

**REVISED FINAL REPORT**

**TASK ORDER 1026**

**ROUND ROBIN STUDY OF *IN-VITRO* BIOACCESSIBILITY ASSAY (IVBA) FOR  
LEAD IN SOIL AND EPA METHOD 3051A FOR LEAD AND ARSENIC:  
FLAT CREEK SOIL REFERENCE MATERIAL**

**Prepared by:**

**QATS Analytical Group**

**Quality Assurance Technical Support Laboratory  
CB&I Federal Services LLC  
2700 Chandler Ave.  
Las Vegas, Nevada 89120**

**February 10, 2017**

**Contract Number: EP-W-16-016**

**Task: 01**

**Statistical Analysis of Round Robin Results**

**Prepared for Sara Duncan**

**Analytical Services Branch  
U.S. Environmental Protection Agency  
Washington, D.C. 20460**

**OFFICE OF SUPERFUND REMEDIATION AND TECHNICAL INNOVATION  
U.S. ENVIRONMENTAL PROTECTION AGENCY  
WASHINGTON, D.C. 20460**

Note: This document is the Revised Final Report for the “Round Robin Study of *In-Vitro* Bioaccessibility Assay (IVBA) for Lead in Soil and EPA Method 3051a for Lead and Arsenic: Flat Creek Soil Reference Material”. This revised report was prepared under Task Order 1026 of the EPA Quality Assurance Technical Support Contract Number EP-W-16-016. This Study was conducted in 2012, and the original final report was submitted to EPA on November 14, 2012. The Study was conducted under Task Order 1026 of the EPA Quality Assurance Technical Support Contract Number EP-W-10-033, managed by Shaw Environmental, Inc. In 2013, Shaw Environmental Inc. was acquired by CB&I Federal Services LLC. All references to “Shaw” or “Shaw Environmental, Inc.” in this revised final report should be considered to be the same as “CB&I Federal Services LLC”. All electronic mail addresses in the report with the domain @shawgrp.com are now @cbifederalservices.com.

Subsequent to the submission of the original report, Syracuse Research Corporation (SRC), under EPA Contract Number EP-W-12-003, conducted an independent statistical analysis of the Study results. Under EPA Contract EP-W-16-016, Task Order 1026, Task 1, CB&I Federal Services LLC has been directed to prepare this revised final report of the 2012 Study to include the independent statistical analysis of the Study results performed by SRC. The results of the SRC independent statistical analysis, in narrative, tabular, and graphic format, have been included in this revised report on Pages 12 through 16 in Section V.E., with the beginning and ending of the SRC independent analysis clearly defined.

## TABLE OF CONTENTS

	PAGE
I. SUMMARY.....	1
II. INTRODUCTION.....	2
III. BACKGROUND.....	2
IV. TECHNICAL APPROACH.....	3
V. RESULTS AND DISCUSSION.....	6
VI. CONCLUSIONS AND RECOMMENDATIONS.....	16
VII. REFERENCES.....	18

## LIST OF APPENDICES

APPENDIX A	FCRM EPA Method 3051A Lead Results and Statistics .....	A-1
APPENDIX B	FCRM EPA Method 3051A Lead Results and t-Test .....	B-1
APPENDIX C	FCRM EPA Method 3051A Arsenic Results and Statistics .....	C-1
APPENDIX D	FCRM Lead IVBA Results and Statistics.....	D-1
APPENDIX E	FCRM Lead IVBA Results and t-Test.....	E-1
APPENDIX F	NIST SRM 2710a Certificate of Analysis .....	F-1
APPENDIX G	Laboratory Submitted Initial Demonstration of Proficiency Forms ....	G-1
APPENDIX H	Laboratory Submitted Round Robin Study Sample Results .....	H-1

## ACRONYMS AND ABBREVIATIONS

<b>ANOVA</b>	Analysis of Variance
<b>dF</b>	Degrees of Freedom
<b>F</b>	Value Calculated
<b>F-C rit</b>	Critical Value
<b>FCRM</b>	Flat Creek Soil Reference Material
<b>HSD</b>	Honestly Significant Difference
<b>ICP-MS</b>	Inductively Coupled Plasma - Mass Spectrometry
<b>ICP-AES</b>	Inductively Coupled Plasma - Atomic Emission Spectrometry
<b>IDP</b>	Initial Demonstration of Proficiency
<b>IVBA</b>	In-Vitro Bioaccessibility Assay
<b>MS</b>	Mean Square
<b>NERL</b>	National Exposure Research Laboratory
<b>NIST</b>	National Institute of Standards and Technology
<b>P-Value</b>	Probability Value
<b>RM</b>	Reference Material
<b>RSD</b>	Relative Standard Deviation
<b>RTP</b>	Research Triangle Park
<b>SAS/STAT</b>	Statistical Analysis Software
<b>SD</b>	Standard Deviation
<b>SEM</b>	Standard Error of the Mean
<b>Shaw</b>	Shaw Environmental, Inc.
<b>Sm</b>	Standard Deviation of the Mean
<b>SOP</b>	Standard Operating Procedure
<b>SOW</b>	Statement of Work
<b>SRC</b>	Syracuse Research Corporation, Inc.
<b>SRM</b>	Standard Reference Material
<b>SS</b>	Sum of Squares
<b>TRW</b>	Technical Review Workgroup
<b>USEPA</b>	United States Environmental Protection Agency
<b>USGS</b>	United States Geological Survey
<b>QATS</b>	Quality Assurance Technical Support Program
<b>QC</b>	Quality Control

**REVISED FINAL REPORT  
TASK ORDER 1026**

**ROUND ROBIN STUDY OF *IN-VITRO* BIOACCESSIBILITY ASSAY (IVBA) FOR  
LEAD IN SOIL AND SOIL-LIKE MATERIALS AND EPA METHOD 3051A FOR LEAD  
AND ARSENIC:  
FLAT CREEK SOIL REFERENCE MATERIAL**

**I. SUMMARY**

The Bioavailability Committee of the USEPA Technical Review Workgroup for Metals and Asbestos (<http://epa.gov/superfund/bioavailability/trw.htm>) conducted a Round Robin Study (herein referred to as Study) of the *In Vitro* Bioaccessibility Assay (IVBA) for Lead<sup>1</sup> and EPA Method 3051A for Lead and Arsenic on the Flat Creek Soil Reference Material (FCRM).

Objectives:

- To derive a mean consensus value for the Lead IVBA for the FCRM, using EPA SOP 9200.2-86
- To report the total amount of lead and arsenic in the FCRM as measured by EPA Method 3051A

This Study included the participation of eight (8) laboratories, each reporting five (5) replicate analysis results for the FCRM Lead IVBA, as well as the Lead and Arsenic EPA Method 3051A digestion, with results totaling three (3) data sets of forty (40) results each. The EPA "Standard Operating Procedure for an *In Vitro* Bioaccessibility Assay" (EPA 9200.2-86) and EPA Method 3051A were provided for the participating laboratories as well as the Scope of Work to be performed. The results were statistically evaluated for IVBA Lead, and total Lead and Arsenic to derive the final consensus values provided in Table 1. No outlying sample results were identified using the Grubb's test either within each laboratory (n=5), or collectively (n=40) for the entire data set.

The associated quality control (QC) sample results provided by the laboratories for the reagent blank, bottle blank, spiked blank, and matrix spike were all within the control limits presented in the standard operating procedure (SOP) EPA 9200.2-86, with the exception of the Lead IVBA control soil. Although there were no outlier sample results identified using the Grubb's test, two (2) of the eight (8) laboratory results for the control soil NIST SRM 2710a exceeded the control limits specified in the EPA SOP 9200.2-86. A statistical comparison (t-test) was performed for the data set for one of the laboratories that had a control soil Lead IVBA result that was outside the control limits specified in EPA SOP 9200.2-86 and could be excluded. The Lead IVBA results for this one (1) laboratory were excluded and the statistical analysis was repeated for the Lead IVBA data set. Similarly, the EPA Method 3051A lead results from another laboratory were statistically evaluated using the t-test, which indicated that these EPA Method 3051A lead results could also be excluded. The revised statistical calculations resulted in lower standard deviation values for the individual and combined Lead IVBA and EPA Method 3051A lead results from the remaining laboratories, thus resulting in narrower Lead IVBA and EPA Method 3051A lead concentration 99-percentile prediction intervals for the new FCRM compared to using the total data set.

---

<sup>1</sup> This method has been incorporated into the SW846 Compendium as Method 1340: <https://www.epa.gov/hw-sw846/sw-846-test-method-1340-vitro-bioaccessibility-assay-lead-soil>.

**Table 1: FCRM Summary of Results (Rounded Values)**

<b>FCRM</b>	<b>Low 99% PI</b>	<b>Mean</b>	<b>High 99% PI</b>	<b>RSD</b>
<b>Lead Method 3051A (mg/Kg)</b>	5490	6440	7400	5.4%
<b>Arsenic Method 3051A (mg/Kg)</b>	550	730	910	8.9%
<b>Lead IVBA Extracted (mg/Kg)</b>	3990	4620	5250	4.9%

## **II. INTRODUCTION**

Utilization of IVBA assays as an estimator of the bioaccessibility and bioavailability of lead in soil has been studied and recognized by the bioavailability scientific community. A comparison of the *in vivo* and *in vitro* assays for lead was conducted in 2007 and the results exhibited a high correlation between the two assays. (2007, EPA OSWER 9285.7-77). The IVBA assay is a viable and less cost prohibitive alternative to an *in vivo* assay (e.g., juvenile swine).

This report provides the Study results for the analysis of the FCRM. The objective of this Study is twofold: (1) derive a mean consensus value for the Lead IVBA for the FCRM, using EPA SOP 9200.2-86, and (2) report the total amount of lead and arsenic in the FCRM as measured by EPA Method 3051A. This report provides the data and statistical analysis of the lead and arsenic results from the Study conducted by the United States Environmental Protection Agency (USEPA), which validates its use as an additional soil reference material for EPA SOP 9200.2-86 and EPA Method 3051A. The FCRM was developed by the United States Geological Survey (USGS) from soil containing high concentrations of metals due to mining activity near an abandoned lead mine in Montana.

The Bioavailability Committee of the USEPA Technical Review Workgroup for Metals and Asbestos initiated the task of verification of the Lead IVBA values for the new FCRM in July, 2011. This Study was coordinated, evaluated, and reported by the USEPA Quality Assurance Technical Support (QATS) Program. The QATS Program was tasked to provide support that included a Study design, the development of the Study instructions in the form of a Statement of Work (SOW), reference material (RM) bottling and shipping, laboratory coordination, statistical analysis of results, and report preparation. Each of the eight (8) laboratories participating in the Study was requested to analyze each of the reference materials in five (5) replicate analyses, along with the EPA SOP 9200.2-86 required QC samples, including blank, matrix spike, and control soil.

## **III. BACKGROUND**

The utilization of IVBA methods as an estimator of the bioavailability of lead in soil matrices has been studied and adopted by the bioavailability community. The IVBA technique is utilized because it is a less expensive method for the estimation of the bioavailability of lead in soil for humans than the previous method of choice, which involved juvenile swine assays. A comparison of the *in vivo* and *in vitro* methods is presented in USEPA OSWER 9285.7-77 (USEPA, 2007). This Study employed two (2) methods: EPA Method 3051A for the determination of total lead and total arsenic, and EPA SOP 9200.2-86 for Lead IVBA determination. These methods are summarized below.

### **III.A. EPA Method 3051A**

EPA Method 3051A was used for the determination of total lead and total arsenic concentrations in the Study samples. Using EPA Method 3051A, solid samples are digested in concentrated nitric acid and concentrated hydrochloric acid using microwave heating with a suitable laboratory

microwave unit. In this Study,  $0.5 \pm 0.001$  grams of sample,  $9 \pm 0.1$  mL of concentrated nitric acid, and  $3 \pm 0.1$  mL of concentrated hydrochloric acid were added to a fluorocarbon polymer microwave vessel. The vessel was then sealed and heated in the microwave unit with power setting(s) that cause the mixture within the vessels to rise to a temperature of  $175^{\circ} \text{C} \pm 5^{\circ} \text{C}$  in approximately  $5.5 \pm 0.25$  minutes, and remain at  $175^{\circ} \text{C} \pm 5^{\circ} \text{C}$  for 4.5 minutes, or for the remainder of the 10 minute digestion period. After cooling, the vessel contents are either filtered, centrifuged, or allowed to settle, and then diluted to a suitable volume and analyzed using either EPA SW-846 Method 6010C (ICP-AES) or EPA SW-846 Method 6020A (ICP-MS).

### III.B. EPA SOP 9200.2-86

EPA SOP 9200.2-86 was the method used for the determination of Lead IVBA results for the Study samples. Throughout this report, the term "Lead IVBA" is used synonymously with EPA SOP 9200.2-86, unless the SOP is specifically referenced. The IVBA method is performed by first retrieving the soil to be assessed for *in vitro* bioaccessibility assay, drying the soil at less than  $40^{\circ} \text{C}$ , and passing the dried material through a sieve to obtain the soil particles that are less than  $250 \mu\text{m}^2$ . One (1) gram of the soil is placed in a plastic bottle, and 100 mL of 0.4 M glycine, at a pH of 1.5, is added. The sample bottle(s), and associated quality control sample bottles, are then placed on a rotary extractor ( $30 \pm 2$  RPM) for one (1) hour while being heated at a constant temperature of  $37^{\circ}\text{C} \pm 2^{\circ}\text{C}$ . The heating of the bottles and rotary extraction apparatus is accomplished by immersion in a temperature controlled water bath (aquarium style), or alternatively, the apparatus can be heated by the flow of temperature controlled air (incubator style). After the prescribed extraction period, the bottles are removed from the extraction apparatus and the supernatant removed using an in-line filter and a 20 mL syringe. The filtered supernatant is then analyzed for lead (or other analytes) by ICP-AES or ICP-MS using the analytical methods cited above.

The Lead IVBA value for the FCRM is expressed as the ratio of the Lead IVBA result divided by EPA Method 3051A lead result, multiplied by 100.

$$IVBA (\%) = \frac{EPA\ SOP\ 9200.2 - 86\ Result}{Method\ 3051A\ Result} \times 100$$

EPA Method 3051A and EPA SOP 9200.2-86 can be accessed using the following USEPA website hyperlinks:

<https://www.epa.gov/hw-sw846/sw-846-test-method-3051a-microwave-assisted-acid-digestion-sediments-sludges-soils-and-oils>

<https://nepis.epa.gov/Exe/ZyPDF.cgi/P100GESL.PDF?Dockkey=P100GESL.PDF>

## IV. TECHNICAL APPROACH

Shaw's QATS Program support included the following subtasks:

- Contacting candidate laboratories with previous IVBA experience;
- Requesting laboratories to complete an Initial Demonstration of Proficiency (IDP) form, if they had not done so in a previous Lead IVBA Round Robin Study;
- Bioavailability Committee of the USEPA Technical Review Workgroup for Metals and Asbestos review of the completed IDP forms and selection of laboratories to participate

---

<sup>2</sup> After this round robin was completed, the recommended sieve size for this method was revised to  $<150 \mu\text{m}$ .

in the Study;

- Study Design;
- Development of a Statement of Work (SOW), including IVBA data reporting forms;
- Shipment of the IVBA samples and associated QC samples; and
- Statistical analysis of the Study results and report preparation.

#### IV.A. Contacting Laboratories, IDP Form, and Participating Laboratory Selection

To identify qualified candidates to participate in the Study, Shaw first contacted laboratories with previous IVBA experience. Most of the laboratories in this Study were participants in a previous Lead IVBA Round Robin Study conducted by the Bioavailability Committee of the USEPA Technical Review Workgroup for Metals and Asbestos and coordinated by Shaw, which was completed in 2011. The laboratories were asked to complete an IDP form to determine their level of experience with the IVBA procedures. The information requested on the IDP form included the total number of IVBA analyses performed by the laboratory, as well as the QC sample results for the most recent ten (10) batches of IVBA analyses conducted at their facility. Only those laboratories that had not been participants in the previous Study were asked to complete the IDP form. From previous participation and the IDP form response, the Bioavailability Committee of the USEPA Technical Review Workgroup for Metals and Asbestos selected a total of eight (8) laboratories for participation in the Study, which are presented in Table 2. In order to maintain the anonymity of the Study participants, the IDP forms provided by the laboratories are presented in Appendix F in redacted format, with an alphanumeric letter used as an identifier, in an order inconsistent with the order presented in Table 2. The IDP forms, without redaction, are available from USEPA HQ Co-Chair for the Technical Review Workgroup (TRW), <http://epa.gov/superfund/bioavailability/trw.htm>.

**Table 2. Laboratories Selected to Participate in the Study**

Laboratory		Address	Contact Name and e-mail Address
1	ACZ Laboratories	2773 Downhill Drive Steamboat Springs, CO 80487	Mr. Tim VanWyngarden (timv@acz.com) Ms. Sue Webber (suew@acz.com)
2	USEPA National Exposure Research Laboratory (NERL) Research Triangle Park (RTP)	109 T.W. Alexander Dr. Research Triangle Park, NC 27709	Dr. Karen Bradham (bradham.karen@epa.gov)
3	Ohio State University	School of Environment and Natural Resources 2021 Coffey Rd. 410 C Kottman Hall Columbus, OH 43210-1043	Dr. Nicholas Basta (basta.4@osu.edu)
4	PRIMA Environmental	5070 Robert J Mathews Pkwy, Suite 300 El Dorado Hills, CA 95762	Dr. Cindy Schreier (cschreier@primaenvironmental.com)
5	USEPA Region 7 Laboratory	300 Minnesota Avenue Kansas City, KS 66101	Michael Davis (davis.michael@epa.gov)



6	USEPA Region 9 Laboratory	1337 South 46th Street, Bldg 201 Richmond, CA 94804	Richard Bauer (bauer.richard@epa.gov)
7	Royal Roads University (Canada)	Royal Roads University 2005 Sooke Road Victoria, BC, Canada V9B 5Y2	Dr. Matt Dodd (Matt.Dodd@RoyalRoads.ca)
8	University of Colorado	Benson Earth Science 2200 Colorado Avenue Boulder, CO 80309	Dr. John Drexler (Drexlerj@Colorado.edu)

## IV.B. Study Design

### IV.B.1. FCRM and Number of Replicates

The FCRM used in this Study was sent to the QATS Laboratory for sub-aliquoting and shipment on February 24, 2012 by USEPA National Exposure Research Laboratory (NERL) personnel, who had previously received the material from the USGS Associate Project Chief. The FCRM was provided in a 500 mL glass bottle, and sufficient FCRM material was mixed before sub-aliquots were bottled for Study sample analysis. The standard reference material (SRM) NIST SRM 2710a used as a control soil in this Study was provided by the NIST Analytical Chemistry Division from a previous Study conducted at the QATS Laboratory in 2009. The QATS Laboratory was provided with 50 grams of NIST SRM 2710a, and a sufficient amount of the material for the Study was combined and mixed before sub-aliquots were bottled for distribution to the laboratories.

The moisture content of the FCRM was <0.5%, and was determined by heating a 5 gram sample in an oven at 105° C for twelve (12) hours. The NIST SRM 2710a moisture content is approximately 2%, and the particle size is <74 µm, as reported on the NIST SRM 2710a Certificate of Analysis.

The Certificate of Analysis for the NIST SRM 2710a is presented in Appendix F. Table 3 provides the lead and arsenic concentration, particle size, and moisture content for this NIST SRM 2710a, derived from the Certificates of Analysis.

**Table 3. NIST SRM 2710a Certificate of Analysis Parameters**

Element	Total Concentration (mg/Kg)	Leachable Concentration (mg/Kg)	Particle Size	Moisture Content
Pb	5520	5100	<74 µm	~2%
As	1540	1400		

The Bioavailability Committee of the USEPA Technical Review Workgroup for Metals and Asbestos determined that five (5) replicate analyses of the FCRM would be conducted by each laboratory participating in the Study. Five (5) replicate analyses were chosen to ensure that a sufficient number of results were available for establishing a statistically sound Lead IVBA mean value and control limits for the new FCRM.

### IV.B.2. QC Samples

In this Study, the laboratories were instructed to analyze the samples in strict accordance with the EPA SOP 9200.2-86 including all of the associated quality control samples, with noted exceptions. Table 4 below provides the EPA SOP 9200.2-86 required QC samples and associated control limits used in this Study.

**Table 4. EPA SOP 9200.2-86 Required QC Samples and Control Limits**

<b>QC Sample</b>	<b>Control Limits</b>
Reagent Blank	<25 µg/L Lead
Bottle Blank	<50 µg/L Lead
Blank Spike (10 mg/L)	85% -115% Recovery
Matrix Spike (10 mg/L)	75% -125% Recovery
Duplicate Sample	± 20% RPD
Control Soil (NIST SRM 2710a)	IVBA Mean = 67.5% Acceptable Range 60.7% - 74.2%

NIST SRM 2710a was used as the control soil for both the Lead IVBA and EPA Method 3051A portions of the Study, followed by analysis. Both the lead and arsenic mean values and range appear in the Addendum to the NIST SRM 2710a Certificate of Analysis titled "Leachable Concentrations Determined Using EPA Methods 200.7 and 3050B." Five (5) replicate aliquots of the FCRM were subjected to the Lead IVBA procedure; therefore, there was no additional duplicate sample analysis requirement in this Study as a measure of analytical precision.

The laboratories were instructed to perform the analysis of one set of QC samples with each RM batch for both the Lead IVBA and the EPA Method 3051A methods.

#### **IV.C. Statement of Work for the Study**

An SOW was developed by QATS personnel and the Bioavailability Committee of the USEPA Technical Review Workgroup for Metals and Asbestos which provided instructions to the participating laboratories on the analysis and reporting of the Study samples. The SOW provided a list of samples for each Lead IVBA batch and a recommended sequence of instrumental analysis of the Lead IVBA samples. The SOW also provided a list of the required associated QC sample analysis and QC sample control limits derived from the EPA SOP 9200.2-86.

#### **IV.D. Shipment of the Study Samples and Associated QC Samples**

The Study samples were shipped to the eight (8) participating laboratories in April, 2012. The laboratories were provided a 30 day turnaround time for submitting the sample results. The Study sample shipments also included hardcopies and CDs of the SOW and the EPA SOP 9200.2-86.

#### **IV.E. Statistical Analysis of the Study Results**

Conventional statistical analysis techniques were used to analyze the data collected from the Study. The statistical analyses were performed in Microsoft® Excel, using Analysis of Variance (ANOVA) and t-test data analysis tools provided by the Excel Analysis Tool Pac add-in package.

The statistical tool ANOVA, single factor (e.g., lead), was used to discern the intralaboratory versus the interlaboratory sources of variance of each FCRM data set derived from the Study. The statistical t-test was used to analyze the data from the different laboratories to determine, for example, if one set of data is statistically different than the others. Specifically, the t-test employed was the two (2) sample, assuming equal variances t-test.

The QC samples, including the reagent blank, bottle blank, spiked blank, matrix spike, and NIST SRM 2710a, were also processed with the Lead IVBA / EPA Method 3051A digested FCRM samples. The results were evaluated to determine if there were any anomalous data submitted by a participating laboratory that should be excluded from the composite results in the course of setting the FCRM statistical values and control limits.

## V. RESULTS AND DISCUSSION

### V.A. Initial Demonstration of Proficiency

The IDP forms provided by the laboratories selected for the Study are presented in Appendix G. As discussed in a previous section, these forms have been redacted to preserve anonymity. The original unredacted forms are available from the USEPA HQ Co-Chair for the TRW. Out of the ten (10) candidate laboratories submitting IDP forms, eight (8) laboratories were selected to be participants in the Study.

### V.B. Study Results

Each of the eight (8) laboratories participating in the Study analyzed the FCRM using five (5) replicate aliquots, providing a total of 40 results for the Lead IVBA procedure, and 40 lead and arsenic results for the EPA Method 3051A procedure. The SOW provided to the laboratories contained several tables that allowed laboratory reporting of the Study sample analysis results using Microsoft® Word. The participating laboratories were asked to submit the results to the QATS Laboratory via electronic mail, and provide hard copies of the results that could not be converted to electronic files. The results provided by the laboratories in the SOW tables are presented in Appendix H in redacted form. The original unredacted SOW forms completed by the laboratories are available from the USEPA HQ Co-Chair for the TRW.

### V.C. FCRM Results and Statistical Analysis

#### V.C.1. Lead Results, EPA Method 3051A

Results of EPA Method 3051A for lead for the FCRM are presented in Appendix A. Table A-1 presents the EPA Method 3051A lead results for the FCRM. The mean lead result from all eight (8) laboratories (n=40) is 6,634 mg/Kg, with a pooled RSD value of 9.1%. The calculated lead 99 percentile prediction interval, based on the EPA Method 3051A results (n=40) alone, is  $\pm 25.0\%$ . As shown in Table A-1, the calculated percent standard deviation of the mean is 1.4%, and the calculated 99 percentile confidence interval of the mean is  $\pm 3.9\%$ . Note that the Laboratory D EPA Method 3051A mean lead result of 7,963 mg/Kg was higher than the results from the other laboratories, and 20% higher than the EPA Method 3051A mean result of 6,634 mg/Kg.

The formulas used for the prediction interval and confidence interval of the mean are provided below in Exhibit 1.

**Exhibit 1:**

$$\text{Prediction Interval: } \bar{x} \pm \left( sd * t \left( \sqrt{1 + \frac{1}{n}} \right) \right)$$

$$\text{Confidence Interval: } \bar{x} \pm (sm * t) \quad \text{where } sm = \frac{sd}{\sqrt{n}}$$

Where:

sd = standard deviation (n-1)

t = Student's t; for n = 40, df=39, t = 2.708, for 99 percentile

sm = standard deviation of the mean

The mean Pb IVBA value is the mean Pb extraction result / the mean Pb EPA Method 3051A digestion result \* 100.

The pooled standard deviation resulting from division is based on the square root of the sum of the squares formula for two independent variables with unequal means formula. Please note that the sd are normalized to percentiles before squaring.

$$sd \text{ Pb IVBAratio} = \left( Pb \text{ IVBAratio} * \left( \sqrt{((sd / \text{mean Extraction})^2 + (sd / \text{mean Digestion})^2)} \right) \right)$$

**Note:** The square root of the sum of variances squared method was used as an estimator of the combined variance for the final lead IVBA result, as the expected means and variances of the IVBA extraction and digestion results are not expected to be equal. The IVBA extraction and the digestion results are not subsets of the same population, and therefore their respective variances are additive, even during the division operation.

Table A-2 presents the EPA Method 3051A lead results for the associated QC samples that were determined with the FCRM. These results include the blank spike recovery, matrix spike recovery, and the NIST SRM 2710a results and percent recovery. All results are within the control limits presented in the EPA SOP 9200.2-86 and in Table 4 above, with the exception of the EPA Method 3051A NIST SRM 2710a lead result of 4,537 mg/Kg for Laboratory E, which is slightly below the lower limit of 4,700 mg/Kg. Although the Laboratory E, EPA Method 3051A NIST SRM 2710a lead result is slightly below the lower control limit for lead based on the Addendum to the NIST SRM 2710a Certificate of Analysis, this did not translate into lower results for the FCRM when compared to the other laboratory results. The EPA Method 3051A NIST SRM 2710a mean lead result for all eight (8) laboratories is 100.8% of the value presented in the Addendum to the NIST SRM 2710a Certificate of Analysis, which indicates good overall accuracy. The Laboratory D NIST SRM 2710a matrix spike recovery of 57% is outside the 75% to 125% matrix spike recovery range; however, because the spiking ratio was less than 1:4 spike to sample concentration, this spike result is not a reliable predictor of accuracy.

Table A-3 presents the ANOVA for the FCRM EPA Method 3051A lead results. For each set of laboratory data, Table A-3 presents the number of sample replicates (n), as well as the sum, mean, and variance (square of the data set standard deviation) values. The table also provides the various statistical calculation values that are used by the ANOVA algorithm to test the variance of all of the results for both within a laboratory, and between laboratories. These calculation results include: sum of squares (SS), degrees of freedom (df), mean square (MS), value calculated (F), critical value of (F-Crit), and probability value (P-value).

The results of the ANOVA assessment which are presented in Table A-3 indicate that the intralaboratory variance is low compared to interlaboratory variance. This is reflected by the large MS value for the interlaboratory group results (1,807,266) compared to the lower intralaboratory group results MS value (49,325). The variance uses the null hypothesis that the data sets provided by the laboratories represent the same samples, analyzed by the same method. The ANOVA

assessment allows the user to select the probability of error of falsely rejecting the hypothesis that all results are from the same population (same samples and method). The error significance level is typically set at 95%, which translates to a 5% chance of wrongly rejecting the hypothesis. The data comparison performed by the algorithm is referred to as a two-tail test, which means that both the upper and the lower ends of data distribution are tested. The ANOVA algorithm calculates (or selects from an algorithm table) the f-critical value, based on the assumption of normal distributions of the intralaboratory results and the composite results. If the calculated f-value, which is based on the ratio of variances displayed by the between laboratory results to the variance of individual laboratory results, is greater than the f-critical value, then the null hypothesis is rejected, which is the case for the lead extraction data sets. The ANOVA results presented in Table A-3 indicate that the variance in interlaboratory data is large relative to the intralaboratory data variances; therefore the null hypothesis is rejected with a high degree of confidence (low P-value). The rejection of the null hypothesis could indicate: 1) different methods were used in the analysis, 2) different samples were being analyzed, or 3) the intralaboratory variance is small compared to what might be expected. The latter choice must be accepted as correct, considering the RSDs for the FCRM for the intralaboratory (n=5) results all quite low (less than 8% RSD for three (3) data sets and less than 3% RSD for the remaining five (5) sets of results).

Appendix B provides the statistical t-test comparison of the Laboratory D EPA Method 3051A lead results. Table B-1 presents the t-test statistical comparison for the FCRM EPA Method 3051A lead results from Laboratory D and the results from the other seven (7) laboratories. This t-test was performed because the EPA Method 3051A lead results from Laboratory D as shown in Table A-1 were higher than the other laboratory results.

The t-test was employed to evaluate if there was a statistical difference between the results from Laboratory D versus the other reported results. The t-test function in Microsoft® Excel was used, which is the 2-sample (assuming equal variances, alpha 0.01, 99-percentile) t-test. The t-test results presented in Table B-1 shows there is a significant difference between the data from Laboratory D compared to the results derived from the other laboratories collectively, as indicated by a P ( $T \leq t$ ) value that is less than 0.01 for the t-tests performed on the data set. A t-Stat value that is greater than the t-critical value also indicates a significant difference between the Laboratory D data and the remaining data sets. The t-test comparison results, which are presented in Table B-1, indicate that the extraction results for Laboratory D can reasonably be excluded with a less than 1% chance of being incorrect. The EPA Method 3051A lead results for this laboratory were omitted and the statistical analysis was repeated for the EPA Method 3051A data set.

Table B-2 presents the revised statistical analysis of the EPA Method 3051A lead results for the FCRM, excluding the results from Laboratory D. The mean of the pooled EPA Method 3051A lead results (n=35) from the FCRM is 6,444 mg/Kg. The calculated pooled standard deviation of the FCRM EPA Method 3051A lead results is 345 mg/Kg, which provides a pooled RSD of 5.4%. The calculated lead 99 percentile prediction interval, based on the EPA Method 3051A lead results (n=35) alone, is  $\pm 14.8\%$  of the mean value of 6,444 mg/Kg. The calculated percent standard deviation of the mean for the FCRM EPA Method 3051A lead results is 0.91%. The calculated 99 percentile confidence interval of the mean EPA Method 3051A lead results for the FCRM is 6,444  $\pm 2.5\%$ .

### **V.C.2. Arsenic Results, Method 3051A**

Appendix C presents the FCRM EPA Method 3051A arsenic results. Table C-1 presents the EPA Method 3051A arsenic results, prediction intervals, and confidence intervals for the FCRM. The mean arsenic result is 728 mg/Kg, with a standard deviation of 65 mg/Kg and RSD of 8.9%. The calculated arsenic 99 percentile prediction interval, based on the EPA Method 3051A arsenic results (n=40) alone, is 728 mg/Kg  $\pm 24.2\%$ . The calculated 99 percentile confidence interval of the EPA Method 3051A mean arsenic result for the FCRM is 728 mg/Kg  $\pm 3.8\%$ .

Table C-2 presents results for the associated arsenic QC samples that were also determined by EPA Method 3051A along with the FCRM. The table includes the blank spike recovery, matrix spike recovery, and the NIST SRM 2710a results and percent recovery. Using the NIST SRM 2710a Certificate of Analysis as a guideline, the NIST SRM 2710a arsenic results are within the range presented in the Certificate of Analysis, with the exception of the results from Laboratories B and F. The arsenic results from Laboratories B and F, at 1,650 mg/Kg and 1,684 mg/Kg, respectively, slightly exceed the 1,600 mg/Kg upper range for arsenic listed in the Certificate of Analysis. The NIST SRM 2710a mean arsenic result for all eight (8) laboratories is 1,592 mg/Kg. This value is 113.7% of the NIST SRM 2710a strong leach value of 1,400 mg/Kg presented in the Addendum to the NIST SRM 2710a Certificate of Analysis, and indicates a high bias in the recovery of arsenic from the EPA Method 3051A digestion relative to the CLP digestion procedures referenced in the Certificate of Analysis. In comparison, the CLP digestions are usually open beaker or block digestions and may result in incomplete digestion, or possible increased losses of arsenic acid vapor during digestion.

Table C-3 presents the ANOVA for the FCRM EPA Method 3051A arsenic results. As with the ANOVA of the FCRM EPA Method 3051A lead results, the ANOVA of the FCRM EPA Method 3051A arsenic results show the intralaboratory variance to be low compared to interlaboratory variance. This is indicated by the large MS value of 20,732 for the interlaboratory group results compared to the lower intralaboratory group results MS value of 585. The RSD values for the FCRM EPA Method 3051A intralaboratory results (n=5) are all 6% or less.

### **V.C.3. Lead IVBA Results (EPA SOP 9200.2-86)**

Appendix D presents the FCRM Lead IVBA results. Table D-1 presents the Study Lead IVBA results for the FCRM along with the mean, standard deviation (n-1 weighting), and RSD values for each sample set. Please note that the values presented in these tables are not rounded. The pertinent rounded values are presented in the Conclusions and Recommendations section of this report. Table D-1 also presents the 99 percentile prediction interval for the Lead IVBA result, in mg/Kg. This 99 percentile prediction interval for the Lead IVBA result was converted to the Lead IVBA prediction interval by dividing the statistically combined Lead IVBA results with the statistically combined EPA Method 3051A lead results. Table D-1 also provides the confidence interval of the mean for the FCRM Lead IVBA value.

Table D-1 presents the mean concentration (n=40) of the FCRM Lead IVBA at 4,700 mg/Kg. This is 70.8% of the EPA Method 3051A mean lead concentration of 6,634 mg/Kg, which is presented in Table A-1, and represents a Lead IVBA value of 70.8%. The calculated pooled RSD value of the FCRM Lead IVBA results is 6.5%. The calculated lead 99 percentile prediction interval based on the Lead IVBA results alone (n=40) is  $\pm 17.7\%$ . The calculated lead 99 percentile prediction interval for the Lead IVBA result, which includes the variance of the EPA Method 3051A results (n=80), is significantly higher at  $\pm 29.7\%$ . The calculated percent standard deviation of the mean for the FCRM is 1.24%. The calculated 99 percentile confidence interval of the Lead IVBA mean result for the FCRM is  $70.8 \pm 3.3\%$ . The Laboratory B results in Table D-1 were observed to be higher than the results from the other laboratories.

Table D-2 presents the Lead IVBA results for the associated QC samples that were processed with the FCRM, as well as the EPA Method 3051A lead QC results. These include results for the reagent blank, bottle blank, blank spike, matrix spike, and the NIST SRM 2710a. All results are within the control limits presented in the EPA SOP 9200.2-86 and in Table 4 above, with the exception of the Lead IVBA NIST SRM 2710a results from laboratories B and C and the reagent blank result for laboratory C.

In Table D-2, row 1, Laboratory C reported a reagent blank result of <40 ug/L, which is greater

than the EPA SOP 9200.2-86 required detection limit of 25 ug/L. The laboratory was contacted after a review of the laboratory's reagent blank results, and confirmed the reported <40 ug/L blank result was correct. However, because the actual analytical sample results are approximately 100 times greater in concentration, this elevated blank result does not impact the Study results.

Row 6 of Table D-2 presents the NIST SRM 2710a Lead IVBA percent recovery based on the Lead IVBA mean recovery of 3,440 mg/Kg from a previous Study. All percent recovery results are within  $\pm 20\%$  of the mean value, with the Laboratory B recovery the highest at 116.3%.

Row 8 of Table D-2 presents the NIST SRM 2710a EPA Method 3051A percent recovery for lead based on the NIST SRM 2710a Certificate of Analysis mean result for the strong leach acid digestion (EPA Method 3050B) of 5,100 mg/Kg, with the acceptance range of (4,700 - 5,800 mg/Kg). All of the lead digestion result recoveries are  $100 \pm 20\%$ ; however, compared with the Certificate of Analysis acceptance range (4,700 – 5,800 mg/Kg), the EPA Method 3051A result for Laboratory E was slightly below the lower limit at 4,537 mg/Kg. However, because the FCRM Lead IVBA results for Laboratory E were not low biased, the results were retained.

Row 9 of Table D-2 presents the Lead IVBA values for the NIST SRM 2710a derived from both the Lead IVBA and EPA Method 3051A results from this Study. The results are within the previously established EPA SOP 9200.2-86 control limits of 60.7% to 74.2% with the exception of the results from Laboratories B and C, which exceeded the 99 percentile control limits at 78.4% and 77.3%, respectively. The FCRM Lead IVBA results from these two (2) laboratories seem to correlate with these high NIST SRM 2710a Lead IVBA results.

Table D-3 presents the statistical summary for the ANOVA for the FCRM Lead IVBA results. The results of the ANOVA assessment, which are presented in Table D-3, indicate that the intralaboratory variance is low compared to interlaboratory variance. This is indicated by the large MS value of 485,588 for the interlaboratory group compared to the substantially lower MS value of 6,469 for the intralaboratory group. The ANOVA results presented in Table D-3 indicate that the variance in interlaboratory data is large relative to the intralaboratory data variances; therefore the null hypothesis is rejected with a high degree of confidence (low P-value). The RSD values for the FCRM for the intralaboratory (n=5) results are all quite low (less than 4% for one (1) of the sets, and less than 2% for the remaining seven (7) sets of results).

Because of the higher than acceptable Lead IVBA results for the NIST SRM 2710a from Laboratories B and C presented in row 9 of Table D-2, a statistical comparison (t-test) was performed between the FCRM data set for Laboratory B (the highest results) and the remaining FCRM data. The t-test was employed to evaluate if there was a statistical difference between the results from Laboratory B versus the results from the remaining seven (7) laboratories. The Excel t-test output for this exercise is presented in Appendix E. The t-test results presented in Table E-1 shows there is a significant difference between the data from Laboratory B compared to the results derived from the other laboratories collectively, as indicated by a P ( $T \leq t$ ) value that is less than 0.01 for the t-tests performed on the data set. The t-test comparison results, which are presented in Table E-1, indicate that the Lead IVBA results for Laboratory B can reasonably be excluded with a less than 1% chance of being incorrect. The Lead IVBA extraction results for this one (1) laboratory were omitted and the statistical analysis was repeated for the remaining Lead IVBA data set.

Table E-2 presents the revised statistical analysis for the FCRM Lead IVBA results, excluding the results from Laboratory B. The FCRM Lead IVBA mean value minus the results from Laboratory B (n=35) is 4,619 mg/Kg. The Lead IVBA value is 69.6% of the EPA Method 3051A mean lead value of 6,634 mg/Kg, presented in Table E-2, and represents a Lead IVBA value of 69.6. The calculated pooled RSD of the FCRM Lead IVBA results is 4.9%. The calculated lead 99 percentile prediction interval based on the Lead IVBA results (n=35) alone, is  $\pm 13.6\%$ . The calculated 99 percentile prediction interval for the Lead IVBA value, which includes the variance of the EPA Method 3051A results (n=75), is  $\pm 27.5\%$ . The calculated percent standard deviation of the mean for the FCRM Lead IVBA results is 1.2%. The calculated 99 percentile confidence interval of the mean Lead IVBA result for the FCRM is  $69.6 \pm 3.2\%$ . Overall, the statistical results for the FCRM, excluding the Laboratory B data set, exhibit slightly increased precision when compared to the full data set.

Table D-4 presents the resulting calculated 99 percentile prediction interval for the Lead IVBA, which includes the variance of the EPA Method 3051A results, but has excluded both the Laboratory B Lead IVBA results and Laboratory D EPA Method 3051A lead results. The Lead IVBA value (n=70) is 71.7%, the SD is 5.2, and the RSD is 7.3%. The calculated 99 percentile prediction interval for the Lead IVBA, which includes the variance for the EPA Method 3051A results (n=70) is  $71.7\% \pm 19.4\%$ . The calculated percent standard deviation of the mean for the FCRM Lead IVBA result is 0.87%. The calculated 99 percentile confidence interval of the Lead IVBA mean result for the FCRM is  $71.7\% \pm 2.3\%$ . Overall, the statistical results for the FCRM, excluding the Laboratory B Lead IVBA data set and the Laboratory D EPA Method 3051A lead data set, exhibit increased precision as compared to the full data set.

#### **V.D. Summary of FCRM Study Results and Prediction Intervals**

Table 5 provides the EPA Method 3051A lead and arsenic results, statistics, and 99 percentile prediction intervals for the FCRM. Table 6 provides the Lead IVBA results, statistics, and 99 percentile prediction intervals for the FCRM.



**Table 5. FCRM EPA Method 3051A Results and Statistics**

Analyte	Low 99% PI	Mean	High 99% PI	RSD	N
Pb	5490	6440	7400	5.4%	35
As	550	730	910	8.9%	40

**Table 6. FCRM Lead IVBA Results and Statistics**

Analyte	Mean Result Lead IVBA (%)	Lead IVBA (%) (± 99% PI)	RSD*	Mean Result Lead IVBA (mg/Kg)	Lead IVBA (mg/Kg) (± 99% PI)	RSD*	N
Pb	71.7	(57.8 – 85.6)	7.3%	4619	(3991 – 5247)	4.9%	35

\* RSD was derived from the replicate Lead IVBA results.

**V.E. Independent Statistical Analysis of the Round Robin Study Results by Syracuse Research Corporation (SRC) and Comparison to Shaw’s Standard Statistical Analysis Results**

SRC, Inc. under contract EP-W-12-003 analyzed the Round Robin Study data using Tukey’s Studentized Range (also known as the Honestly Significant Difference [HSD] test). This test evaluates whether the data from each laboratory are significantly different from the others while controlling the type 1 error rate (at  $\alpha = 0.05$ ) when multiple statistical comparisons are performed. When data from one or more laboratories were identified as different from the others, these datasets were further evaluated visually to determine if they should be excluded from the final dataset used to calculate prediction intervals (PIs) and confidence intervals (CIs). If the Tukey’s HSD test or visual examination of the data did not identify any datasets that differed significantly from the others, or if the test indicated many differences among the datasets with no clear grouping, then all laboratory results were included in the final dataset.

The final dataset for each measurement endpoint included all laboratories that were not excluded for QC issues and were not identified as significantly different from the other laboratories by Tukey’s HSD or visual inspection of the data. These data were used to calculate the PIs and CIs for the measurement endpoint. The PI for a Reference Material (RM) refers to a specific measurement and is used to determine if a laboratory result is acceptable, while the CI is an estimated range of values that is likely (with probability of  $\alpha$ ) to include the mean of a population. The formulas used for the prediction and CI are provided in Exhibit 2.

**Exhibit 2.**

99% Prediction Interval:  $\bar{x} \pm \left( sd * t \left( \sqrt{1 + \frac{1}{n}} \right) \right)$

99% Confidence Interval:  $x \pm (sem * t)$  where  $sem = \frac{sd}{\sqrt{n}}$

Where:

- SD = standard deviation
- t = Student’s critical t value for  $\alpha = 0.01$  (two-tailed)
- n = number of samples
- SEM = standard error of the mean

The final datasets for lead by EPA method 3051A and by IVBA Method SOP 9200.2-86 were used to estimate the IVBA as a percentage of the total lead (i.e., IVBA by Method EPA SOP 9200.2-86 / lead by Method 3051A). Fieller confidence intervals for the ratio of bi-variate normal random variables were calculated (Fieller, 1954; Dilba et al., 2006; Tamhane and Logan, 2004) using the T-

Test procedure in SAS (Exhibit 3; SAS/STAT software, Version 9.3 of the SAS System for Windows).

### Exhibit 3.

99% Confidence Interval (unequal variance):  $-\frac{bu}{a_u} + \frac{\sqrt{(b_u^2 - b_u c_u)}}{a_u}, -\frac{bu}{a_u} - \frac{\sqrt{(b_u^2 - b_u c_u)}}{a_u}$

Where:

$$a_u = \frac{sp t_{1-\frac{\alpha}{2}, df_u}^2}{n_2} - \bar{x}_2^2$$

$$b_u = \bar{x}_1 * \bar{x}_2$$

$$c_u = \frac{sp t_{1-\frac{\alpha}{2}, df_u}^2}{n_1} - \bar{x}_1^2$$

$$df_u = \left( \frac{s_1^2}{n_1} + \frac{s_2^2 \bar{x}_2^2}{n_2} \right)^2 / \left( \frac{s_1^4}{n_1^2(n_1-1)} + \frac{s_2^4 \bar{x}_2^4}{n_2^2(n_2-1)} \right) \text{ (Satterthwaite approximate degrees of freedom; Tamhane and Logan, 2004)}$$

$x_1$  = average of IVBA lead (mg/kg)

$x_2$  = average of lead by 3051A

$s_1$  = standard deviation of IVBA lead (mg/kg)

$s_2$  = standard deviation of lead by 3051A

Prediction intervals account for the variability among individual measurements as well as the uncertainty in the estimate of mean. The prediction interval was estimated by extending the Fieller confidence limits to account for the estimated variability of the ratio (Exhibit 4).

### Exhibit 4.

99% Prediction Interval (unequal variance):  $LL - s_r * t_{1-\frac{\alpha}{2}, df_u} UL + s_r * t_{1-\frac{\alpha}{2}, df_u}$

Where:

$LL, UL$  = lower limit, upper limit of the 99% Fieller type CI (unequal variance) for the ratio (Exhibit 2)

$$s_r = \frac{\bar{x}}{y} \sqrt{\left( \frac{s_1}{x_1} \right)^2 + \left( \frac{s_2}{x_2} \right)^2} \text{ (first order Taylor series approximation; Stuart & Ord, 1986)}$$

$$df_u = \left( \frac{s_1^2}{n_1} + \frac{s_2^2 \bar{x}_2^2}{n_2} \right)^2 / \left( \frac{s_1^4}{n_1^2(n_1-1)} + \frac{s_2^4 \bar{x}_2^4}{n_2^2(n_2-1)} \right) \text{ (Welch-Satterthwaite degrees of freedom; Tamhane and Logan, 2004)}$$

$x_1$  = average of IVBA (mg/kg)

$x_2$  = average of lead by 3051A

$s_1$  = standard deviation of IVBA (mg/kg)

$s_2$  = standard deviation of lead by 3051A

The SRC estimates based on the methods described above are presented in Table 7 below. The results of Pb 3051A and As 3051A are identical to what was determined using the standard statistical tests, compare to results in Tables B-2 and C-1, respectively. The SRC estimates of Pb IVBA (mg/kg) differ from the standard statistical results (compare Table 7 with Table E-2) because different laboratories were selected for inclusion in the estimate of Pb IVBA (mg/kg).

For the Pb 3051A data, SRC found the results of the ANOVA assessment indicated that data from at least one laboratory were significantly ( $p < 0.05$ ) different from the other data. Analysis of the data using Tukey's HSD test showed that the data from Laboratory D were different from all of the remaining laboratories; this difference is also evident in Figure 1 (below). As data from Laboratory D were significantly different from all other laboratories, these data were omitted from the final dataset. Laboratory D was also eliminated from the estimate of Pb 3051A in the analysis using the standard statistical approach (see Table B-2).

For the As 3051A data, SRC found the results of the ANOVA assessment indicated a significant ( $p < 0.05$ ) difference among the laboratories. Analysis of the data using Tukey's Studentized Range Test indicated many differences among the datasets with no clear majority grouping (see Figure 2 below). Therefore, all laboratories were included in the final dataset. All laboratories were also included in the estimate of As 3051A in the standard statistical analysis (see Table C-1).

For the Pb IVBA data, SRC removed the results from laboratories B and C because these laboratories exceeded the previously established EPA SOP 9200.2-86 control limits of 60.7% to 74.2% on the NIST SRM 2710a control soil. In addition, the results of the ANOVA assessment indicated a significant ( $p < 0.05$ ) difference among the laboratories. While Tukey's HSD found the data from laboratory E to be significantly different from all of the remaining laboratories, visual inspection of Figure 3 (below) does not indicate the mean IVBA from laboratory E is substantially different from the mean IVBAs from the other laboratories; therefore, the data from laboratory E were retained in the final dataset.

Since SRC eliminated both laboratories B and C data for the Pb IVBA (mg/kg) results but the Shaw analysis only eliminated laboratory B (see section 'V.C.3 Lead IVBA Results (EPA SOP 9200.2-86)' for a discussion on why laboratory B data was eliminated but laboratory C was retained), there is a difference in the Pb IVBA results with  $4562 \pm 183$  mg/kg for the SRC analysis versus  $4619 \pm 227$  mg/kg from Table E-2. Consequently, the Pb % IVBA results differ as well,  $71 \pm 4.7\%$  from the SRC analysis versus  $71.7 \pm 5.2\%$  from Table D-4.

**Table 7: SRC Statistical Results for FCRM**

Results	Mean	SD	99% PI	99% CI of mean	n
<b>Pb 3051A (mg/kg)</b>	6444	345	5489-7399	6285-6603	35
<b>As 3051A (mg/kg)</b>	728	65	550-905	700-756	40
<b>Pb IVBA (mg/kg)</b>	4562	183	4049-5076	4470-4654	30
<b>Pb IVBA (%)</b>	71	4.7	56-86	69-73 <sup>1</sup>	65 <sup>2</sup>

<sup>1</sup>A Fieller's method modified by Dilba was used to calculate the 99% confidence interval for the % IVBA.

<sup>2</sup>Based on thirty (30) IVBA extraction results and thirty-five (35) Method 3051A digestion results.

Figure 1. All Laboratory Results, Lead by Method 3051a  
(Mean  $\pm$  2 SEM)

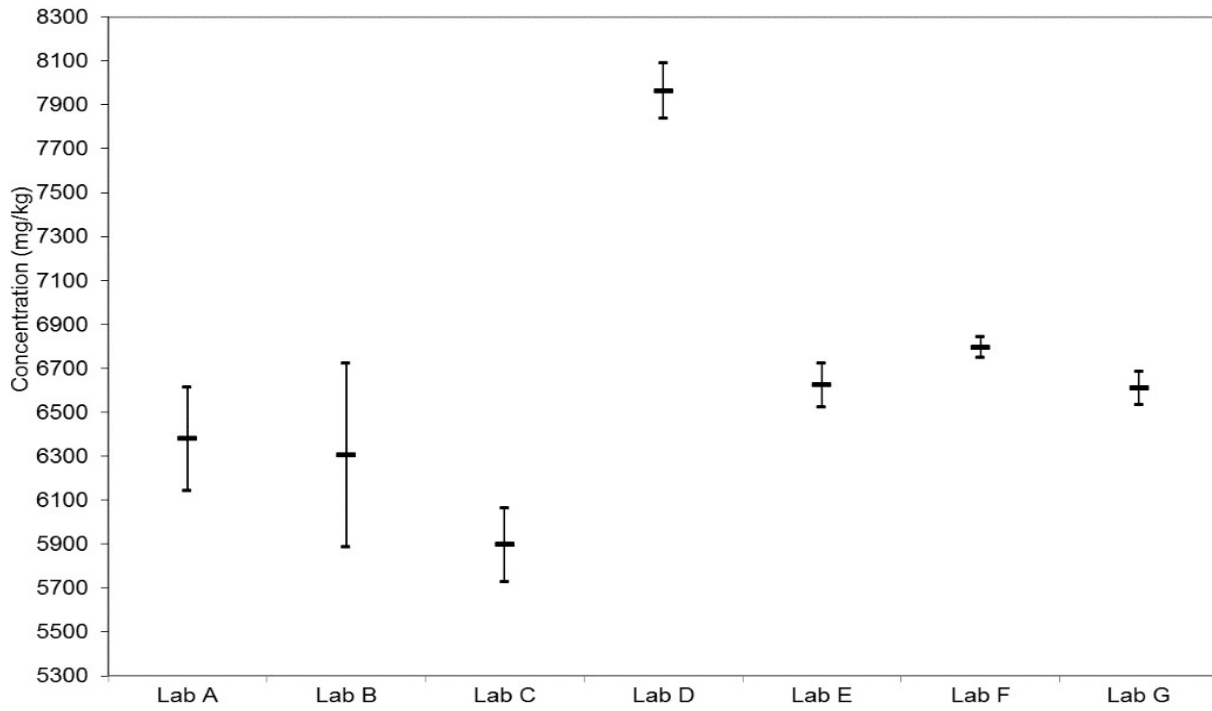


Figure 2. All Laboratory Results, Arsenic by Method 3051a  
(Mean  $\pm$  2 SEM)

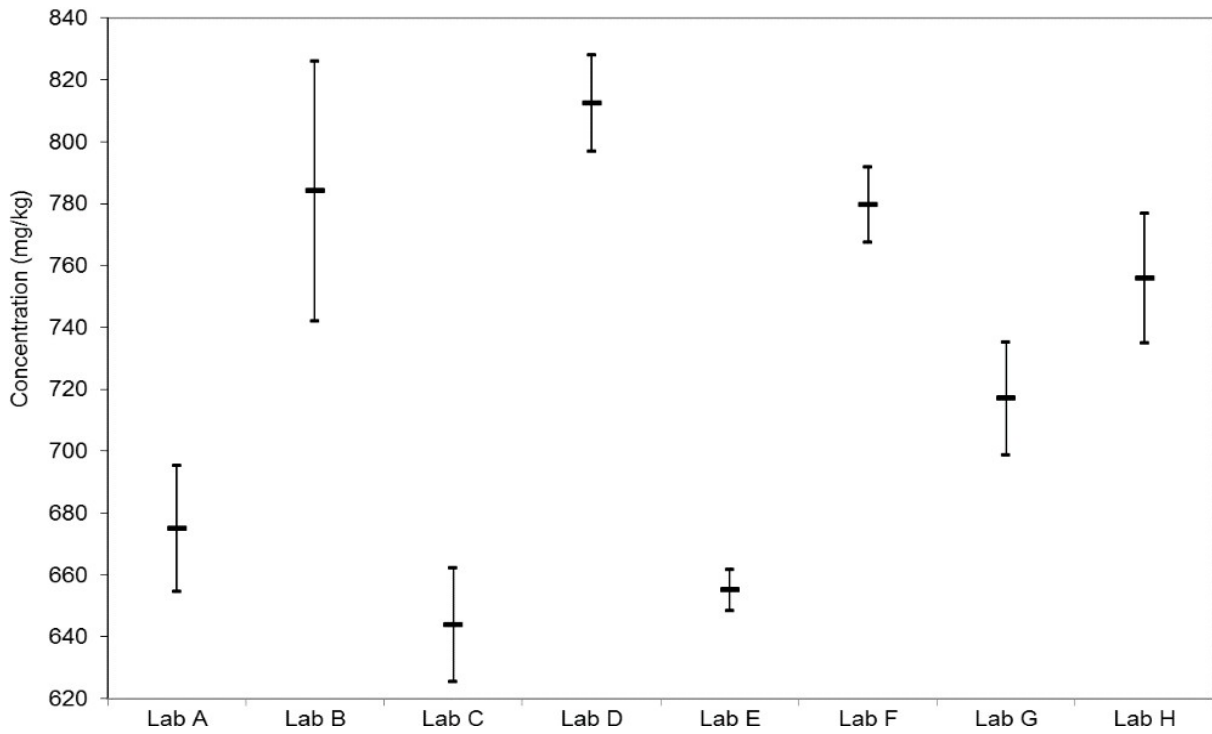
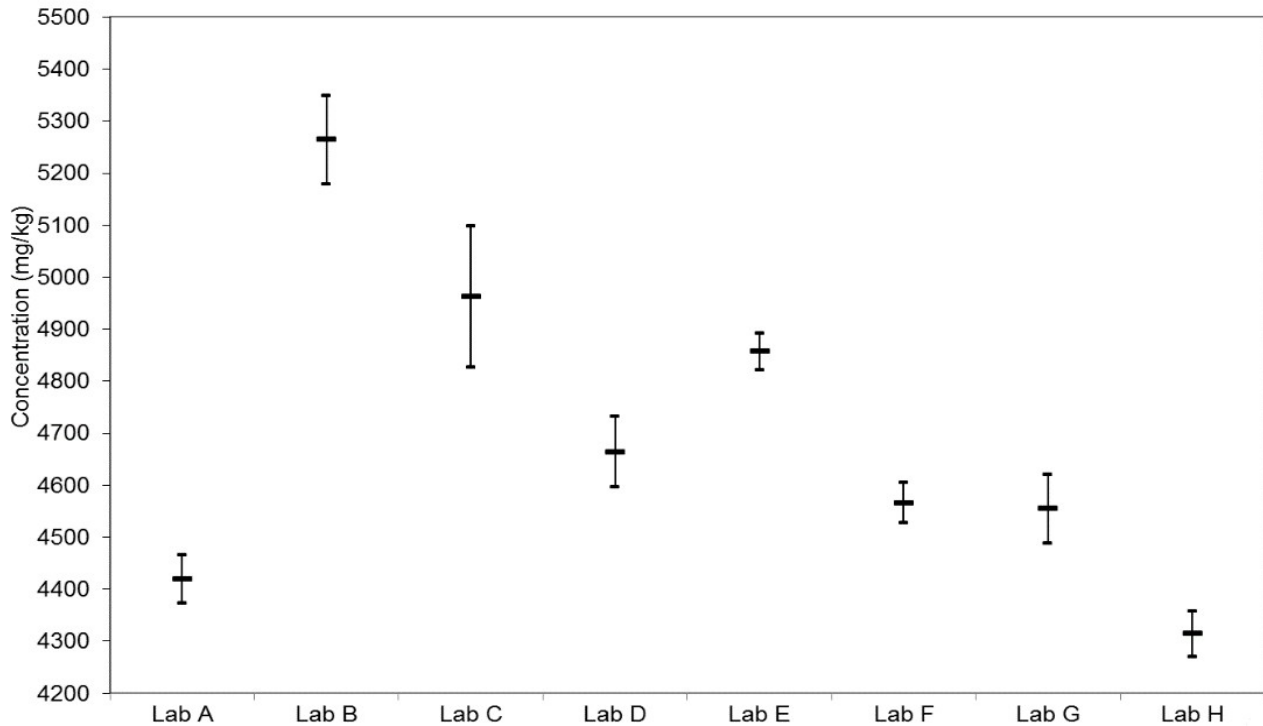


Figure 3. All Laboratory Results, Lead IVBA  
(Mean  $\pm$  2 SEM)



Note: Conclusion of Section V.E. – “Independent Statistical Analysis of the Round Robin Study Results by Syracuse Research Corporation (SRC) and Comparison to Shaw’s Standard Statistical Analysis Results”

The TRW IVBA committee selected the Shaw results provided in this report as the final results for the Round Robin Study, noting that there is little difference between the statistical results provided by the Shaw standard statistical approach and the results from the SRC statistical approach.

## VI. CONCLUSIONS AND RECOMMENDATIONS

The main objectives of this Study were to derive a Lead IVBA mean with known confidence for the FCRM, as well as to estimate the 99 percentile prediction interval. Another objective was to derive a mean value with known confidence for the lead and arsenic concentrations for the FCRM based on the EPA Method 3051A results from this Study. The Study results from the eight (8) participating laboratories were all determined to be acceptable using conventional statistics and the Grubb’s test for outliers. However, the t-test allowed for the exclusion of one laboratory’s Lead IVBA results and another laboratory’s EPA Method 3051A lead results, which allowed for the establishment of a Lead IVBA value for the FCRM with known and acceptable precision. This Study also provided for the determination of the lead and arsenic concentrations of the FCRM with known and acceptable precision. The associated QC results provided by the participating laboratories were all within the EPA SOP 9200.2-86 defined control limits, with a few noted exceptions.

Table 8 presents the final rounded values for the mean result and 99 percentile prediction intervals for the FCRM Lead IVBA results, as well as the EPA Method 3051A lead and arsenic values and

prediction intervals based on the pooled Study results. The prediction intervals for the EPA Method 3051A lead and arsenic values are presented in mg/Kg, and the Lead IVBA prediction intervals are presented in both mg/Kg and as Lead IVBA values.

**Table 8. Rounded Values for the FCRM Lead IVBA and EPA Method 3051A Lead and Arsenic Results**

<b>FCRM</b>	<b>Low 99% PI</b>	<b>Mean</b>	<b>High 99% PI</b>
<b>Lead Method 3051A (mg/Kg)</b>	5490	6440	7400
<b>Arsenic Method 3051A (mg/Kg)</b>	550	730	910
<b>Lead IVBA (mg/Kg)</b>	3990	4620	5250
<b>Lead IVBA (%)</b>	57.8	71.7	85.6

## VII. REFERENCES

Dilba, G., Bretz, F. and V.Guiard. 2006. Simultaneous Confidence Sets and Confidence Intervals for Multiple Ratios. *Journal of Statistical Planning and Inference*. 136, 2640-2658.

Fieller, E.C. 1954. Some Problems in Interval Estimation. *Journal of the Royal Statistical Society. Series B (Methodological)*, Vol. 16, No. 2, 175-185.

NFESC (2003). *Guide for Incorporating Bioavailability Adjustments into Human Health and Ecological Risk Assessments at U. S. Department of Defense Facilities, Naval Facilities Engineering Service Center (NFESC), Air Force Center for Environmental Excellence (AFCEE), Army Environmental Center (AEC), Part 2, Appendix C, June 2003.*

Ruby, M., Drexler, J., "*In Vitro Method for Determination of Lead Bioaccessibility Standard Operating Procedure for Stomach Phase Extraction*", June 2003.

Tamhane, A.C. and Logan, B.R. 2004. Finding the Maximum Safe Dose Level for Heteroscedastic Data. *J. Biopharmaceutical Statistics*, Vol.14, No. 4, 843-856.

US EPA (2012) *Standard Operating Procedure for an In Vitro Bioaccessibility Assay for Lead in Soil*, USEPA OSWER 9200.2-86, July 2012. On-line at: [http://epa.gov/superfund/bioavailability/pdfs/EPA\\_Pb\\_IVBA\\_SOP\\_040412\\_FINAL\\_SRC.pdf](http://epa.gov/superfund/bioavailability/pdfs/EPA_Pb_IVBA_SOP_040412_FINAL_SRC.pdf)

USEPA SW-846 EPA *Method 3051A Microwave Assisted Acid Digestion of Sediments, Sludges, Soils, and Oils*, USEPA Rev. 1, February, 2007. On-line at: <http://www.epa.gov/osw/hazard/testmethods/sw846/pdfs/3051a.pdf>

US EPA (2007). *Estimation of Relative Bioavailability of Lead in Soil and Soil-like Materials using In Vivo and In Vitro methods*, USEPA OSWER 9285.7-77, May 2007.

**Appendix A  
FCRM EPA Method 3051A Lead Results and Statistics**

**Table A-1. FCRM EPA Method 3051A Lead Results  
With Prediction Intervals and Confidence Intervals – All Labs**

<b>FCRM EPA Method 3051A Lead Results (mg/Kg)</b>																
<b>Laboratory &gt;</b>	<b>A</b>	<b>B</b>	<b>C</b>	<b>D</b>	<b>E</b>	<b>F</b>	<b>G</b>	<b>H</b>								
Replicate 1	6180	5600	5762	7812	6788	6838	6670	6246								
Replicate 2	6036	6500	6019	8141	6543	6742	6470	6513								
Replicate 3	6657	6350	6156	8087	6687	6815	6605	6471								
Replicate 4	6579	6870	5845	7878	6566	6739	6670	6538								
Replicate 5	6439	6200	5699	7898	6533	6844	6630	6737								
<b>Mean</b>	6378	6304	5896	7963	6623	6796	6609	6501								
<b>SD</b>	264	466	188	143	111	51	82	175								
<b>RSD</b>	4.1%	7.4%	3.2%	1.8%	1.7%	0.8%	1.2%	2.7%								
<table border="1" style="margin-left: auto; margin-right: auto;"> <thead> <tr> <th colspan="2"><b>Pooled Results (n-1) n=40</b></th> </tr> </thead> <tbody> <tr> <td><b>Mean</b></td> <td>6634</td> </tr> <tr> <td><b>SD</b></td> <td>604</td> </tr> <tr> <td><b>RSD</b></td> <td>9.1%</td> </tr> </tbody> </table>									<b>Pooled Results (n-1) n=40</b>		<b>Mean</b>	6634	<b>SD</b>	604	<b>RSD</b>	9.1%
<b>Pooled Results (n-1) n=40</b>																
<b>Mean</b>	6634															
<b>SD</b>	604															
<b>RSD</b>	9.1%															
<b>FCRM EPA Method 3051A Lead – 99 Percentile Prediction Interval (mg/Kg)</b>																
<b>Low 99 % PI</b>	<b>Mean</b>				<b>High 99 % PI</b>											
4978	6634				8290											
<b>± 99 % Prediction Interval = 25.0% of the Mean Value</b>																
<i>The range above should be used to determine if a laboratory EPA Method 3051A lead result is acceptable.</i>																
<b>FCRM EPA Method 3051A Lead – 99 Percentile Confidence Interval of the Mean</b>																
6634 = Mean	96 = SD of the Mean				1.4% = RSD of the Mean											
<b>Low 99 % CI</b>	<b>Mean</b>				<b>High 99 % CI</b>											
6375	6634				6892											
<b>± 99 Percentile of the Confidence Interval of the Mean = 3.9% of the Mean Value</b>																
<i>The range above can be used to statistically assess the confidence in the accuracy of the mean result.</i>																

SD = Standard Deviation  
RSD = Relative Standard Deviation  
CI = Confidence Interval  
PI = Prediction Interval



**Appendix A  
FCRM EPA Method 3051A Lead Results and Statistics**

**Table A-2. FCRM EPA Method 3051A Batch QC Sample Lead Results**

Laboratory>	A	B	C	D	E	F	G	H	Mean
Blank Spike Recovery (Nominal: 10 mg/L) (Range: 85% to 115%)	93.2%	111.8%	96.3%	108.0%	96.9%	104.0%	101.0%	105.0%	102.0%
FCRM Matrix Spike Recovery (Nominal: 10 mg/L) (Range: 75% to 125%)	101.9%	88%	83.2%	57.0%	98.1%	76.5%	80.0%	81.2%	83.2%
NIST SRM 2710a Digestion Lead Results NIST Certificate (Nominal: 5100 mg/Kg) (Range: 4700- 5800 mg/kg)	5554	5370	4882	4912	(4537)	5491	5195	5181	5140
Lead IVBA NIST SRM 2710a Recovery Based on NIST Certificate Leachable Value of 5100 mg/Kg	108.9%	105.3%	95.7%	96.3%	89.0%	107.7%	101.9%	101.6%	100.8%

Values in parentheses are outside the associated control limits.

**Table A-3. FCRM EPA Method 3051A Lead Digestion Analysis of Variance**

Excel ANOVA: Single Factor (Lead) Note: alpha at 0.05 (95 percentile)						
SUMMARY						
Groups	Count	Sum	Mean	Variance		
Laboratory A	5	31891	6378	69533		
Laboratory B	5	31520	6304	216830		
Laboratory C	5	29480	5896	35459		
Laboratory D	5	39816	7963	20328		
Laboratory E	5	33117	6623	12215		
Laboratory F	5	33978	6796	2648		
Laboratory G	5	33045	6609	6805		
Laboratory H	5	32505	6501	30784		
ANOVA						
Source of	SS	df	MS	F	P-value	F-Crit
Interlaboratory	12650866	7	1807266	36.6	1.59 E-13	2.31
Intralaboratory	1578406	32	49325			
Total	14229273	39				
SS = Sum of Squares Df = Degrees of Freedom MS = Mean Square F = F Value Calculated F-Crit = Critical Value of F P-value = Probability Value						

**Appendix B  
FCRM EPA Method 3051A Lead Results and t-Test**

**Table B-1. FCRM Lead Results and t-Test for Laboratory D Digestion Data**

FCRM EPA Method 3051A Lead Results t-Test (mg/Kg)									
Laboratory>	A	B	C	E	F	G	H	Laboratory>	D
Replicate 1	6180	5600	5762	6788	6838	6670	6246	Replicate 1	7812
Replicate 2	6036	6500	6019	6543	6742	6470	6513	Replicate 2	8141
Replicate 3	6657	6350	6156	6687	6815	6605	6471	Replicate 3	8087
Replicate 4	6579	6870	5845	6566	6739	6670	6538	Replicate 4	7878
Replicate 5	6439	6200	5699	6533	6844	6630	6737	Replicate 5	7898
Mean	6378	6304	5896	6623	6796	6609	6501	Mean	7963
SD	264	466	188	111	51	82	175	SD	143
RSD	4.1%	7.4%	3.2%	1.7%	0.76%	1.2%	2.7%	RSD	1.8%

Labs A-H		Percent Difference	Lab D	
	n=35			N=5
Mean	6444	21.1%	Mean	7963
SD	345		SD	143
RSD	5.4%		RSD	1.8%

**Excel t-Test: Two-Sample Assuming Equal Variances**  
alpha = 0.01 1 in 99)

	Lab A-H	Lab D
Mean	6444	7963
Variance	119094	20328
Observations	35	5
Pooled Variance	108698	
Hypothesized Mean Difference	0	
Df	38	
t-Stat	9.64	
P(T ≤ t) (two-tail)	9.40 E-12	
t-Critical (two-tail)	2.71	

The t-Stat value of 9.64 is greater than t-Critical two-tail value of 2.71; therefore, the null hypothesis that the means are not significantly different (zero difference, same population), can be rejected.

A P(T ≤ t) two tail value of less than 0.01 indicates a greater than 99% probability that the means of the two groups do not come from the same population.

Based on the above statements and t-test results, the results from Laboratory D can be excluded.

**Appendix B  
FCRM EPA Method 3051A Lead Results and t-Test**

**Table B-2. FCRM EPA Method 3051A Lead Results  
With Prediction Intervals and Confidence – Minus Laboratory D**

<b>FCRM EPA Method 3051A Lead Results (mg/Kg)</b>																
<b>Laboratory &gt;</b>	<b>A</b>	<b>B</b>	<b>C</b>	<b>D</b>	<b>E</b>	<b>F</b>	<b>G</b>	<b>H</b>								
<b>Replicate 1</b>	6180	5600	5762		6788	6838	6670	6246								
<b>Replicate 2</b>	6036	6500	6019		6543	6742	6470	6513								
<b>Replicate 3</b>	6657	6350	6156		6687	6815	6605	6471								
<b>Replicate 4</b>	6579	6870	5845		6566	6739	6670	6538								
<b>Replicate 5</b>	6439	6200	5699		6533	6844	6630	6737								
<b>Mean</b>	6378	6304	5896		6623	6796	6609	6501								
<b>SD</b>	264	466	188		111	51	82	175								
<b>RSD</b>	4.1%	7.4%	3.2%		1.7%	0.8%	1.2%	2.7%								
<table border="1" style="margin: auto;"> <thead> <tr> <th colspan="2"><b>Pooled Results (n-1) n=35</b></th> </tr> </thead> <tbody> <tr> <td><b>Mean</b></td> <td>6444</td> </tr> <tr> <td><b>SD</b></td> <td>345</td> </tr> <tr> <td><b>RSD</b></td> <td>5.4%</td> </tr> </tbody> </table>									<b>Pooled Results (n-1) n=35</b>		<b>Mean</b>	6444	<b>SD</b>	345	<b>RSD</b>	5.4%
<b>Pooled Results (n-1) n=35</b>																
<b>Mean</b>	6444															
<b>SD</b>	345															
<b>RSD</b>	5.4%															
<b>FCRM EPA Method 3051A Lead Results – 99 Percentile Prediction Interval (mg/Kg)</b>																
<b>Low 99 % PI</b>			<b>Mean</b>			<b>High 99 % PI</b>										
5489			6444			7399										
<b>± 99 % Prediction Interval = 14.8% of the Mean Value</b>																
<i>The range above should be used to determine if a laboratory EPA Method 3051A lead result is acceptable.</i>																
<b>FCRM EPA Method 3051A Lead Results – 99 Percentile Confidence Interval of the Mean</b>																
6444 = Mean			58 =SD of the Mean			0.91% = RSD of the Mean										
<b>Low 99 % CI</b>			<b>Mean</b>			<b>High 99 % CI</b>										
6285			6444			6603										
<b>± 99 Percentile of the Confidence Interval of the Mean = 2.5% of the Mean Value</b>																
<i>The range above can be used to statistically assess the confidence in the accuracy of the mean result.</i>																

SD = Standard Deviation  
RSD = Relative Standard Deviation  
CI = Confidence Interval  
PI = Prediction Interval

**Appendix C  
FCRM EPA Method 3051A Arsenic Results and Statistics**

**Table C-1. FCRM Laboratory Arsenic Results  
With Prediction Intervals and Confidence Intervals – All Labs**

<b>FCRM EPA Method 3051A Arsenic Results (mg/Kg)</b>																
<b>Laboratory &gt;</b>	<b>A</b>	<b>B</b>	<b>C</b>	<b>D</b>	<b>E</b>	<b>F</b>	<b>G</b>	<b>H</b>								
<b>Replicate 1</b>	652	716	621	800	649	776	715	748								
<b>Replicate 2</b>	653	760	673	841	656	789	700	731								
<b>Replicate 3</b>	700	792	655	816	649	770	712	742								
<b>Replicate 4</b>	696	830	639	799	654	765	706	790								
<b>Replicate 5</b>	674	822	631	806	667	798	752	768								
<b>Mean</b>	675	784	644	812	655	779	717	756								
<b>SD</b>	23	47	21	17	7	14	21	24								
<b>RSD</b>	3.4%	6.0%	3.2%	2.1%	1.1%	1.8%	2.9%	3.1%								
<table border="1" style="margin: auto;"> <thead> <tr> <th colspan="2"><b>Pooled Results n=40</b></th> </tr> </thead> <tbody> <tr> <td><b>Mean</b></td> <td>728</td> </tr> <tr> <td><b>Std Dev</b></td> <td>65</td> </tr> <tr> <td><b>RSD</b></td> <td>8.9%</td> </tr> </tbody> </table>									<b>Pooled Results n=40</b>		<b>Mean</b>	728	<b>Std Dev</b>	65	<b>RSD</b>	8.9%
<b>Pooled Results n=40</b>																
<b>Mean</b>	728															
<b>Std Dev</b>	65															
<b>RSD</b>	8.9%															
<b>FCRM EPA Method 3051A Arsenic – 99 Percentile Prediction Interval (mg/Kg)</b>																
<b>Low 99 % PI</b>			<b>Mean</b>			<b>High 99 % PI</b>										
550			728			905										
<b>± 99 Prediction Interval = 24.2% of the Mean Value</b>																
<i>The range above should be used to determine if a laboratory EPA Method 3051A arsenic result is acceptable.</i>																
<b>FCRM EPA Method 3051A Arsenic – 99 Percentile Confidence Interval of the Mean (mg/Kg)</b>																
728 = Mean			10.2 SD of the Mean			1.41% = RSD of the Mean										
<b>Low 99 % CI</b>			<b>Mean</b>			<b>High 99 % CI</b>										
700			728			756										
<b>± 99 Percentile of the Confidence Interval of the Mean = 3.8% of the Mean Value</b>																
<i>The range above can be used to assess the confidence in the accuracy of the mean result.</i>																

SD = Standard Deviation  
RSD = Relative Standard Deviation  
CI = Confidence Interval  
PI = Prediction Interval

**Appendix C**  
**FCRM EPA Method 3051A Arsenic Results and Statistics**

**Table C-2. FCRM EPA Method 3051A Batch QC Sample Arsenic Results**

Laboratory>	A	B	C	D	E	F	G	H	Mean
<b>Blank Spike Recovery (Range: 85% to 115%)</b>	90.8%	114.8%	90.5%	110.0%	95.8%	102.0%	99.9%	104.0%	101.0%
<b>RM Matrix Spike Recovery (Range: 75% to 125%)</b>	103.8%	NA	106.7%	106.0%	120.7%	97.9%	100.0%	97.1%	104.5%
<b>NIST SRM 2710a Arsenic (Mean: 1400 mg/Kg) (Range: 1300-1600 mg/Kg)</b>	1592	(1650)	1460	1546	1322	(1684)	1505	1577	1592

Values in parentheses are outside the associated control limits.

**Table C-3. FCRM EPA Method 3051A Arsenic Results Analysis of Variance**

Excel ANOVA: Single Factor (Arsenic) Note: alpha at 0.05 (95 percentile)						
SUMMARY						
Groups	Count	Sum	Mean	Variance		
Laboratory A	5	3375	675	520		
Laboratory B	5	3920	784	2206		
Laboratory C	5	3220	644	430		
Laboratory D	5	4062	812	301		
Laboratory E	5	3274	655	56.4		
Laboratory F	5	3897	779	187		
Laboratory G	5	3584	717	424		
Laboratory H	5	3778	756	553		
ANOVA						
Source of Variation	SS	Df	MS	F	P-value	F-Crit
Interlaboratory	145124	7	20732	35.5	2.51 E-13	2.31
Intralaboratory	18708	32	585			
<b>Total</b>	<b>163833</b>	<b>39</b>				
<b>SS = Sum of Squares</b> <b>df = Degrees of Freedom</b> <b>MS = Mean Square</b> <b>F = F Value Calculated</b> <b>F-Crit = Critical Value of F</b> <b>P-value = Probability Value</b>						

**Appendix D  
FCRM Lead IVBA Results and Statistics**

**Table D-1. FCRM Lead IVBA Results  
With Prediction Intervals and Confidence Intervals**

FCRM Lead IVBA Results (mg/Kg)																
Laboratory >	A	B	C	D	E	F	G	H								
Replicate 1	4360	5210	4870	4762	4921	4609	4538	4314								
Replicate 2	4491	5420	5000	4639	4840	4604	4434	4285								
Replicate 3	4387	5260	5060	4622	4849	4549	4584	4267								
Replicate 4	4448	5260	5130	4576	4857	4563	4589	4393								
Replicate 5	4409	5170	4750	4720	4816	4505	4626	4310								
Mean	4419	5264	4962	4664	4856	4566	4554	4314								
SD	52	95	152	76	39	43	74	48								
RSD	1.17%	1.81%	3.07%	1.62%	0.81%	0.94%	1.63%	1.12%								
<table border="1" style="margin-left: auto; margin-right: auto;"> <thead> <tr> <th colspan="2">Pooled Results (n-1) n=40</th> </tr> </thead> <tbody> <tr> <td>Mean</td> <td>4700</td> </tr> <tr> <td>SD</td> <td>304</td> </tr> <tr> <td>RSD</td> <td>6.5%</td> </tr> </tbody> </table>									Pooled Results (n-1) n=40		Mean	4700	SD	304	RSD	6.5%
Pooled Results (n-1) n=40																
Mean	4700															
SD	304															
RSD	6.5%															
FCRM Lead IVBA – 99 Percentile Prediction Interval (mg/Kg)																
Low 99 % PI			Mean			High 99 % PI										
3866			4700			5534										
± 99 % Prediction Interval = 17.7% of the Mean Value																
<i>The range above should be used to determine if a laboratory EPA SOP 9200.2-86 IVBA extracted lead result is acceptable.</i>																
FCRM Lead IVBA – 99 Percentile Prediction Interval																
Low 99 % PI			Mean			High 99 % PI										
49.8			70.8			91.9										
± 99 % Prediction Interval = 29.7% of the Mean Value																
The pooled EPA SOP 9200.2-86 lead extraction results have been divided by the pooled EPA Method 3051A digestion results to derive an EPA 9200.2-86 Lead IVBA value that includes the variance of both extraction and digestion results																
IVBA = 70.8 or 70.8%			SD = 7.9			RSD = 11.2%										
<i>The range above should be used to determine if a laboratory EPA SOP 9200.2-86 lead IVBA result is acceptable.</i>																
FCRM Lead IVBA – Confidence Interval of the Mean																
70.8 = Mean			0.88 SD of the Mean			1.24% = RSD of the Mean										
Low 99 % PI			Mean			High 99 % PI										
68.5			70.8			73.2										
± 99 Percentile of the Confidence Interval of the Mean = 3.3% of the Mean Value																
<i>The range above can be used to statistically assess the confidence in the accuracy of the mean result.</i>																

SD = Standard Deviation  
RSD = Relative Standard Deviation  
CI = Confidence Interval  
PI = Prediction Interval

**Appendix D  
FCRM Lead IVBA Results and Statistics**

**Table D-2. FCRM Lead IVBA and EPA Method 3051A Lead Batch QC Sample Results**

Laboratory>		A	B	C	D	E	F	G	H	Mean
1	Reagent Blank <25 ug/L	1	<3	<40	<1	1.8	<2.5	<11	4.5	NA
2	Bottle Blank ug/L <50 ug/L	1	<3	<40	<1	1.6	<2.5	<11	4.1	NA
3	Blank Lead Spike Recovery (Control Limits: 85% to 115%)	91.4%	104.0%	108.3%	100.0%	98.0%	113.0%	102.0%	98.4%	101.9%
4	FCRM Lead Matrix Spike Recovery (Control Limits: 75% to 125%)	100.2%	117.6%	116.2%	62%	103.1%	62%	122%	92.2%	96.9%
<hr/>										
5	NIST SRM 2710a mg/Kg Lead IVBA Results (Nominal = 3440 mg/Kg)	3325	4000	3943	3595	3615	3400	3393	3332	3575
6	NIST SRM 2710a Lead IVBA Percent Recovery (Nominal: 3440 mg/Kg) (Control Limits: 80% to 120%)	96.7%	116.3%	114.6%	104.5%	105.1%	98.8%	98.6%	96.9%	103.9%
<hr/>										
7	NIST SRM 2710a EPA Method 3051A Digestion Lead Results (mg/Kg) NIST Certificate (Nominal: 5100 mg/Kg) (Range: 4700 to 5800 mg/Kg)	5554	5370	4882	4912	(4537)	5491	5195	5181	5140
8	NIST SRM 2710a EPA Method 3051A Lead Percent Recovery Based on NIST Certificate Leachable Value of 5100 mg/Kg (Control Limits: 80% to 120%)	108.9%	105.3%	95.7%	96.3%	89.0%	107.7%	101.9%	101.6%	100.8%
<hr/>										
9	Lead IVBA value for NIST SRM 2710a, based on the mean EPA 3051A lead value using EPA SOP 9200.2-86 criteria. (Mean 67.5%: Control Limits: 60.7% - 74.2%)	65.2	(78.4)	(77.3)	70.5	70.9	66.7	66.5	65.3	70.1
<hr/>										
10	NIST SRM 2710a Lead IVBA Results Based on both the IVBA Lead Extraction and EPA 3051A Digestion of NIST SRM 2710a During this Study Lead IVBA: (Mean 67.5%: Control Limits: 60.7% to 74.2%)	59.9	(74.5)	(80.8)	73.2	(79.7)	61.9	65.3	64.3	69.9
11	Lead IVBA value for NIST SRM 2710a based on the Study Lead IVBA and EPA Method 3051A results. (i.e. Row 10 divided by IVBA 67.5%)	88.7%	110.4%	119.7%	108.4%	118.0%	91.7%	96.8%	95.3%	103.6%

NA = Not Applicable

Values in parentheses are outside the associated control limits.

**Appendix D**  
**FCRM Lead IVBA Results and Statistics**

**Table D-3. FCRM Lead IVBA - Analysis of Variance Results**

Excel ANOVA: Single Factor (Lead) Note: alpha at 0.05 (95 percentile)						
SUMMARY						
<i>Groups</i>	<i>Count</i>	<i>Sum</i>	<i>Mean</i>			
<i>Variance</i>						
Laboratory A	5	22095	4419			
Laboratory B	5	26320	5264			
Laboratory C	5	24810	4962			
Laboratory D	5	23319	4664			
Laboratory E	5	24282	4856			
Laboratory F	5	22830	4566			
Laboratory G	5	22771	4554			
Laboratory H	5	21569	4314			
ANOVA						
<i>Source of</i>	<i>SS</i>	<i>df</i>	<i>MS</i>	<i>F</i>	<i>P-value</i>	<i>F-Crit</i>
Interlaboratory	3399119	7	485588	75.1	4.83 E-18	2.31
Intralaboratory	207022	32	6469			
<b>Total</b>	<b>3606142</b>	<b>39</b>				
<p><b>SS = Sum of Squares</b>  <b>df = Degrees of Freedom</b>  <b>MS = Mean Square</b>  <b>F = F Value Calculated</b>  <b>F-Crit = Critical Value of F</b>  <b>P-value = Probability Value</b></p>						



**Appendix D  
FCRM Lead IVBA Results and Statistics**

**Table D-4. FCRM Lead IVBA Prediction and Confidence Intervals  
Minus Lab B Lead IVBA and Lab D EPA Method 3051A Lead Results**

<b>FCRM Lead IVBA – 99 Percentile Prediction Interval</b>		
<b>Low 99 % PI</b>	<b>Mean</b>	<b>High 99 % PI</b>
57.8	71.7	85.6
<b>± 99 Prediction Interval = 19.4% of the Mean Value</b>		
<b>The pooled extraction results been divided by the pooled digestion results to derive a Lead IVBA and includes the variance of both extraction and digestion results</b>		
IVBA = 71.7 or 71.7%	SD = 5.2	RSD = 7.3%
<i>The range above should be used to determine if a laboratory lead IVBA result is acceptable.</i>		
<b>FCRM Lead IVBA – 99 Percentile Confidence Interval of the Mean</b>		
71.7 = Mean	0.62 SD of the Mean	0.87% = RSD of the Mean
<b>Low 99 % CI</b>	<b>Mean</b>	<b>High 99 % CI</b>
70.0	71.7	73.3
<b>± 99 Percentile of the Confidence Interval of the Mean = 2.3% of the Mean Value</b>		
<i>The range above can be used to statistically assess the confidence in the accuracy of the mean result.</i>		

SD = Standard Deviation  
RSD = Relative Standard Deviation  
CI = Confidence Interval  
PI = Prediction Interval

**Appendix E  
FCRM Lead IVBA Results and t-Test**

**Table E-1. FCRM Lead IVBA Results and t-Test for Laboratory B**

FCRM Lead IVBA Results t-Test (mg/Kg)									
Laboratory >	A	C	D	E	F	G	H	Laboratory >	B
Replicate 1	4360	4870	4762	4921	4609	4538	4314	Replicate 1	5210
Replicate 2	4491	5000	4639	4840	4604	4434	4285	Replicate 2	5420
Replicate 3	4387	5060	4622	4849	4549	4584	4267	Replicate 3	5260
Replicate 4	4448	5130	4576	4857	4563	4589	4393	Replicate 4	5260
Replicate 5	4409	4750	4720	4816	4505	4626	4310	Replicate 5	5170
Mean	4419	4962	4664	4856	4566	4554	4314	Mean	5264
SD	52	152	76	39	43	74	48	SD	95
RSD	1.2%	3.1%	1.6%	0.8%	0.94%	1.6%	1.1%	RSD	1.8%
<b>Labs A-H</b>		<b>Percent Difference</b>					<b>Lab B</b>		
	<b>n=35</b>							<b>n=5</b>	
Mean	4619	13.1%					Mean	5264	
SD	227						SD	95	
RSD	4.9%						RSD	1.8%	

Excel t-Test: Two-Sample Assuming Equal Variances		
alpha = 0.01 (1 in 99)		
	Labs A-H	Lab B
Mean	4619	5264
Variance	51519	9030
Observations	35	5
Pooled Variance	47047	
Hypothesized Mean Difference	0	
Df	38	
t-Stat	6.22	
P(T ≤ t) two-tail	2.87 E-07	
t-Critical two-tail	2.71	
<p>The t-Stat value of 6.22 is greater than t-Critical two-tail value of 2.71; therefore, the null hypothesis that the means are not significantly different (zero difference, same population), can be rejected.</p> <p>A P(T ≤ t) two tail value of less than 0.01 indicates a greater than 99% probability that the means of the two groups do not come from the same population.</p> <p>Based on the above statements and t-test results, the results from Laboratory B can be excluded.</p>		

SD = Standard Deviation  
RSD = Relative Standard Deviation

**Appendix E  
FCRM Lead IVBA Results and t-Test**

**Table E-2. FCRM Lead IVBA Results  
With Prediction Intervals and Confidence Intervals - Minus Lab B**

FCRM Lead IVBA Results (mg/Kg)																
Laboratory >	A	B	C	D	E	F	G	H								
Replicate 1	4360		4870	4762	4921	4609	4538	4314								
Replicate 2	4491		5000	4639	4840	4604	4434	4285								
Replicate 3	4387		5060	4622	4849	4549	4584	4267								
Replicate 4	4448		5130	4576	4857	4563	4589	4393								
Replicate 5	4409		4750	4720	4816	4505	4626	4310								
<b>Mean</b>	4419		4962	4664	4856	4566	4554	4314								
<b>SD</b>	52		152	76	39	43	74	48								
<b>RSD</b>	1.2%		3.1%	1.6%	0.8%	0.94%	1.6%	1.1%								
<table border="1" style="margin: auto;"> <thead> <tr> <th colspan="2">Pooled Results (n-1) n=35</th> </tr> </thead> <tbody> <tr> <td>Mean</td> <td>4619</td> </tr> <tr> <td>SD</td> <td>227</td> </tr> <tr> <td>RSD</td> <td>4.9%</td> </tr> </tbody> </table>									Pooled Results (n-1) n=35		Mean	4619	SD	227	RSD	4.9%
Pooled Results (n-1) n=35																
Mean	4619															
SD	227															
RSD	4.9%															
FCRM Lead IVBA – 99 Percentile Prediction Interval (mg/Kg)																
<b>Low 99 % PI</b>	<b>Mean</b>			<b>High 99 % PI</b>												
3991	4619			5247												
<b>± 99 % Prediction Interval = 13.6% of the Mean Value</b>																
<i>The range above should be used to determine if a laboratory IVBA extracted lead result is acceptable.</i>																
FCRM Lead IVBA – 99 Percentile Prediction Interval																
<b>Low 99 % PI</b>	<b>Mean</b>			<b>High 99 % PI</b>												
50.5	69.6			88.8												
<b>± 99 % Prediction Interval = 27.5% of the Mean Value</b>																
<b>The pooled extraction results been divided by the pooled digestion results to derive an Lead IVBA and includes the variance of both extraction and digestion results</b>																
IVBA = 69.6 or 69.6%	SD = 7.2			RSD = 10.3%												
<i>The range above should be used to determine if a laboratory lead IVBA result is acceptable.</i>																
FCRM Lead IVBA – 99 Percentile Confidence Interval of the Mean																
69.6 = Mean	0.83 = SD of the Mean			1.2% = RSD of the Mean												
<b>Low 99 % CI</b>	<b>Mean</b>			<b>High 99 % CI</b>												
67.4	69.6			71.8												
<b>± 99 Percentile of the Confidence Interval of the Mean = 3.2% of the Mean Value</b>																
<i>The range above can be used to statistically assess the confidence in the accuracy of the mean result.</i>																

SD = Standard Deviation  
RSD = Relative Standard Deviation  
CI = Confidence Interval  
PI = Prediction Interval

Appendix F  
NIST SRM 2710a Certificate of Analysis



National Institute of Standards & Technology

# Certificate of Analysis

Standard Reference Material<sup>®</sup> 2710a

Montana I Soil

Highly Elevated Trace Element Concentrations

This Standard Reference Material (SRM) is intended primarily for use in the analysis of soils, sediments, or other materials of a similar matrix. One unit of SRM 2710a consists of 50 g of the dried, powdered soil, blended with lead oxide.

**Certified Values:** The certified concentrations for 22 elements, expressed as mass fractions [1] on a dry-mass basis, are provided in Table 1. Certified values are based on results obtained from critically evaluated independent analytical techniques. A NIST certified value is a value for which NIST has the highest confidence in its accuracy in that all known or suspected sources of bias have been investigated or taken into account [2].

**Reference Values:** The reference values for 13 constituents, expressed as mass fractions on a dry-mass basis, are provided in Table 2. Ten reference values are based on results obtained from a single NIST analytical method, and three are based on results from two NIST analytical methods. Reference values are non-certified values that are the best estimate of the true value; however, the values do not meet NIST criteria for certification and are provided with associated uncertainties that may not include all sources of uncertainty [2].

**Information Values:** The values for 13 elements are provided in Table 3 for information purposes only. These are non-certified values with no uncertainty assessed. The information values included in this certificate are based on results obtained from one NIST method.

**Expiration of Certification:** The certification of SRM 2710a is valid, within the measurement uncertainties specified, until **1 January 2019**, provided the SRM is handled in accordance with the instructions given in this certificate (see "Instructions for Use"). This certification is nullified if the SRM is damaged, contaminated, or otherwise modified.

**Maintenance of SRM Certification:** NIST will monitor this SRM over the period of its certification. If substantive technical changes occur that affect the certification before the expiration of this certificate, NIST will notify the purchaser. Registration (see attached sheet) will facilitate notification.

E.A. Mackey and R.R. Greenberg of the NIST Analytical Chemistry Division were responsible for coordination of the technical measurements leading to certification.

Statistical analyses were performed by J.H. Yen of the NIST Statistical Engineering Division.

Support aspects involved in the issuance of this SRM were coordinated through the NIST Measurement Services Division.

Stephen A. Wise, Chief  
Analytical Chemistry Division

Gaithersburg, MD 20899  
Certificate Issue Date: 7 April 2009

Robert L. Watters, Jr., Chief  
Measurement Services Division

# Appendix F

## NIST SRM 2710a Certificate of Analysis

### INSTRUCTIONS FOR USE

**Sampling:** The SRM should be thoroughly mixed by repeatedly inverting and rotating the bottle horizontally before removing a test portion for analysis. A minimum mass of 250 mg (dry mass - see *Instructions for Drying*) should be used for analytical determinations to be related to the mass fraction values in this Certificate of Analysis.

To obtain the certified values, sample preparation procedures should be designed to effect complete dissolution. If volatile elements (i.e., arsenic, mercury, selenium) will be determined, precautions should be taken in the dissolution of SRM 2710a to avoid volatilization losses.

**Drying:** To relate measurements to the certified, reference, and information values that are expressed on a dry-mass basis, users should determine a drying correction at the time of each analysis. The recommended drying procedure is oven drying for 2 h at 110 °C. Note that analytical determination of volatile elements (i.e., arsenic, mercury, selenium) should be determined on samples as received; separate samples should be dried as previously described to obtain a correction factor for moisture. Correction for moisture must be made to the data for volatile elements before comparing to the certified values. This procedure ensures that these elements are not lost during drying. The mass loss on drying for this material as bottled was approximately 2 %, but this value may change once the bottle is opened and the soil is exposed to air.

### SOURCE, PREPARATION, AND ANALYSIS

**Source and Preparation of Material<sup>1</sup>:** The U.S. Geological Survey (USGS), under contract to NIST, collected and processed the material for SRM 2710a. The original collection site used for SRM 2710 was no longer available due to remediation efforts by the Montana Department of Environmental Quality. An alternative nearby site, located within the flood plain of the Silver Bow Creek, was selected. The site is approximately five miles west of Butte, Montana. Soil for SRM 2710a was placed in 22 plastic-lined five-gallon buckets using a common garden spade. The buckets were sealed and transferred to the USGS using a commercial freight carrier. At the USGS, the SRM 2710a soil was dried at room temperature, disaggregated, and sieved to remove coarse material ( $\geq 2$  mm). The resulting soil was ball-milled in 50 kg portions together with an amount of lead oxide sufficient to achieve a mass fraction of 0.55 % lead in the final product. The entire ball-milled batch of soil was transferred to a cross-flow V-blender for mixing. The blended soil was radiation sterilized prior to bottling. In the final preparation step the blended material was split into containers using a custom-designed spinning riffler, which was used to divide the material into smaller batches, and then used to apportion approximately 50 g into each pre-cleaned bottle.

Every 100th bottle was set aside for chemical analyses designed to assess material homogeneity using X-ray fluorescence spectrometry (XRF), inductively coupled plasma optical emission spectrometry (ICP-OES), and inductively coupled plasma mass spectrometry (ICP-MS) at the USGS. Homogeneity assessments were performed at NIST as well, and results indicated that additional processing was needed to achieve optimum homogeneity. The material from all bottles was combined, and then ground in batches between stainless steel plates for a time sufficient to produce a powder of which  $\geq 95$  %, by mass, passed through a 200 mesh (74  $\mu$ m) sieve. The resulting powder was blended, and 50 g portions were dispensed into bottles using the spinning riffler. Results from additional analyses indicated material homogeneity was acceptable (see below).

**Analysis:** The homogeneity was assessed for selected elements in the bottled material using X-ray fluorescence spectrometry and instrumental neutron activation analysis (INAA). The estimated relative standard deviation for material inhomogeneity is  $< 1$  % and no component for inhomogeneity was included in the expanded uncertainties of the certified or reference values.

Analyses of this material were performed at NIST and at the USGS (Denver, CO). Results from NIST were used to provide the certified, reference, and information values shown in Tables 1, 2, and 3 respectively. Results from the USGS were used to confirm those values. The analytical techniques used for each element are listed in Table 4; the analysts are listed in Tables 5 and 6.

---

<sup>1</sup> Certain commercial equipment, instruments, or materials are identified in this certificate in order to specify adequately the experimental procedure. Such identification does not imply recommendation or endorsement by the National Institute of Standards and Technology, nor does it imply that the materials or equipment identified are necessarily the best available for the purpose.

## Appendix F

### NIST SRM 2710a Certificate of Analysis

Table 1. Certified Values <sup>(a,b)</sup> (Dry-Mass Basis) for Selected Elements in SRM 2710a

Element	Mass Fraction (%)	Element	Mass Fraction (mg/kg)
Aluminum	5.95 ± 0.05	Antimony	52.5 ± 1.6
Arsenic	0.154 ± 0.010	Barium	792 ± 36
Calcium	0.964 ± 0.045	Cadmium	12.3 ± 0.3
Copper	0.342 ± 0.005	Cobalt	5.99 ± 0.14
Iron	4.32 ± 0.08	Lanthanum	30.6 ± 1.2
Lead	0.552 ± 0.003	Mercury	9.88 ± 0.21
Magnesium	0.734 ± 0.038	Strontium	255 ± 7
Manganese	0.214 ± 0.006	Uranium	9.11 ± 0.30
Phosphorus	0.105 ± 0.004		
Potassium	2.17 ± 0.13		
Silicon	31.1 ± 0.4		
Sodium	0.894 ± 0.019		
Titanium	0.311 ± 0.007		
Zinc	0.418 ± 0.015		

<sup>(a)</sup> Certified values for all elements except lead and mercury are the equally weighted means of results from two or three analytical methods. The uncertainty listed with each value is an expanded uncertainty about the mean. The expanded uncertainty is calculated as  $U = k u_c$ , where  $u_c$  is intended to represent, at the level of one standard deviation, the combined effect of between-method and within-method components of uncertainty, following the ISO Guide [3,4]. The coverage factor ( $k$ ) is determined from the Student's  $t$ -distribution corresponding to the appropriate associated degrees of freedom and approximately 95 % confidence for each analyte.

<sup>(b)</sup> The certified values for lead and mercury are each results from a single NIST method (see Table 4) for which a complete evaluation of all sources of uncertainty has been performed. The uncertainties for the certified values for these elements represent expanded uncertainties with a coverage factor of 2, with uncertainty components combined following the ISO Guide [4].

## Appendix F

### NIST SRM 2710a Certificate of Analysis

Table 2. Reference Values <sup>(a,b,c)</sup> (Dry-Mass Basis) for Selected Elements in SRM 2710a

Element	Mass Fraction (mg/kg)
Cesium	8.25 ± 0.11
Chromium	23 ± 6
Europium	0.82 ± 0.01
Gadolinium	3.0 ± 0.1
Lutetium	0.31 ± 0.01
Neodymium	22 ± 2
Nickel	8 ± 1
Rubidium	117 ± 3
Samarium	4.0 ± 0.2
Scandium	9.9 ± 0.1
Thallium	1.52 ± 0.02
Thorium	18.1 ± 0.3
Vanadium	82 ± 9

<sup>(a)</sup> Reference values for all elements except chromium, nickel, samarium, and vanadium are based on results from one analytical method at NIST (see Table 4) and the uncertainties represent the expanded uncertainties, which include the combined Type A and Type B with a coverage factor of 2, following the ISO Guide [4].

<sup>(b)</sup> Reference values for nickel and samarium are the equally weighted means of results from two analytical methods for nickel and two INAA experiments for samarium. The uncertainty listed with each value is an expanded uncertainty about the mean. The expanded uncertainty is calculated as  $U = k u_c$ , where  $u_c$  is intended to represent, at the level of one standard deviation, the combined effect of between-method and within-method components of uncertainty, following the ISO Guide [3,4]. The coverage factor ( $k$ ) is determined from the Student's  $t$ -distribution corresponding to the appropriate associated degrees of freedom and approximately 95 % confidence for each analyte.

<sup>(c)</sup> Reference values for chromium and vanadium are based on a weighted mean calculated based on the Dersimonian-Laird method [5], which incorporates an estimate of the between-method variance into the weights. The expanded uncertainty listed with these values is calculated as  $U = k u_c$ , where  $k = 2$ , and  $u_c$  is intended to represent, at the level of one standard deviation, the combined effect of between-method and within-method components of uncertainty.

## Appendix F

### NIST SRM 2710a Certificate of Analysis

Table 3. Information Values <sup>(a)</sup> (Dry-Mass Basis) for Selected Elements in SRM 2710a

Element	Mass Fraction (mg/kg)
Boron	20
Cerium	60
Dysprosium	3
Gold	0.2
Hafnium	7
Indium	7
Selenium	1
Silver	40
Tantalum	0.9
Terbium	0.5
Tungsten	190
Ytterbium	2
Zirconium	200

<sup>(a)</sup> Information values are based on results from one analytical method at NIST



## Appendix F NIST SRM 2710a Certificate of Analysis

Table 4. NIST Methods Used for the Analysis of SRM 2710a

Element	Methods	Element	Methods
Ag	INAA	Na	INAA; XRF
Al	INAA; XRF	Nd	INAA
As	CCT-ICP-MS; INAA; XRF	Ni	ICP-MS; ICP-OES
Au	INAA	P	ICP-OES; XRF
B	PGAA	Pb	ID-ICP-MS
Ba	INAA; XRF	Rb	INAA
Ca	INAA; XRF	Sb	ICP-MS; INAA
Cd	ID-ICP-MS; PGAA	Sc	INAA
Ce	INAA	Se	CCT-ICP-MS
Co	INAA; ICP-OES	Si	PGAA; XRF
Cr	INAA; XRF	Sm	INAA <sup>(a)</sup>
Cs	INAA	Sr	ICP-OES; XRF
Cu	INAA; XRF	Ta	INAA
Dy	INAA	Tb	INAA
Eu	INAA	Th	INAA
Fe	INAA; PGAA; XRF	Ti	PGAA; XRF
Gd	PGAA	Tl	ICP-MS
Hf	INAA	U	ICP-MS; INAA
Hg	CV-ID-ICPMS	V	INAA; XRF
K	INAA; PGAA; XRF	W	INAA
La	INAA <sup>(a)</sup>	Yb	INAA
Lu	INAA	Zn	INAA; XRF
Mg	INAA; XRF	Zr	XRF
Mn	INAA; PGAA; XRF		

### NIST Methods of Analysis

CCT-ICP-MS	Collision cell inductively coupled plasma mass spectrometry
CV-ID-ICP-MS	Cold vapor isotope dilution inductively coupled plasma mass spectrometry
ICP-MS	Inductively coupled plasma mass spectrometry
ICP-OES	Inductively coupled plasma optical emission spectrometry
ID-ICP-MS	Isotope dilution inductively coupled plasma mass spectrometry
INAA	Instrumental neutron activation analysis
PGAA	Prompt gamma-ray activation analysis
XRF	X-ray fluorescence spectrometry

### USGS Methods of Analysis<sup>(b)</sup>

WD-XRF-2	Wavelength dispersive X-ray fluorescence spectrometry at USGS
ICP-OES-2	Inductively coupled plasma optical emission spectrometry at USGS
ICP-MS-2	Inductively coupled plasma mass spectrometry at USGS

<sup>(a)</sup>Two different INAA experiments, performed using different sub-samples and different analytical conditions, were used to provide certified and reference values for lanthanum and samarium, respectively.

<sup>(b)</sup>USGS Methods of Analysis were used to confirm results from certification methods.

## Appendix F

### NIST SRM 2710a Certificate of Analysis

Table 5. Participating NIST Analysts:

S.J. Christopher	S.A. Rabb
R.D. Day	J.R. Sieber
S.E. Long	R.O. Spatz
E.A. Mackey	R.S. Popelka-Filcoff
A.F. Marlow	B.E. Tomlin
J.L. Molloy	L.J. Wood
K.E. Murphy	L.L. Yu
R.L. Paul	R. Zeisler

Table 6. Participating USGS Laboratory and Analysts

Laboratory	Analysts
U.S. Geological Survey Branch of Geochemistry Denver, CO, USA	M.G. Adams Z.A. Brown P.L. Lamothe J.E. Taggart S.A. Wilson

#### REFERENCES

- [1] Thompson, A.; Taylor, B.N.; *Guide for the Use of the International System of Units (SI)*, NIST Special Publication 811 (2008); available at <http://www.physics.nist.gov/Pubs/contents.html>.
- [2] May, W.E.; Gills, T.E.; Parris, R.; Beck, II, C.M.; Fassett, J.D.; Gettings, R.J.; Greenberg, R.R.; Guenther, F.R.; Kramer, G.; MacDonald, B.S.; Wise, S.A.; *Definitions of Terms and Modes Used at NIST for Value-Assignment of Reference Materials for Chemical Measurements*, NIST Special Publication 260-136 (1999); available at [http://www.cstl.nist.gov/nist839/NIST\\_special\\_publications.htm](http://www.cstl.nist.gov/nist839/NIST_special_publications.htm).
- [3] Levenson, M.S.; Banks, D.L.; Eberhardt, K.R.; Gill, L.M.; Guthrie, W.F.; Liu, H.K.; Vangel, M.G.; Yen, J.H.; Zhang, N.F.; *J. Res. NIST* 105, pp. 571-579 (2000).
- [4] ISO; *Guide to the Expression of Uncertainty in Measurement*, ISBN 92-67-10188-9, 1st ed.; International Organization for Standardization: Geneva, Switzerland (1993); see also Taylor, B.N.; Kuyatt, C.E.; *Guidelines for Evaluating and Expressing the Uncertainty of NIST Measurement Results*, NIST Technical Note 1297, U.S. Government Printing Office, Washington, DC (1994); available at <http://www.physics.nist.gov/Pubs/contents.html>.
- [5] DerSimonian, R.; Laird, N.; *Controlled Clinical Trials* 7, 177-188 (1986).

*Users of this SRM should ensure that the certificate in their possession is current. This can be accomplished by contacting the SRM Program at: telephone (301) 975-2200; fax (301) 926-4751; e-mail [srminfo@nist.gov](mailto:srminfo@nist.gov); or via the Internet at <http://www.nist.gov/srm>.*

Appendix F  
NIST SRM 2710a Certificate of Analysis

Addendum to Certificate  
Standard Reference Material® 2710a

Montana I Soil

Highly Elevated Trace Element Concentrations

Leachable Concentrations Determined Using USEPA Methods 200.7 and 3050B

The mass fraction values contained in the NIST Certificate of Analysis for SRM 2710a represent the total element content of the material. The measurement results used to provide the certified, reference or information values are obtained from methods that require complete sample decomposition, or from nondestructive analytical methods such as instrumental neutron activation analysis or prompt gamma-ray activation analysis. Where complete sample decomposition is required, it can be accomplished by digestion with mixed acids or by fusion. For mixed-acid decomposition, hydrofluoric acid must be included in the acid mixture used to totally decompose siliceous materials such as soils and sediments.

In its monitoring programs, the U.S. Environmental Protection Agency (USEPA) has established a number of leach methods for the preparation of soil samples for the determination of extractable elements. Six laboratories participated, five of which used USEPA Method 200.7; the remaining laboratory used USEPA SW-846 Method 3050B for preparation of soil samples. All elements were determined in leachates by inductively coupled plasma optical emission spectrometry. All laboratories provided individual results from duplicate portions, and these results were averaged together to provide one result for each element from each participating laboratory. Results rejected as outliers by the USEPA Contract Laboratory Program (CLP) officials were not included. Results are summarized in Table A1. The ranges of mass fraction values, median values (to two significant figures), and the number of results included for each are given for 23 elements. The percent recovery values based on the ratios of the median values to the total element content (from the certified, reference, or information values in the Certificate of Analysis) are listed in the last column of Table A1. **Note that the certified values provided as total mass fractions in the Certificate of Analysis are the best estimate of the true mass fraction values for this material.**

This USEPA CLP Study was coordinated by Clifton Jones, Quality Assurance and Technical Support Program (QATS), Shaw Environmental & Infrastructure Group, Las Vegas, NV, under the direction of John Nebelsick, USEPA, Analytical Services Branch. The participating laboratories are listed in Table A2.

## Appendix F NIST SRM 2710a Certificate of Analysis

Table A1. Results from Laboratories Participating in the EPA Contract Laboratory Program Study.

Element	n	Range (mg/kg)	Median (mg/kg)	Recovery (%)
Aluminum	6	8200 - 12000	10000	17
Antimony	6	5.0 - 12	9.6	18
Arsenic	6	1300 - 1600	1400	92
Barium	6	490 - 540	510	65
Beryllium	6	0.24 - 0.51	0.48	—
Cadmium	5	9.6 - 12	11	86
Calcium	6	1700 - 2000	1800	19
Chromium	6	9.2 - 11	10	41
Cobalt	6	2.8 - 5.2	3.8	64
Copper	6	3100 - 3500	3300	95
Iron	6	30000 - 36000	34000	79
Lead	6	4700 - 5800	5100	93
Magnesium	6	3200 - 3600	3500	48
Manganese	6	1500 - 1800	1700	77
Mercury	6	9.3 - 11.7	10	104
Nickel	5	4.8 - 6.1	5.5	69
Potassium	6	3800 - 4700	4100	19
Selenium	2	1.5 - 2.6	2.0	200
Silver	6	31 - 39	36	91
Sodium	6	550 - 650	590	7
Thallium	3	1.3 - 3.6	3.2	213
Vanadium	6	35 - 43	38	48
Zinc	6	3300 - 4400	3800	90

Table A2. List of CLP and non-CLP Participating Laboratories

A4 Scientific, Inc.  
Bonner Analytical Testing Co.  
Chem Tech Consulting Group  
Datachem Laboratories, Inc.  
Liberty Analytical Corporation  
SVL Analytical, Inc.

## **APPENDIX G**

### **Laboratory Submitted Initial Demonstration of Proficiency Forms**

# Laboratory A

**Appendix G  
Laboratory Submitted IDP Forms**

**LAB A Initial Demonstration of Proficiency (IDP) Form  
For IVBA Round Robin of NIST 2710a and 2711a (ver. 06-30-10)**

Before the USEPA initiates the Round Robin analysis of the NIST 2710a and 2711a materials they have requested that each of the laboratories that wish to participate in the study complete the following Initial Demonstration of Proficiency (IDP) Form, Clifton Jones (Quality Assurance Technical Support Laboratory) US (702 895-8713) clifton.jones@shawgrp.com

**General and Facility Questions**

1	<b>Number of IVBA analyses your facility has performed for lead using the attached SOP?</b>	
2	<b>Will your facility conduct the extraction? (Yes/No)</b>	Yes
3	<b>If the answer to question 2 is no, please provide the name of the laboratory that will be conducting the extraction. (Lab Name)</b>	
4	<b>Will your facility conduct the extract analysis? (Yes/No)</b>	No
5	<b>If the answer to question 4 is no, please provide the name of the laboratory that will be conducting the analysis. (Lab Name)</b>	Other lab name was here
6	<b>Will your facility be able to conduct the attached IVBA Method EPA 9200.1-86 as written? (air controlled temperature is OK) (Yes, or Provide comment Below in 7)</b>	No
7	<b>If the answer the question 6 is no, please provide the deviation from the EPA 9200.1-86 method in the field provided here. Comment-</b> The apparatus we use is different from the one described in the SOP. It consists of a locally built Plexiglas/LPDE basket attached to via a pulley to a motor that operates at 30 rpm in an end over end rotation. The basket holds up to ten 125 ml HPDE bottles. The basket containing the bottles is immersed in a water bath maintained at a temperature of $37 \pm 2^{\circ}\text{C}$ . We have been using this apparatus for IVBA determination since 2002 (mostly for arsenic and lead) but have compiled relevant data for lead since 2007. We will use the protocol as written including matrix spikes which we have not included frequently in the past.	

**Apparatus**

8	<b>Does the IVBA apparatus your facility has use air or water as the 37°C thermal conducting/controlling medium. (Air, Water)</b>	Water
9	<b>How many bottle positions does your apparatus have?</b>	10

**Analytical**

10	<b>Type of analytical instrument use for the final Determination (ICP-AES) (ICP-MS) (GFAA) or specify other instrument type.</b>	ICP-MS
11	<b>Please provide the instrumental detection limit for the procedure that you currently use for the IVBA method. (<math>\mu\text{g/L}</math>)</b>	0.1 $\mu\text{g/L}$
12	<b>Name of Control Soil - Reference Material typically used by your facility for the IVBA extraction. ( e.g., NIST 2710 or 2711, or other)</b>	NIST 2711
13	<b>Blank spike amount (mg/L) used in your procedure.</b>	10 mg/L
14	<b>Matrix spike amount (mg/L) used in your procedure.</b>	10 mg/L

**Appendix G**  
**Laboratory Submitted IDP Forms**

**Analytical (continued)**

**Table of Batch IVBA Results**

No	Batch Date	Reagent Blank µg/L	Bottle Blank µg/L	Spiked Blank Result	Spike Blank Percent Recovery	Matrix Spike Percent Recovery	Duplicate Relative Percent Difference	Reference Material Name	Control Soil Result (mg/L) (analytical solution)	Control Soil RPD	Control Soil IVBA
<b>A</b>	<b>Date</b>	<b>&lt;25 µg/L</b>	<b>&lt;50 µg/L</b>	<b>(mg/L)</b>	<b>85-115%</b>	<b>75-125%</b>	<b>&lt;20%RPD</b>			<b>&lt;10%RPD</b>	<b>IVBA%</b>
<b>B</b>	<b>mm/dd/yyyy</b>	<b>&lt;25 µg/L</b>	<b>&lt;50 µg/L</b>	<b>9.2</b>	<b>92.4%</b>	<b>87.3%</b>	<b>7.4%</b>	<b>NIST 2711</b>	<b>9.12 mg/L</b>	<b>7.1%</b>	<b>75.5%</b>
1	01/15/2007	<50	<50	9.5	95.0	N/A	11.9	NIST 2711	10.1	9.54	84.6
2	10/02/2007	<50	<50	9.5	95.0		0.0	NIST 2711	9.42	2.17	81.4
3	11/26/2007	<50	<50	9.5	94.7		9.5	NIST 2711	9.85	6.83	84.8
4	11/28/2007	<50	<50	9.3	93.3		2.4	NIST 2711	9.59	4.01	82.5
5	12/03/2007	<50	<50	9.4	94.1		27.0	NIST 2711	9.44	2.39	81.2
6	12/04/2007	<50	<50	9.5	94.7		6.9	NIST 2711	9.75	5.75	83.9
7	12/05/2007	<50	<50	9.5	95.0		31.6	NIST 2711	9.85	6.83	84.8
8	12/05/2007	<50	<50	9.5	95.2		14	NIST 2711	10.0	8.46	86.1
9	12/05/2007	<50	<50	9.5	95.2		5.1	NIST 2711	10.1	9.54	86.9
10	12/09/2007	<50	<50	9.8	97.8		10.0	NIST 2711	9.42	2.17	81.1
11	03/25/2008	<0.1	1.1	9.3	93.0		6.2	NIST 2711	10.2	10.6	89.2
12	03/28/2008	<0.1	0.7	10.5	105		2.3	NIST 2711	10.7	16.1	90.6
13	05/08/2008	<0.1	0.7	9.8	98		0.4	NIST 2711	10.9	18.2	91.5
14	05/08/2008	<0.1	0.8	10.2	102		2.7	NIST 2711	10.1	9.54	86.8
15	11/26/2008	<0.1	4.2	9.8	98		2.8	NIST 2711	9.79	6.18	84.3
16	11/27/2008	<0.1	1.0	10.5	105		4.2	NIST 2711	10.4	12.8	89.5
17	02/03/2009	<0.1	3.5	9.9	99		3.6	NIST 2711	9.50	3.04	80.6
18	02/04/2009	<0.1	0.9	9.9	99		7.7	NIST 2711	9.77	5.97	82.6
19	02/05/2009	<0.1	0.5	10.5	105		2.3	NIST 2711	8.97	-2.71	75.7
20	02/05/2009	<0.1	0.5	9.9	99		1.5	NIST 2711	9.34	1.30	79.3
21	02/05/2009	<0.1	0.4	9.9	99		1.5	NIST 2711	8.97	-2.71	75.7
22	02/10/2009	<0.1	12.5	11.1	111		14.5	NIST 2711	10.0	8.46	84.4
23	02/11/2009	<0.1	0.2	10.3	103		1.6	NIST 2711	10.1	9.54	86.2
24	02/12/2009	<0.1	0.2	10.2	102		2.2	NIST 2711	10.2	10.6	87.1
25	02/12/2009	<50	<50	10.3	103		2.9	NIST 2711	9.34	1.30	78.4



**Appendix G**  
**Laboratory Submitted IDP Forms**

26	10/05/2009	<50	<50	9.5	95.0		4.5	NIST 2711	10.6	15.0	91.6
27	11/23/2009	<0.1	<0.5	10.3	103		11.9	NIST 2711	10.4	12.8	88.6
28	01/12/2010	<0.1	2.3	10.7	107		0.1	NIST 2711	10.40	12.8	88.2
29	01/13/2010	<0.1	1.0	10.8	108		0.9	NIST 2711	10.80	17.1	91.2
30	02/09/2010	<0.1	0.6	10.7	107		1.7	NIST 2711	8.10	-12.1	69.7
31	02/09/2010	<0.1	0.6	10.3	103		8.0	NIST 2711	9.57	3.80	82.4
32	02/13/2010	<0.2	<0.2	10.3	103		9.2	NIST 2711	10.0	8.46	86.1
33	02/15/2010	<5	<5	9.5	95.0		12.2	NIST 2711	10.4	12.8	86.7
34	02/15/2010	<5	<5	9.8	98.2		0.0	NIST 2711	10.0	8.46	85.3
35	02/17/2010	<0.2	0.5	10.4	104		2.4	NIST 2711	9.19	-0.33	78.6
36	02/19/2010	<0.1	0.4	10.3	103		2.3	NIST 2711	9.82	6.51	82.1
37	02/23/2010	<0.1	0.3	10.5	105		2.0	NIST 2711	9.57	3.80	80.1
38	02/23/2010	<0.1	1.4	10.2	102		2.6	NIST 2711	9.66	4.77	80.1
39	02/24/2010	<0.1	0.6	10.2	102		0.1	NIST 2711	9.24	0.22	79.5
40	03/01/2010	<0.1	0.6	10.2	102		1.2	NIST 2711	9.52	3.25	80.8
41	03/02/2010	<0.1	0.4	10.3	103		1.9	NIST 2711	9.36	1.52	80.3
42	03/03/2010	<0.1	1.1	10.4	104		8.5	NIST 2711	9.76	5.86	81.0
43	03/04/2010	<0.1	0.3	10.3	103		3.5	NIST 2711	9.48	2.82	81.0
44	03/05/2010	<0.1	0.3	10.4	104		2.2	NIST 2711	9.74	5.64	82.9
45	03/09/2010	<0.1	0.8	10.4	104		79	NIST 2711	9.96	8.03	82.4
46	03/09/2010	<0.1	0.5	10.5	105		0.7	NIST 2711	9.84	6.72	83.1

**Note Row A presents the quality control acceptance criteria from the USEPA IVBA Method EPA 9200.1-86, and Row B provides an example.**

# Laboratory B

**Appendix G  
Laboratory Submitted IDP Forms**

**Lab B Initial Demonstration of Proficiency (IDP) Form  
For IVBA Round Robin of NIST 2710a and 2711a (ver. 06-30-10)  
(submitted 7-08-2010)**

Before the USEPA initiates the Round Robin analysis of the NIST 2710a and 2711a materials they have requested that each of the laboratories that wish to participate in the study complete the following Initial Demonstration of Proficiency (IDP) Form, Clifton Jones (Quality Assurance Technical Support Laboratory) US (702 895-8713) clifton.jones@shawgrp.com

**General and Facility Questions**

1	Number of IVBA analyses your facility has performed for lead using the attached SOP?	~ 50 for Pb (> 150 for As)
2	Will your facility conduct the extraction? (Yes/No)	Yes
3	If the answer to question 2 is no, please provide the name of the laboratory that will be conducting the extraction. (Lab Name)	
4	Will your facility conduct the extract analysis? (Yes/No)	No
5	If the answer to question 4 is no, please provide the name of the laboratory that will be conducting the analysis. (Lab Name)	Other lab name was here.
6	Will your facility be able to conduct the attached IVBA Method EPA 9200.1-86 as written? (air controlled temperature is OK) (Yes, or Provide comment Below in 7)	Yes
7	If the answer the question 6 is no, please provide the deviation from the EPA 9200.1-86 method in the field provided here. Comment-	

**Apparatus**

8	Does the IVBA apparatus your facility has use air or water as the 37°C thermal conducting/controlling medium. (Air, Water)	water
9	How many bottle positions does your apparatus have?	10

**Analytical**

10	Type of analytical instrument use for the final Determination (ICP-AES) (ICP-MS) (GFAA) or specify other instrument type	ICP-MS
11	Please provide the instrumental detection limit for the procedure that you currently use for the IVBA method. (□g/L)	0.106 □g/L
12	Name of Control Soil - Reference Material typically used by your facility for the IVBA extraction. ( e.g., NIST 2710 or 2711, or other)	NIST 2711
13	Blank spike amount (mg/L) used in your procedure.	10.0 mg/L
14	Matrix spike amount (mg/L) used in your procedure.	n.a.

**Appendix G  
Laboratory Submitted IDP Forms**

**Analytical (continued)**

**Table of Batch IVBA Results**

No	Batch Date	Reagent Blank µg/L	Bottle Blank µg/L	Spiked Blank Result	Spike Blank Percent Recovery	Matrix Spike Percent Recovery	Duplicate Relative Percent Difference	Reference Material Name	Control Soil Result (mg/L) (analytical solution)	Control Soil RPD	Control Soil IVBA
<b>A</b>	<b>Date</b>	<b>&lt;25 µg/L</b>	<b>&lt;50 µg/L</b>	<b>(mg/L)</b>	<b>85-115%</b>	<b>75-125%</b>	<b>&lt;20%RPD</b>			<b>&lt;10%RPD</b>	<b>IVBA%</b>
<b>B</b>	<b>mm/dd/yyyy</b>	<b>&lt;25 µg/L</b>	<b>&lt;50 µg/L</b>	<b>9.2</b>	<b>92%</b>	<b>87%</b>	<b>7%</b>	<b>NIST 2711</b>	<b>9.12 mg/L</b>	<b>7.1%</b>	<b>75.5%</b>
1	4/26/2005	n.m.	< 5	9.6	96%	n.m.	n.m.	NIST 2711	11	n.m.	95%^
2	8/22/2005	< 5	< 5	1.0*	100	n.m.	0	NIST 2711	12	n.m.	103%^
3	8/30/2005	n.m.	< 5	11	110	n.m.	10%	NIST 2711	10, 10, 10, 11**	10%	86%^
4	9/1/2005	n.m.	< 5	8.9	89	n.m.	3%	NIST 2711	9.6, 9.5, 9.8, 9.6**	3%	83%^
5	9/12/2005	n.m.	< 5	11	110	n.m.	1%	NIST 2711	10,10, 9.9, 10**	1%	86%^
6	9/19/2005	n.m.	< 5	11	110	n.m.	9.5%	NIST 2711	10, 10, 11, 11**	9.5%	91%^
7	9/21/06	< 5	8	11	110	n.m.	n.m.	NIST 2711	9.5	n.m.	82%^
8	9/22/2006	< 5	9	11	110	n.m.	n.m.	NIST 2711	15	n.m.	130%^
9	8/22/2008	< 5	< 5	11	110	n.m.	0	NIST 2711	10	n.m.	86%^
10											

\* Spiked to 1.0 mg/L Pb.

\*\* NIST soil extracted 4 times during this data set.

^ Assumes concentration of lead in NIST 2711 soil is 1162 mg/kg, per certificate of analysis.

Note Row A presents the quality control acceptance criteria from the USEPA IVBA Method EPA 9200.1-86, and Row B provides an example.

# Laboratory C

**Appendix G  
Laboratory Submitted IDP Forms**

**LAB C Initial Demonstration of Proficiency (IDP) Form  
For IVBA Round Robin of NIST 2710a and 2711a (ver. 07-02-10)  
(Submitted 7-26-2010)**

Before the USEPA initiates the Round Robin analysis of the NIST 2710a and 2711a materials they have requested that each of the laboratories that wish to participate in the study complete the following Initial Demonstration of Proficiency (IDP) Form, Clifton Jones (Quality Assurance Technical Support Laboratory) US (702 895-8713) clifton.jones@shawgrp.com

**General and Facility Questions**

1	<b>Number of IVBA analyses your facility has performed for lead using the attached SOP?</b>	1,926 (MS Access data base query, includes QC)
2	<b>Will your facility conduct the extraction? (Yes/No)</b>	yes
3	<b>If the answer to question 2 is no, please provide the name of the laboratory that will be conducting the extraction. (Lab Name)</b>	
4	<b>Will your facility conduct the extract analysis? (Yes/No)</b>	yes
5	<b>If the answer to question 4 is no, please provide the name of the laboratory that will be conducting the analysis. (Lab Name)</b>	
6	<b>Will your facility be able to conduct the attached IVBA Method EPA 9200.1-86 as written? (air controlled temperature is OK) (Yes, or Provide comment Below in 7)</b>	Yes
7	<b>If the answer the question 6 is no, please provide the deviation from the EPA 9200.1-86 method in the field provided here. Comment-</b>	

**Apparatus**

8	<b>Does the IVBA apparatus your facility has use air or water as the 37°C thermal conducting/controlling medium. (Air, Water)</b>	Water
9	<b>How many bottle positions does your apparatus have?</b>	10

**Analytical**

10	<b>Type of analytical instrument use for the final Determination (ICP-AES) (ICP-MS) (GFAA) or specify other instrument type.</b>	ICP-AES or ICP-MS (We have both)
11	<b>Please provide the aqueous method detection limit for the procedure that you currently use for the IVBA method. (□g/L)</b>	ICP 40 ug/L & ICP-MS 0.1 ug
12	<b>Name of Control Soil - Reference Material typically used by your facility for the IVBA extraction. ( e.g., NIST 2710 or 2711, or other)</b>	NIST 2711
13	<b>Blank spike amount (mg/L) used in your procedure.</b>	High 10 mg/L Low is 1 mg/L
14	<b>Matrix spike amount (mg/L) used in your procedure.</b>	High 10 mg/L Low is 1 mg/L

**Appendix G  
Laboratory Submitted IDP Forms**

**Analytical (continued)**

**Table of Batch IVBA Results**

No	Batch Date	Reagent Blank µg/L	Bottle Blank µg/L	Spiked Blank Result	Spike Blank Percent Recovery	Matrix Spike Percent Recovery	Duplicate Relative Percent Difference	Reference Material Name	Control Soil Result (mg/L) (analytical solution)	Control Soil RPD	Control Soil IVBA
<b>A</b>	<b>Date</b>	<b>&lt;25 µg/L</b>	<b>&lt;50 µg/L</b>	<b>(mg/L)</b>	<b>85-115%</b>	<b>75-125%</b>	<b>&lt;20%RPD</b>			<b>&lt;10%RPD</b>	<b>IVBA%</b>
<b>B</b>	<b>mm/dd/yyyy</b>	<b>&lt;25 µg/L</b>	<b>&lt;50 µg/L</b>	<b>9.2</b>	<b>92.4%</b>	<b>87.3%</b>	<b>7.4%</b>	<b>NIST 2711</b>	<b>9.12 mg/L</b>	<b>7.1%</b>	<b>75.5%</b>
1	06/04/2009	<25ug/L	<40 ug/L	10.42	104.3	121.8	2.2	NIST 2711	9.48	2.4	82
2	06/29-2008	<25ug/L	<40 ug/L	9.62	96.2	92.5	0.6	NIST 2711	9.13	0.4	79
3	06/28/2008	<25ug/L	<40 ug/L	9.69	96.9	95.7	3.2	NIST 2711	9.36	0.1	81
4	02//05/2008	<25ug/L	<40 ug/L	9.81	98.1	84.2	0.8	NIST 2711	9.47	2.6	81
5	02/07/2008	<25ug/L	<40 ug/L	9.94	99.4	85.5	0.2	NIST 2711	8.21	2.6	71
6	02/07/2008	<25ug/L	<40 ug/L	9.53	95.3	89.2	0.1	NIST 2711	9.20	2.5	79
7	02/07/2008	<25ug/L	<40 ug/L	9.43	94.3	89.00	1.8	NIST 2711	9.11	0.6	78
8	04/24/2008	<25ug/L	<40 ug/L	9.89	98.9	92.3	1.1	NIST 2711	9.66	2.2	83
9	05/16/2008	<25ug/L	<40 ug/L	9.43	94.3	Lab C-M3 FLAG* SEE Below	0.7	NIST 2711	9.10	0.8	78
10	08/08/2009	<25ug/L	<40 ug/L	9.28	92.8	Lab C-M3 FLAG* SEE Below	2.5	NIST 2711	8.92	2.7	77

Note Row A presents the quality control acceptance criteria from the USEPA IVBA Method EPA 9200.1-86, and Row B provides an example.

M3 Flag on Lab -X's reports. M3 = The Spike Recovery value is unusable since the analyte concentration in the sample was disproportionate to the spike level. The recovery of associated control samples (LFB & LCS) was acceptable. In this case the samples were so high in Pb the spike values were unusable

Control Soil IVBA % were based on TV of 1162, which is the value used by the EPA in the 2007b validation document, (Drexler and Brattin 2007: EPA 2007b)

# Laboratory D



**Appendix G  
Laboratory Submitted IDP Forms**

**LAB D Initial Demonstration of Proficiency (IDP) Form  
For IVBA Round Robin of NIST 2710a and 2711a (ver. 07-02-10)  
(Submitted 7-21-2010)**

Before the USEPA initiates the Round Robin analysis of the NIST 2710a and 2711a materials they have requested that each of the laboratories that wish to participate in the study complete the following Initial Demonstration of Proficiency (IDP) Form, Clifton Jones (Quality Assurance Technical Support Laboratory) US (702 895-8713) clifton.jones@shawgrp.com

**General and Facility Questions**

1	Number of IVBA analyses your facility has performed for lead using the attached SOP?	~9-10,000
2	Will your facility conduct the extraction? (Yes/No)	Yes
3	If the answer to question 2 is no, please provide the name of the laboratory that will be conducting the extraction. (Lab Name)	
4	Will your facility conduct the extract analysis? (Yes/No)	Yes
5	If the answer to question 4 is no, please provide the name of the laboratory that will be conducting the analysis. (Lab Name)	
6	Will your facility be able to conduct the attached IVBA Method EPA 9200.1-86 as written? (air controlled temperature is OK) (Yes, or Provide comment Below in 7)	Yes
7	If the answer the question 6 is no, please provide the deviation from the EPA 9200.1-86 method in the field provided here. Comment-	

**Apparatus**

8	Does the IVBA apparatus your facility has use air or water as the 37°C thermal conducting/controlling medium. (Air, Water)	Either
9	How many bottle positions does your apparatus have?	We have two 10 position

**Analytical**

10	Type of analytical instrument use for the final Determination (ICP-AES) (ICP-MS) (GFAA) or specify other instrument type.	ICP/MS
11	Please provide the aqueous method detection limit for the procedure that you currently use for the IVBA method. (□g/L)	.02 ug/l
12	Name of Control Soil - Reference Material typically used by your facility for the IVBA extraction. ( e.g., NIST 2710 or 2711, or other)	NIST 2710 , 2711, or 2710A
13	Blank spike amount (mg/L) used in your procedure.	1 mg/l
14	Matrix spike amount (mg/L) used in your procedure.	1 mg/l

**Appendix G  
Laboratory Submitted IDP Forms**

**Analytical (continued)**

**Table of Batch IVBA Results**

No	Batch Date	Reagent Blank µg/L	Bottle Blank µg/L	Spiked Blank Result	Spike Blank Percent Recovery	Matrix Spike Percent Recovery	Duplicate Relative Percent Difference	Reference Material Name	Control Soil Result (mg/L) (analytical solution)	Control Soil RPD	Control Soil IVBA
<b>A</b>	<b>Date</b>	<b>&lt;25 µg/L</b>	<b>&lt;50 µg/L</b>	<b>(mg/L)</b>	<b>85-115%</b>	<b>75-125%</b>	<b>&lt;20%RPD</b>			<b>&lt;10%RPD</b>	<b>IVBA%</b>
<b>B</b>	<b>mm/dd/yyyy</b>	<b>&lt;25 µg/L</b>	<b>&lt;50 µg/L</b>	<b>9.2</b>	<b>92.4%</b>	<b>87.3%</b>	<b>7.4%</b>	<b>NIST 2711</b>	<b>9.12 mg/L</b>	<b>7.1%</b>	<b>75.5%</b>
1	05/19/10	0.05	0.05	2603	100	112	6				
2	03/19/10	-.09	-.09	2669	107	99	3				
3	03/07/10	-.08	-.08	2789	111	108	9				
4	02/03/10	.07	.07	2658	106	107	34	2711	0.611	2.2	
5	12/03/09	.23	.23	2744	110	102	6				
6	12/02/09	.1	.1	2614	105	102	16	2711	0.567	8.9	
7	11/09/09	.17	.17	2497	100	94	23				
8	12.03/09	.08	.08	2667	107	93	3				
9	12/04/09	.1	.1	2737	109	101	12				
10	12/01/09	-.04	-.04	2615	105	102	1				

Note Row A presents the quality control acceptance criteria from the USEPA IVBA Method EPA 9200.1-86, and Row B provides an example.

# Laboratory E

## Appendix G Laboratory Submitted IDP Forms

### Lab E Initial Demonstration of Proficiency (IDP) Form For IVBA Round Robin of NIST 2710a and 2711a (ver. 07-02-10)

Before the USEPA initiates the Round Robin analysis of the NIST 2710a and 2711a materials they have requested that each of the laboratories that wish to participate in the study complete the following Initial Demonstration of Proficiency (IDP) Form, Clifton Jones (Quality Assurance Technical Support Laboratory) US (702 895-8713) [clifton.jones@shawgrp.com](mailto:clifton.jones@shawgrp.com)

#### General and Facility Questions

1	Number of IVBA analyses your facility has performed for lead using the attached SOP?	~ 420 analyses
2	Will your facility conduct the extraction? (Yes/No)	Yes
3	If the answer to question 2 is no, please provide the name of the laboratory that will be conducting the extraction. (Lab Name)	
4	Will your facility conduct the extract analysis? (Yes/No)	Yes
5	If the answer to question 4 is no, please provide the name of the laboratory that will be conducting the analysis. (Lab Name)	
6	Will your facility be able to conduct the attached IVBA Method EPA 9200.1-86 as written? (air controlled temperature is OK) (Yes, or Provide comment Below in 7)	Yes
7	If the answer the question 6 is no, please provide the deviation from the EPA 9200.1-86 method in the field provided here. Comment-	

#### Apparatus

8	Does the IVBA apparatus your facility has use air or water as the 37°C thermal conducting/controlling medium. (Air, Water)	Water
9	How many bottle positions does your apparatus have?	12

#### Analytical

10	Type of analytical instrument use for the final Determination (ICP-AES) (ICP-MS) (GFAA) or specify other instrument type.	ICP-MS
11	Please provide the aqueous method detection limit for the procedure that you currently use for the IVBA method. (µg/L)	0.08 µg/L
12	Name of Control Soil - Reference Material typically used by your facility for the IVBA extraction. ( e.g., NIST 2710 or 2711, or other)	2710 (used through 2/10/09 when we ran out of this SRM)
13	Blank spike amount (mg/L) used in your procedure.	10 mg/L
14	Matrix spike amount (mg/L) used in your procedure.	10 mg/L

Appendix G  
Laboratory Submitted IDP Forms

Analytical (continued)

**Table of Batch IVBA Results for Pb**

No	Batch Date	Reagent Blank µg/L	Bottle Blank µg/L	Spiked Blank Result	Spike Blank Percent Recovery	Matrix Spike Percent Recovery	Duplicate Relative Percent Difference	Reference Material Name	Control Soil Result (mg/L) (analytical solution)	Control Soil RPD	Control Soil IVBA
<b>A</b>	Date	<25 µg/L	<50 µg/L	(mg/L)	85-115%	75-125%	<20%RPD			<10%RPD	IVBA%
<b>B</b>	mm/dd/yyyy	<25 µg/L	<50 µg/L	9.2	92.4%	87.3%	7.4%	NIST 2711	9.12 mg/L	7.1%	75.5%
1	2/4/09	<5 µg/L	<5 µg/L	9.9	99.5	100	0.0	NIST 2710	40.8	1.9	73.6
2	2/4/09	<5 µg/L	<5 µg/L	10.2	101.8	99.0	1.0	NIST 2710	40.7	1.2	74.1
3	2/5/09	<5 µg/L	<5 µg/L	10.2	101.6	105	4.6	NIST 2710	46.3	6.0	79.5
4	2/5/09	<5 µg/L	<5 µg/L	10.2	102.3	103	3.2	NIST 2710	NA	NA	NA
5	2/9/09	<5 µg/L	<5 µg/L	10.0	100.5	NA	NA	NIST 2710	NA	NA	NA
6	2/10/09	<5 µg/L	<5 µg/L	10.1	101.4	99.3	0.7	NIST 2710	42.3	0.4	75.3
7	2/10/09	<5 µg/L	<5 µg/L	NA	NA	96.6	3.4	NIST 2710	43.5	3.45	77.6
8											
9											
10											

Note Row A presents the quality control acceptance criteria from the USEPA IVBA Method EPA 9200.1-86, and Row B provides an example.

# Laboratory F

**Appendix G**  
**Laboratory Submitted IDP Forms**

**LAB F Initial Demonstration of Proficiency (IDP) Form**  
**For IVBA Round Robin of NIST 2710a and 2711a** (ver. 07-02-10)

(Submitted 7-13-2010)

Before the USEPA initiates the Round Robin analysis of the NIST 2710a and 2711a materials they have requested that each of the laboratories that wish to participate in the study complete the following Initial Demonstration of Proficiency (IDP) Form, Clifton Jones (Quality Assurance Technical Support Laboratory) US (702 895-8713) clifton.jones@shawgrp.com

**General and Facility Questions**

1	<b>Number of IVBA analyses your facility has performed for lead using the attached SOP?</b>	60
2	<b>Will your facility conduct the extraction? (Yes/No)</b>	Yes
3	<b>If the answer to question 2 is no, please provide the name of the laboratory that will be conducting the extraction. (Lab Name)</b>	
4	<b>Will your facility conduct the extract analysis? (Yes/No)</b>	Yes
5	<b>If the answer to question 4 is no, please provide the name of the laboratory that will be conducting the analysis. (Lab Name)</b>	
6	<b>Will your facility be able to conduct the attached IVBA Method EPA 9200.1-86 as written? (air controlled temperature is OK) (Yes, or Provide comment Below in 7)</b>	Yes, air controlled
7	<b>If the answer the question 6 is no, please provide the deviation from the EPA 9200.1-86 method in the field provided here. Comment-</b>	

**Apparatus**

8	<b>Does the IVBA apparatus your facility has use air or water as the 37°C thermal conducting/controlling medium. (Air, Water)</b>	Air
9	<b>How many bottle positions does your apparatus have?</b>	12

**Analytical**

10	<b>Type of analytical instrument use for the final Determination (ICP-AES) (ICP-MS) (GFAA) or specify other instrument type.</b>	ICP-AES
11	<b>Please provide the aqueous method detection limit for the procedure that you currently use for the IVBA method. (□g/L)</b>	25 ug/L
12	<b>Name of Control Soil - Reference Material typically used by your facility for the IVBA extraction. ( e.g., NIST 2710 or 2711, or other)</b>	2711
13	<b>Blank spike amount (mg/L) used in your procedure.</b>	10 mg/L
14	<b>Matrix spike amount (mg/L) used in your procedure.</b>	10 mg/L

**Appendix G  
Laboratory Submitted IDP Forms**

**Analytical (continued)**

**Table of Batch IVBA Results**

No	Batch Date	Reagent Blank ug/L	Bottle Blank ug/L	Spiked Blank Result	Spike Blank Percent Recovery	Matrix Spike Percent Recovery	Duplicate Relative Percent Difference	Reference Material Name	Control Soil Result (mg/L) (analytical solution)	Control Soil RPD	Control Soil IVBA
<b>A</b>	<b>Date</b>	<b>&lt;25 ug/L</b>	<b>&lt;50 ug/L</b>	<b>(mg/L)</b>	<b>85-115%</b>	<b>75-125%</b>	<b>&lt;20%RPD</b>			<b>&lt;10%RPD</b>	<b>IVBA%</b>
<b>B</b>	<b>mm/dd/yyyy</b>	<b>&lt;25 ug/L</b>	<b>&lt;50 ug/L</b>	<b>9.2</b>	<b>92.4%</b>	<b>87.3%</b>	<b>7.4%</b>	<b>NIST 2711</b>	<b>9.12 mg/L</b>	<b>7.1%</b>	<b>75.5%</b>
1	06/16/2010	<25 ug/L	NA	9.6	95.5%	92.3	2.8	NIST 2711	9.36 mg/L	0.7%	84.4%
2	06/28/2010	<25 ug/L	NA	9.6	95.9%	91.6	1.8	NIST 2711	9.20 mg/L	-0.8%	84.4%
3	06/30/2010	<25 ug/L	NA	9.6	96.5%	96.0	2.2	NIST 2711	9.42 mg/L	1.2%	84.4%
4	07/06/2010	<25 ug/L	NA	9.5	94.8%	94.2	3.1	NIST 2711	9.31 mg/L	0.2%	84.4%
5	07/07/2010	<25 ug/L	NA	9.5	94.8%	89.1	1.2	NIST 2711	9.19 mg/L	-0.8%	84.4%
6											
7											
8											
9											
10											

Note Row A presents the quality control acceptance criteria from the USEPA IVBA Method EPA 9200.1-86, and Row B provides an example.

*Note (LAB F): 75.5% IVBA listed in example should be for NIST 2710 according to USEPA IVBA Method EPA 9200.*



# Laboratory G

**Appendix G  
Laboratory Submitted IDP Forms**

**LAB G Demonstration of Proficiency (IDP) Form  
For IVBA Round Robin of NIST 2710a and 2711a (ver. 07-02-10)**

Before the USEPA initiates the Round Robin analysis of the NIST 2710a and 2711a materials they have requested that each of the laboratories that wish to participate in the study complete the following Initial Demonstration of Proficiency (IDP) Form, Clifton Jones (Quality Assurance Technical Support Laboratory) US (702 895-8713) clifton.jones@shawgrp.com

**General and Facility Questions**

1	Number of IVBA analyses your facility has performed for lead using the attached SOP?	228
2	Will your facility conduct the extraction? (Yes/No)	Yes
3	If the answer to question 2 is no, please provide the name of the laboratory that will be conducting the extraction. (Lab Name)	
4	Will your facility conduct the extract analysis? (Yes/No)	Yes
5	If the answer to question 4 is no, please provide the name of the laboratory that will be conducting the analysis. (Lab Name)	
6	Will your facility be able to conduct the attached IVBA Method EPA 9200.1-86 as written? (air controlled temperature is OK) (Yes, or Provide comment Below in 7)	Yes. * However, we do not have riffle splitter to mix and split the samples. We use in air incubator set at 37 C.
7	If the answer the question 6 is no, please provide the deviation from the EPA 9200.1-86 method in the field provided here. Comment- 1. Per method comparison, We normally dry our samples at 105 deg. Celsius instead recommended <40 deg. Celsius per item # 6. May need some clarification. 2. Cost of splitter is \$500- recommended but not required per specified method.	

**Apparatus**

8	Does the IVBA apparatus your facility has use air or water as the 37°C thermal conducting/controlling medium. (Air, Water)	Air
9	How many bottle positions does your apparatus have?	8 per each

**Analytical**

10	Type of analytical instrument use for the final Determination (ICP-AES) (ICP-MS) (GFAA) or specify other instrument type.	ICP-AES
11	Please provide the aqueous method detection limit for the procedure that you currently use for the IVBA method. (□g/L)	50 □g/L
12	Name of Control Soil - Reference Material typically used by your facility for the IVBA extraction. ( e.g., NIST 2710 or 2711, or other)	2711
13	Blank spike amount (mg/L) used in your procedure.	100 □g/L
14	Matrix spike amount (mg/L) used in your procedure.	100 □g/L

**Appendix G  
Laboratory Submitted IDP Forms**

**Analytical (continued)**

**Table of Batch IVBA Results**

No	Batch Date	Reagent Blank µg/L	Bottle Blank µg/L	Spiked Blank Result	Spike Blank Percent Recovery	Matrix Spike Percent Recovery	Duplicate Relative Percent Difference	Reference Material Name	Control Soil Result (mg/L) (analytical solution)	Control Soil RPD	Control Soil IVBA
<b>A</b>	<b>Date</b>	<25 µg/L	<50 µg/L	<b>(mg/L)</b>	<b>85-115%</b>	<b>75-125%</b>	<b>&lt;20%RPD</b>			<b>&lt;10%RPD</b>	<b>IVBA%</b>
<b>B</b>	<b>mm/dd/yyyy</b>	<25 µg/L	<50 µg/L	<b>9.2</b>	<b>92.4%</b>	<b>87.3%</b>	<b>7.4%</b>	<b>NIST 2711</b>	<b>9.12 mg/L</b>	<b>7.1%</b>	<b>75.5%</b>
<b>1</b>	06/14/2010 (1)		<50 ug/L	4.37	83.2%	-132, -266	2.3, 4.6, 10.4, 4.8, 4.1	NIST 2711	8.39 mg/L		77.7%
<b>2</b>	06/14/2010 (2)		<50 ug/L	4.36	83.0%	80.1, 76.7	2.6, 6.6, 6.3, 0.9	NIST 2711	8.89 mg/L		80.8%
<b>3</b>	06/14/2010 (3)		<50 ug/L	4.35	82.9%	81.3, 72.9	1.6	NIST 2711	8.51 mg/L		76.7%
<b>4</b>	01/28/2010	<50 ug/L		4.21	84.2%	406, 403	0.6, 4.4, 2.1	NIST 2711	8.78 mg/L		85.0%
<b>5</b>	12/14/2009			0.745	74.5%	596, 287		NIST 2711	2.12 mg/L		67.5%
<b>6</b>	08/27/2009		<50 ug/L	3.75	75.0%	76.6, 86.5	6.3, 9.2, 5.8, 8.6	NIST 2711	8.10 mg/L		74.3%
<b>7</b>	06/30/2009		<50 ug/L	3.88	77.6%	73.7, 71.5	86.0, 0.9, 0.5, 4.8	NIST 2711	8.78 mg/L		81.3%
<b>8</b>	06/25/2009		<50 ug/L	4.16	83.2%	69.4, 57.0	70.2, 16.3, 1.0	NIST 2711	8.50 mg/L		78.0%
<b>9</b>	06/02/2009		<50 ug/L	4.02	80.4%	77.0, 91.0	3.2, 3.0, 6.5	unknown	8.20 mg/L		74.5%
<b>10</b>	05/26/2009		<50 ug/L			25.5, 38.9	16.4, 0.8, 8.8, 0.4, 18.8	unknown	5.70 mg/L		52.8%

**Note Row A presents the quality control acceptance criteria from the USEPA IVBA Method EPA 9200.1-86, and Row B provides an example.**

Data Notes: All batches had a matrix spike and matrix spike duplicate. Most batches had duplicate analyses on multiple samples.  
 Row 1 MS/MSD were spiked at <10% of native concentration.  
 Row 4 MS/MSD were spiked at ~30% of native concentration.  
 Row 5 MS/MSD were spiked at <15% of native concentration.  
 Row 10 MS/MSD were spiked at ~15% of native concentration.

# Laboratory H

**Appendix G  
Laboratory Submitted IDP Forms**

**LAB H Demonstration of Proficiency (IDP) Form  
For Lead IVBA Round Robin of new RM, with Microwave Digestion of  
RM for Lead and Arsenic using EPA Method 3051A (ver. 09-22-11)**

Before the USEPA initiates the Round Robin analysis the new RM they have requested that each of the laboratories that wish to participate in the study complete the following Initial Demonstration of Proficiency (IDP) Form, Clifton Jones (Quality Assurance Technical Support Laboratory) US (702 895-8713) clifton.jones@shawgrp.com

**General and Facility Questions**

<b>IVBA</b>		
<b>1</b>	<b>Number of IVBA analyses your facility has performed for lead using the attached IVBA SOP EPA 9200.1-86?</b>	<b>* See below</b>
<b>2</b>	<b>Will your facility conduct the extraction? (Yes/No)</b>	<b>Yes</b>
<b>3</b>	<b>If the answer to question 2 is no, please provide the name of the laboratory that will be conducting the Extraction. (Lab Name)</b>	
<b>4</b>	<b>Will your facility conduct the extract analysis? (Yes/No)</b>	<b>Yes</b>
<b>5</b>	<b>If the answer to question 4 is no, please provide the name of the laboratory that will be conducting the Analysis. (Lab Name)</b>	
<b>6</b>	<b>Will your facility be able to conduct the attached IVBA Method EPA 9200.1-86 as written? (air controlled temperature is OK) (Yes, or Provide comment Below in 7)</b>	<b>Yes, see LAB H Lab SOP 256.</b>
<b>7</b>	<b>If the answer the question 6 is no, please provide the deviation from the EPA 9200.1-86 method in the field provided here. Comment-</b>	

\* The LAB H has performed 9200.1-86 on 143 client samples. 80 of these were tested for lead and the remaining 63 were tested for arsenic. These numbers do not count laboratory QC samples or work performed during method development and documentation of acceptable performance prior to running client samples.

**Appendix G  
Laboratory Submitted IDP Forms**

<b>Microwave Digestion using 3051A</b>		
<b>8</b>	<b>Total number of analyses your facility has performed for lead and arsenic using the attached EPA Method 3051A.</b>	Typically has been used for oil or tissue matrix only, not soil or sediment. Currently performing MDLs and precision and accuracy studies for soil and recently updated soil procedure in SOP 420. No client soil samples in several years.
<b>9</b>	<b>Will your facility conduct the digestion? (Yes/No)</b>	Yes
<b>10</b>	<b>If the answer to question 2 is no, please provide the name of the laboratory that will be conducting the Digestion. (Lab Name)</b>	
<b>11</b>	<b>Will your facility conduct the digest analysis? (Yes/No)</b>	Yes
<b>12</b>	<b>If the answer to question 4 is no, please provide the name of the laboratory that will be conducting the analysis. (Lab Name)</b>	
<b>13</b>	<b>Will your facility be able to conduct the attached EPA Method 3051A as written? (Yes, or Provide comment Below in 7)</b>	See below.
<b>14</b>	<b>If the answer the question 6 is no, please provide the deviation from EPA Method 3051A in the field provided here. Comment-</b>  See appendix A of LAB H Lab SOP 420 for deviations.	

**Apparatus IVBA**

<b>16</b>	<b>Does the IVBA apparatus your facility has use air or water as the 37°C thermal conducting/controlling medium. (Air, Water)</b>	Air
<b>17</b>	<b>How many bottle positions does your apparatus have?</b>	It holds 12 x 2L bottles. Each 2L bottle can hold about ten 125 mL IVBA extraction bottles. Total = 120
<b>18</b>	<b>Does your usual protocol allow for the pre-incubation of the extraction solution to 37°C before initiation of the IVBA extraction.</b>	Yes
<b>19</b>	<b>How do you measure temperature of the controlling apparatus?</b>	Digital thermometer with data logger.
<b>20</b>	<b>If your lab uses air control, are you using a commercially available extraction apparatus? If possible, please provide the vendor and part number, or Model number.</b>	Associated Designs 3740-12BRE (12 place TCLP rotary

## Appendix G Laboratory Submitted IDP Forms

		agitator)
21	If your lab uses air control, what type of temperature control device is being used (i.e., benchtop, upright, or walk-in incubator)?	Walk-in
22	Does your lab use a pH probe which compensates for temperature (i.e., Automatic Temperature Control (ATC probe))?	Yes
23	If not, how does your lab control for temperature when measuring the pH?	

### Analytical IVBA

24	Type of analytical instrument typically used for the final Determination (ICP-AES) (ICP-MS) (GFAA) or specify other instrument type.	Typically use ICP/AES. ICP/MS could be used if necessary.
25	Please provide the aqueous method detection limit for the procedure that you currently use for the IVBA method for both Lead and Arsenic. (□g/L)	As = 10 ug/L Pb = 15 ug/L
26	Name of Control Soil - Reference Material typically used by your facility for the IVBA extraction. ( e.g., NIST 2710 or 2711, or other)	NIST 2711A
27	Blank spike amount (mg/L) used in your procedure.	1 mg/L
28	Matrix spike amount (mg/L) used in your procedure.	5 mg/L

### Apparatus Microwave

29	Does the Microwave apparatus at your facility have temperature or pressure control.	temperature
30	How many vessel positions does your apparatus have?	12
31	Please provide the manufacturer and model of your microwave Apparatus.	CEM MARS Xpress
32	What procedure do you use for the microwave power calibration?	Not performed, use temp control
33	When was your microwave apparatus last power calibrated?	N/A

**Appendix G  
Laboratory Submitted IDP Forms**

**Table of Batch Lead IVBA Results Table modified by CLJ - QATS**

No	Batch Date	Reagent Blank µg/L	Bottle Blank µg/L	Spiked Blank Result	Spike Blank Percent Recovery	Matrix Spike Percent Recovery	Duplicate Relative Percent Difference	Reference Material Name	Control Soil Result (mg/L) (analytical solution)	Control Soil RPD	Control Soil IVBA	Determination by ICP-AES or ICP-MS
<b>A</b>	<b>Date</b>	<b>&lt;25 µg/L</b>	<b>&lt;50 µg/L</b>	<b>(mg/L)</b>	<b>85-115%</b>	<b>75-125%</b>	<b>&lt;20%RPD</b>			<b>&lt;10%RPD</b>	<b>IVBA%</b>	
<b>B</b>	<b>Mm/dd/yyyy</b>	<b>&lt;25 µg/L</b>	<b>&lt;50 µg/L</b>	<b>9.2</b>	<b>92.4%</b>	<b>87.3%</b>	<b>7.4%</b>	<b>NIST 2711</b>	<b>9.12 mg/L</b>	<b>7.1%</b>	<b>82.9%</b>	
1	04/11/2011	<15	?	0.99	99	93	0.1	NIST 2711a	10.5	3.7	75	ICP-AES
2	04/13/2011	<15	?	5.21	104	104	7	NIST 2711a	11.6	6.2	83	ICP-AES
3	10/03/2011	<15	?	0.98	98	80	4	NIST 2711a	11	2.8	76	ICP-AES
4												
5												
6												
7												
8												
9												
10												

**Note Row A presents the quality control acceptance criteria from the USEPA IVBA Method EPA 9200.1-86, and Row B provides an example.**



**Appendix G  
Laboratory Submitted IDP Forms**

**Table of EPA Microwave Method 3051a Soil Batch Lead Results**

No	Batch Date	Reagent Blank µg/L	Matrix Spike Percent Recovery	Duplicate Sample Relative Percent Difference	LCS or Reference Material Name	LCS or Reference Material Nominal Value (mg/Kg)	Reference Material Result (mg/ Kg)	Reference Material Result Percent Recovery	Determination by ICP-AES or ICP-MS
<b>A</b>	<b>Date</b>	<b>&lt;25 µg/L</b>	<b>75-125%</b>	<b>&lt;20%RPD</b>					
<b>B</b>	<b>Mm/dd/yyyy</b>	<b>&lt;25 µg/L</b>	<b>87.3%</b>	<b>7.4%</b>	<b>NIST 2711</b>	<b>1100 mg/kg</b>	<b>912 mg/L</b>	<b>82.9%</b>	
1									
2									
3									
4									
5									
6									
7									
8									
9									
10									

Note: Row A presents typical quality control acceptance criteria from the USEPA Method 6010, and Row B provides an example.

**Appendix G  
Laboratory Submitted IDP Forms**

**Table of EPA Microwave Method 3051a Soil Batch Arsenic Results**

No	Batch Date	Reagent Blank µg/L	Matrix Spike Percent Recovery	Duplicate Sample Relative Percent Difference	LCS or Reference Material Name	LCS or Reference Material Nominal Value (mg/Kg)	Reference Material Result (mg/ Kg)	Reference Material Result Percent Recovery	Determination by ICP-AES or ICP-MS
<b>A</b>	<b>Date</b>	<b>&lt;25 µg/L</b>	<b>75-125%</b>	<b>&lt;20%RPD</b>					
<b>B</b>	<b>Mm/dd/yyyy</b>	<b>&lt;25 µg/L</b>	<b>87.3%</b>	<b>7.4%</b>	<b>NIST 2711</b>	<b>90 mg/kg</b>	<b>81 mg/kg</b>	<b>90.0%</b>	
1									
2									
3									
4									
5									
6									
7									
8									
9									
10									

**Note: Row A presents typical quality control acceptance criteria from the USEPA Method 6010, and Row B provides an example.**

## **APPENDIX H**

### **Laboratory Submitted Round Robin Study Sample Results**

# Laboratory A

## Appendix H Laboratory Submitted Study Results

### LAB A- Statement of Work for the Lead IVBA and EPA Method 3051A (for Lead and Arsenic) Round Robin Analyses of a New Reference Material (RM)

(version 4, April 9, 2012)

**Introduction:** The purpose of this Statement of Work (SOW) is to provide specific information and procedures for the analysis and reporting for (1) EPA SOP EPA 9200.2-86 (the lead only IVBA) and (2) EPA method 3051A (for analysis lead and arsenic) Round Robin analyses for the New EPA IVBA Reference Material (RM). Please read carefully. Analyses of the New RM must be performed in strict accordance with the EPA SOP EPA 9200.2-86 (see attachment 1) and EPA Method 3051A (see attachment 2). Any exceptions to the SOP procedures will be provided in this Statement of Work. **Please note the sample extraction, analyses, and reporting are to be completed within a thirty (30) day-turn-around time.**

**Suggestions or Edits to the EPA SOP:** If you have any suggested changes to the Lead IVBA SOP 9200.2-86 that might be helpful, either editorial or technical, it would be great if you could e-mail (or provide on CD) a word document with the suggested changes along with the Round Robin Study results. If you wish, you could use the "Tools - Tract Changes" feature of WORD to provide the suggested changes to the WORD copy of the Lead IVBA SOP 9200.2-86 that you received on the CD with the Round Robin Study samples.

**Sample Receipt:** Two 30 mL Nalgene (polyethylene) wide mouth bottles will be provided to you. One bottle will contain approximately twenty (20) grams of the **New IVBA RM Sample**, and the second bottle will also contain approximately five (5) grams of **NIST SRM 2710a**. The bottles will be logged in to your usual sample receipt login system; however, these soil materials will not require refrigeration.

#### (1) EPA SOP EPA 9200.2-86 (Lead IVBA)

**Required Quality Assurance/Quality Control:** During the EPA review of the Initial Demonstration of Proficiency Forms (IDP) Forms submitted by the laboratories participating in the Round Robin Study, it was noted that not all laboratories performed each of the Quality Control samples that are presented in the SOP EPA 9200.2-86. **It is imperative for this study that all of the required quality control samples are prepared and analyzed as specified in the SOP EPA 9200.2-86.** It was also noted during the reviewed of the IDP Forms that different laboratories use varying acceptance criteria for the quality control parameters. **It is a requirement for this study** that the acceptance criteria presented in the SOP EPA 9200.2-86 be used for quality control sample results. Below is a table of the required quality control samples and the control limits, which was derived from Section 9 of the SOP EPA 9200.2-86. Limits that are lower than those specified for the Reagent Blank and the Bottle Blank are acceptable. Please note that a designated duplicate sample is **not** required for these analyses.

QC Sample	Control Limits
Reagent blank	<25 µg/L lead
Bottle blank	<50 µg/L lead
Blank spike (10 mg/L)	85-115% recovery
Matrix spike (10 mg/L)	75-125% recovery
Duplicate sample	±20% RPD
Control soil (NIST 2710a )	NIST 2710a Mean 67.5% (Acceptable Range 60.7- 74.2%)

All quality Control Samples must be run on every batch extraction of the NIST materials. **The New RM must each be extracted in a batch with a complete set of quality control QC samples.** Please note that the control soil NIST SRM 2710a range is based on the NIST Addendum to the Certificate of Analysis leachable median lead value of 5100 mg/kg, **not** the total dissolution Certificate of Analysis lead value 5520 mg/kg.

## Appendix H Laboratory Submitted Study Results

**Sample preparation:** The provided **New IVBA RM Sample** and **NIST SRM 2710a** should be used as is. The oven drying and the sieving to less than 250um should **not** be performed. Also, riffle splitting should **not** be performed on these SRM materials. The New RM must be extracted in a single extraction batch, with five (5) replicate RM samples, along with complete associated QC samples for each batch. To insure homogeneity, the New RM and NIST SRM 2710a bottles **must** be rotated along the x, y, and z axes for at least one minute before sub-sampling for extraction. **Note: the New RM and NIST SRM 2710a materials used in this study must be weighted out to 1.000 +/- 0.001 g, which is a more precise weighing than the SOP requirement.** The extraction apparatus may have the extraction temperature controlled to  $37 \pm 2$  °C by either air (incubator type) or water (aquarium type). For either incubator or aquarium type of extractor, the sample rotation speed must be 28 RPM as specified in the SOP.

The batch sequence that **must** be used for this study is provided in Table 1 below. Again, please note that a designated duplicate sample is not required. The sample extraction will proceed as presented in the SOP.

**Table 1. IVBA Lead Extraction Batch for Round Robin Analysis of the New RM**

IVBA Extraction Batch for Lead		
Extractor Position	Sample Name	Comment
1	New RM	
2	New RM	
3	New RM	
4	New RM	
5	New RM	
6	New RM Matrix Spike	10 mg/L Pb
7	Bottle Blank	
8	Blank Spike	10 mg/L Pb
9	Control Soil NIST SRM 2710a	

**Sample Filtering and Analysis:** Sample filtering and analysis should proceed as indicated in the SOP. The analysis will be performed using either EPA SW-846 method 6010C (ICP-AES) or 6020A (ICP-MS); however, the analytical sequence should be exactly as specified in Table 2.

**Reporting:** Tables 3 and 4 **must** be used for reporting the IVBA analysis results for the New RM and the associated QC samples results. The laboratory **must** provide copies of the calibration and the raw data print out from the instrumental analysis for both batches as part of the data submission.

Please complete the Results Tables 3 and 4 and e-mail to [clifton.jones@shawgrp.com](mailto:clifton.jones@shawgrp.com), followed by a 2<sup>nd</sup> day Fed-Ex mailing of the Results Tables 3 and 4, along with the copies of the calibration and the raw data print outs from the instrumental analysis to the address provided below. Please provide any other pertinent information regarding the RM extraction and analysis with the data submission.

Clifton Jones  
Shaw Environmental - QATS  
2700 Chandler Avenue, Bldg C  
Las Vegas, Nevada, USA  
89120  
Tel. (702) 895-8713

## Appendix H Laboratory Submitted Study Results

**Table 2. Analytical Sequence for Lead IVBA Extraction Batch for the New RM**

Position	Sample Name	Comment
Initial Standard Calibration and Beginning QC Samples	Initial Standard Calibration	
	Interference Check Sample (s)	
	Initial Calibration Verification and/or Continuing Calibration Standards and Blanks, as per EPA Methods 6010C or 6020A.	
10(<<proxy position no.)	Reagent Blank	
11	Bottle Blank	(from Extractor Position 7)
12	New RM ( Extractor Position 1)	
13	New RM (Extractor Position 2)	
14	New RM (Extractor Position 3)	
15	New RM (Extractor Position 4)	
16	New RM (Extractor Position 5)	
17	Control Soil NIST SRM 2710a	(from Extractor Position 9)
18	Blank Spike	10 mg/L (from Extractor Position 8)
19	New RM Matrix Spike	10 mg/L (from Extractor Position 6)
20	Continuing Calibration Verification Standard	
21	Continuing Calibration Verification Blank	
Analytical Run Closing QC Samples-	Interference Check Sample etc. as required by either EPA Methods 6010C or 6020A.	

**Appendix H  
Laboratory Submitted Study Results**

**(1) EPA SOP EPA 9200.2-86 (Lead IVBA)**

Table3: Laboratory, Instrument, Instrumental MDL, and IVBA Extraction Batch  
Data Reporting Form for new RM

Laboratory Performing Extraction	Lab A			
Laboratory Performing Analysis	Lab A			
<b>IVBA Extraction Batch Results new RM: Lead</b>				
Instrument Type? (ICP-AES or ICP-MS)	ICP-MS	Instrument Method Detection Limit (MDL) (ug/L)	2.0 ug/L	
Extraction Date	May 10, 2012			
Extraction Lead Standard Manufacturer and Lot #	Ultra Scientific ICP-082, Lot# L00394			
Analysis Date(s)	May 18, 2012			
Analysis Lead Standard Manufacturer and Lot #	Inorganic Ventures, E2-MEB393062			
Initial Calibration Verification Standard Source and Lot #	High Purity Standards, 1125704			
Interference Check Sample Source and Lot #	n/a			
<b>Sample Name</b>	<b>Instrument result for the analytical solution (ug/L)</b>	<b>Dilution Factor</b>	<b>Final Instrumental result analytical solution (corrected for dilution) (ug/L)</b>	<b>Result in mg/Kg (corrected for 1g/100mL extraction) (i.e., ug/L times 100/1000 = mg/kg)</b>
EXAMPLE SOIL (NIST 2710a)	70	10	700	70
Reagent Blank	1.01	1	1.01	
Bottle Blank	0.72	1	0.72	
RM (Extractor Position 1)	43600	1	43600	4360
RM (Extractor Position 2)	45100	1	45100	4491
RM (Extractor Position 3)	43900	1	43900	4387
RM (Extractor Position 4)	44700	1	44700	4448
RM (Extractor Position 5)	44500	1	44500	4409
Control Soil SRM 2710a	33300	1	3300	3325
Blank Spike	9140	1	9140	914
RM Matrix Spike	56400	1	56400	5547



**Appendix H  
Laboratory Submitted Study Results**

**Table 4. Lead Extraction Batch Spiked Blank and Spiked Sample Results for new RM**

<b>Laboratory Performing Extraction</b>	<b>Lab A</b>
<b>Laboratory Performing Analysis</b>	<b>Lab A</b>
<b>IVBA Extraction Batch Spiked Blank and Spiked Sample Results for New RM: Lead</b>	
<b>Bottle Blank Result (mg/L)</b>	0.0010
<b>Blank Spike Result (mg/L)</b>	0.00072 (9140) CLJ
<b>Blank Spike Percent Recovery</b>	91.4%
<b>Average (5) Result RM (mg/L)</b>	44.2
<b>RM Matrix Spike Result (mg/L)</b>	55.4
<b>RM Matrix Spike Percent Recovery</b>	100.2

**Appendix H  
Laboratory Submitted Study Results**

**(2) SW-846 METHOD 3051A MICROWAVE ASSISTED ACID DIGESTION OF SEDIMENTS, SLUDGES, SOILS, AND OILS**

Table 3. Laboratory, Instrument, Instrumental MDL, and 3051A Digestion  
Data Reporting Form for new RM: Results for Lead

Laboratory Performing Extraction	Lab A			
Laboratory Performing Analysis	Lab A			
<b>Digestion Batch Results new RM: Lead</b>				
Instrument Type? (ICP-AES or ICP-MS)	ICP-MS	Instrument Method Detection Limit (MDL) (ug/L)		
Digestion Date	May 9, 2012			
Digestion Lead Standard Manufacturer and Lot #	Ultra Scientific ICP-082, Lot# L00394			
Analysis Date(s)	May 18, 2012			
Analysis Lead Standard Manufacturer and Lot #	n/a			
Initial Calibration Verification Standard Source and Lot #	n/a			
Interference Check Sample Source and Lot #	n/a			
Sample Name	Instrument result for the analytical solution (ug/L)	Dilution Factor	Final Instrumental result analytical solution (corrected for dilution) (ug/L)	Result in mg/Kg (corrected for 0.5g/100mL extraction)(i.e ug/L times 200/1000 = mg/kg)
EXAMPLE SOIL (NIST 2710a)	35	10	350	70
Reagent Blank	0.94	1	0.94	
RM (Sample 1)	30900	1	30900	6180
RM (Sample 2)	30300	1	30300	6036
RM (Sample 3)	33300	1	33300	6657
RM (Sample 4)	32900	1	32900	6579
RM (Sample 5)	32600	1	32600	6439
Control Soil SRM 2710a	27800	1	27800	5554
Blank Spike	9320	1	9320	
RM Matrix Spike	42800	1	42800	

**Appendix H  
Laboratory Submitted Study Results**

**Table 4. 3051A Digestion Spiked Blank and Spiked Sample  
Results for new RM: Results for Lead**

<b>Laboratory Performing Extraction</b>	Lab A
<b>Laboratory Performing Analysis</b>	Lab A
<b>3051A Digestion Spiked Blank and Spiked Sample Results for New RM: Lead</b>	
<b>Blank Spike Result (mg/L)</b>	0.00094 mg/L(9.320) CLJ
<b>Blank Spike Percent Recovery</b>	93.2%
<b>Average (5) Result RM (mg/L)</b>	32.0
<b>RM Matrix Spike Result (mg/L)</b>	42.8
<b>RM Matrix Spike Percent Recovery</b>	101.9%

**Appendix H  
Laboratory Submitted Study Results**

**Table 5. Laboratory, Instrument, Instrumental MDL, and 3051A Digestion  
Data Reporting Form for new RM: Results for Arsenic**

Laboratory Performing Extraction	Lab A			
Laboratory Performing Analysis	Lab A			
<b>Digestion Batch Results new RM: Arsenic</b>				
Instrument Type? (ICP-AES or ICP-MS)	ICP-MS	Instrument Method Detection Limit (MDL) (ug/L)		
Digestion Date				
Digestion Arsenic Standard Manufacturer and Lot #	Ultra Scientific ICP-033, Lot# L00431A			
Analysis Date(s)				
Analysis Arsenic Standard Manufacturer and Lot #	Inorganic Ventures, E2-MEB393062			
Initial Calibration Verification Standard Source and Lot #	High Purity Standards, 1125704			
Interference Check Sample Source and Lot #	n/a			
Sample Name	Instrument result for the analytical solution (ug/L)	Dilution Factor	Final Instrumental result analytical solution (corrected for dilution) (ug/L)	Result in mg/Kg (corrected for 0.5g/100mL extraction)(i.e ug/L times 200/1000 = mg/kg)
EXAMPLE SOIL (NIST 2710a)	35	10	350	70
Reagent Blank	0.80	1	0.80	
RM (Sample 1)	3260	1	3260	652
RM (Sample 2)	3280	1	3280	653
RM (Sample 3)	3500	1	3500	700
RM (Sample 4)	3480	1	3480	696
RM (Sample 5)	3410	1	3410	674
Control Soil SRM 2710a	7970	1	7970	1592
Blank Spike	9080	1	9080	
RM Matrix Spike	13900	1	13900	

**Appendix H  
Laboratory Submitted Study Results**

**Table 6. 3051A Digestion Spiked Blank and Spiked Sample  
Results for new RM: Results for Arsenic**

Laboratory Performing Extraction	Lab A
Laboratory Performing Analysis	Lab A
<b>3051A Digestion Spiked Blank and Spiked Sample Results for New RM : Arsenic</b>	
Blank Spike Result (mg/L)	0.00080 mg/L (9.08) CLJ
Blank Spike Percent Recovery	90.8%
Average (5) Result RM (mg/L)	3.386 mg/L
RM Matrix Spike Result (mg/L)	13.9
RM Matrix Spike Percent Recovery	103.8%

# Laboratory B

**Appendix H**  
**Laboratory Submitted Study Results**

**(1) EPA SOP EPA 9200.2-86 (Lead IVBA)**

Table 3. Laboratory, Instrument, Instrumental MDL, and IVBA Extraction Batch Data Reporting Form for New RM

Laboratory Performing Extraction	Lab B			
Laboratory Performing Analysis	Lab B			
<b>IVBA Extraction Batch Results New RM: Lead</b>				
Instrument Type? (ICP-AES or ICP-MS)	ICP-MS	Instrument Method Detection Limit (MDL) (ug/L)	333, 0.17 (CLJ)	
Extraction Date	05/14/2012			
Extraction Lead Standard Manufacturer and Lot #	CPI INT'L	#11L036		
Analysis Date(s)	5/16/2012			
Analysis Lead Standard Manufacturer and Lot #	Inorganic Ventures		E2-MEB373122	
Initial Calibration Verification Standard Source and Lot #	ACCUSTANDARD		211055033	
Interference Check Sample Source and Lot #	Inorganic Ventures		E2-MEB348034	
Sample Name	Instrument result for the analytical solution (ug/L)	Dilution Factor	Final Instrumental result analytical solution (corrected for dilution) (ug/L)	Result in mg/Kg (corrected for 1g/100mL extraction) (i.e ug/L times 100/1000 = mg/kg)
EXAMPLE SOIL	70	10	700	70
Reagent Blank	0.44	5	2.2	0
Bottle Blank	0.88	5	4.4	0
New RM (Extractor Position 1)	521	100	52100	5210
New RM (Extractor Position 2)	542	100	54200	5420
New RM (Extractor Position 3)	526	100	52600	5260
New RM (Extractor Position 4)	526	100	52600	5260
New RM (Extractor Position 5)	517	100	51700	5170
Control Soil NIST SRM 2710a	400	100	40000	4000
Blank Spike	2075	5	10375	1040
New RM Matrix Spike	644	100	64400	6440

**Appendix H  
Laboratory Submitted Study Results**

**Table 4. Lead Extraction Batch Spiked Blank and Spiked Sample Results for New RM**

<b>Laboratory Performing Extraction</b>	<b>Lab B</b>
<b>Laboratory Performing Analysis</b>	<b>Lab B</b>
<b>IVBA Extraction Batch Spiked Blank and Spiked Sample Results for New RM: Lead</b>	
<b>Bottle Blank Result (mg/L)</b>	0
<b>Blank Spike Result (mg/L)</b>	10375
<b>Blank Spike Percent Recovery</b>	104%
<b>Average (5) Result New RM (mg/L)</b>	5264
<b>New RM Matrix Spike Result (mg/L)</b>	6440
<b>New RM Matrix Spike Percent Recovery</b>	117.6%



**Appendix H**  
**Laboratory Submitted Study Results**

**(2) SW-846 METHOD 3051A MICROWAVE ASSISTED ACID DIGESTION OF SEDIMENTS, SLUDGES, SOILS, AND OILS**

Table 3. Laboratory, Instrument, Instrumental MDL, and 3051A Digestion  
Data Reporting Form for New RM: Results for Lead

Laboratory Performing Extraction	Lab B			
Laboratory Performing Analysis	Lab B			
<b>Digestion Batch Results New RM: Lead</b>				
Instrument Type? (ICP-AES or ICP-MS)		Instrument Method Detection Limit (MDL) (ug/L)	333 , 0.17 (CLJ)	
Digestion Date	05/14/2012			
Digestion Lead Standard Manufacturer and Lot #	CPI INT'L	#11L036		
Analysis Date(s)	5/16/2012			
Analysis Lead Standard Manufacturer and Lot #	Inorganic Ventures		E2-MEB373122	
Initial Calibration Verification Standard Source and Lot #	ACCUSTANDARD		211055033	
Interference Check Sample Source and Lot #	Inorganic Ventures		E2-MEB348034	
Sample Name	Instrument result for the analytical solution (ug/L)	Dilution Factor	Final Instrumental result analytical solution (corrected for dilution) (ug/L)	Result in mg/Kg (corrected for 0.5g/100mL extraction)(i.e ug/L times 200/1000 = mg/kg)
EXAMPLE SOIL	35	10	350	70
Reagent Blank	0.128	2000	256	ND@1.0
New RM (Sample 1)	279.8	20000	5596000	5600
New RM (Sample 2)	325.3	20000	6506000	6500
New RM (Sample 3)	317.5	20000	6350000	6350
New RM (Sample 4)	343.7	20000	6870000	6870
New RM (Sample 5)	310.2	20000	6200000	6200
Control Soil NIST SRM 2710a	268.6	20000	5372000	5370
Blank Spike	13.98	2000	27960	27.96
New RM Matrix Spike	342.4	20000	6848000	6850

**Appendix H  
Laboratory Submitted Study Results**

**Table 4. 3051A Digestion Spiked Blank and Spiked Sample  
Results for New RM: Results for Lead**

<b>Laboratory Performing Extraction</b>	<b>Lab B</b>
<b>Laboratory Performing Analysis</b>	<b>Lab B</b>
<b>3051A Digestion Spiked Blank and Spiked Sample Results for New RM: Lead</b>	
<b>Blank Spike Result (mg/L)</b>	<b>27.96</b>
<b>Blank Spike Percent Recovery</b>	<b>111.8</b>
<b>Average (5) Result New RM (mg/L)</b>	<b>6300</b>
<b>New RM Matrix Spike Result (mg/L)</b>	<b>6850</b>
<b>New RM Matrix Spike Percent Recovery</b>	<b>220 (250 MG/KG SPK)</b>

**Appendix H  
Laboratory Submitted Study Results**

**Table 5. Laboratory, Instrument, Instrumental MDL, and 3051A Digestion  
Data Reporting Form for New RM: Results for Arsenic**

Laboratory Performing Extraction	Lab B			
Laboratory Performing Analysis	Lab B			
<b>Digestion Batch Results New RM: Arsenic</b>				
Instrument Type? (ICP-AES or ICP-MS)	ICP-MS	Instrument Method Detection Limit (MDL) (ug/L)	333	
Digestion Date	05/14/2012			
Digestion Arsenic Standard Manufacturer and Lot #	CPI INT'L	#11L036		
Analysis Date(s)	5/16/2012			
Analysis Arsenic Standard Manufacturer and Lot #	Inorganic Ventures		E2-MEB373122	
Initial Calibration Verification Standard Source and Lot #	ACCUSTANDARD		211055033	
Interference Check Sample Source and Lot #	Inorganic Ventures		E2-MEB348034	
Sample Name	Instrument result for the analytical solution (ug/L)	Dilution Factor	Final Instrumental result analytical solution (corrected for dilution) (ug/L)	Result in mg/Kg (corrected for 0.5g/100mL extraction)(i.e ug/L times 200/1000 = mg/kg)
EXAMPLE SOIL	35	10	350	70
Reagent Blank	0.38	2000	760	ND @ 1.0
New RM (Sample 1)	358	2000	716000	716
New RM (Sample 2)	380	2000	760000	760
New RM (Sample 3)	396	2000	792000	792
New RM (Sample 4)	416	2000	832000	830
New RM (Sample 5)	411	2000	822000	822
Control Soil NIST SRM 2710a	826	2000	1652000	1650
Blank Spike	14.33	2000	28660	28.7
New RM Matrix Spike	358.3	2000	716600	717

**Appendix H  
Laboratory Submitted Study Results**

**Table 6. 3051A Digestion Spiked Blank and Spiked Sample  
Results for New RM: Results for Arsenic**

<b>Laboratory Performing Extraction</b>	<b>Lab B</b>
<b>Laboratory Performing Analysis</b>	<b>Lab B</b>
<b>3051A Digestion Spiked Blank and Spiked Sample Results for New RM: Arsenic</b>	
<b>Blank Spike Result (mg/L)</b>	<b>28.7</b>
<b>Blank Spike Percent Recovery</b>	<b>114.8</b>
<b>Average (5) Result New RM (mg/L)</b>	<b>784</b>
<b>New RM Matrix Spike Result (mg/L)</b>	<b>717</b>
<b>New RM Matrix Spike Percent Recovery</b>	<b>-268 (25 MG/KG SPK)</b>

# Laboratory C

**Appendix H**  
**Laboratory Submitted Study Results**

**(1) EPA SOP EPA 9200.2-86 (Lead IVBA)**

Table 3. Laboratory, Instrument, Instrumental MDL, and IVBA Extraction Batch Data Reporting Form for New RM

Laboratory Performing Extraction	Lab C			
Laboratory Performing Analysis	Lab C			
<b>IVBA Extraction Batch Results New RM: Lead</b>				
Instrument Type? (ICP-AES or ICP-MS)	ICP	Instrument Method Detection Limit (MDL) (ug/L)	40 ug/L	
Extraction Date	04/23/12			
Extraction Lead Standard Manufacturer and Lot #	CPI International	Lot# 11G022		
Analysis Date(s)	04/26/12			
Analysis Lead Standard Manufacturer and Lot #	CPI International	Lot#	11G022	
Initial Calibration Verification Standard Source and Lot #	SPEX	Lot#	6-171CR	
Interference Check Sample Source and Lot #	SPEX	Lot#	3-50YP	
Sample Name	Instrument result for the analytical solution (ug/L)	Dilution Factor	Final Instrumental result analytical solution (corrected for dilution) (ug/L)	Result in mg/Kg (corrected for 1g/100mL extraction) (i.e ug/L times 100/1000 = mg/kg)
EXAMPLE SOIL	70	10	700	70
Reagent Blank	<40	1	<40	<4
Bottle Blank	<40	1	<40	<4
New RM (Extractor Position 1)	9734	5	48700	4870
New RM (Extractor Position 2)	9994	5	50000	5000
New RM (Extractor Position 3)	10112	5	50600	5060
New RM (Extractor Position 4)	10261	5	51300	5130
New RM (Extractor Position 5)	9497	5	47500	4750
Control Soil NIST SRM 2710a	7886	5	39430	3943
Blank Spike	2166	5	10830	1083
New RM Matrix Spike	12248	5	61240	6124

**Appendix H  
Laboratory Submitted Study Results**

**Table 4. Lead Extraction Batch Spiked Blank and Spiked Sample Results for New RM**

<b>Laboratory Performing Extraction</b>	<b>Lab C</b>
<b>Laboratory Performing Analysis</b>	<b>Lab C</b>
<b>IVBA Extraction Batch Spiked Blank and Spiked Sample Results for New RM: Lead</b>	
<b>Bottle Blank Result (mg/L)</b>	<0.04
<b>Blank Spike Result (mg/L)</b>	10.83
<b>Blank Spike Percent Recovery</b>	108.3%
<b>Average (5) Result New RM (mg/L)</b>	49.62
<b>New RM Matrix Spike Result (mg/L)</b>	61.24
<b>New RM Matrix Spike Percent Recovery</b>	116.2%*

- The spike recovery value is unusable since the analyte concentration is disproportionate to the spike level. The recovery of the associated control sample (LCS or LFB) was acceptable.

**Appendix H**  
**Laboratory Submitted Study Results**

**2) SW-846 METHOD 3051A MICROWAVE ASSISTED ACID DIGESTION OF SEDIMENTS, SLUDGES, SOILS, AND OILS**

Table 3. Laboratory, Instrument, Instrumental MDL, and 3051A Digestion  
Data Reporting Form for New RM: Results for Lead

Laboratory Performing Extraction	Lab C			
Laboratory Performing Analysis	Lab C			
<b>Digestion Batch Results New RM: Lead</b>				
Instrument Type? (ICP-AES or ICP-MS)	ICP0	Instrument Method Detection Limit (MDL) (ug/L)		
Digestion Date	05/02/12	40		
Digestion Lead Standard Manufacturer and Lot #	CPI	Lot# 11G022		
Analysis Date(s)	05/03/12			
Analysis Lead Standard Manufacturer and Lot #	CPI	Lot#	11G022	
Initial Calibration Verification Standard Source and Lot #	SPEX	Lot#	6-171CR	
Interference Check Sample Source and Lot #	SPEX	Lot#	3-50YP	
Sample Name	Instrument result for the analytical solution (ug/L)	Dilution Factor	Final Instrumental result analytical solution (corrected for dilution) (ug/L)	Result in mg/Kg (corrected for 0.5g/100mL extraction)(i.e ug/L times 200/1000 = mg/kg)
EXAMPLE SOIL	35	10	350	70
Reagent Blank	<40	100	<4000	<400
New RM (Sample 1)	57615	100	5761500	576150
New RM (Sample 2)	60189	100	6018900	601890
New RM (Sample 3)	61556	100	6155600	615560
New RM (Sample 4)	58450	100	5845000	584500
New RM (Sample 5)	56994	100	5699400	569940
Control Soil NIST SRM 2710a	24410	200	4882000	488200
Blank Spike	9634	1	9634	963
New RM Matrix Spike	42683	200	8536600	853660



**Appendix H  
Laboratory Submitted Study Results**

**Table 4. 3051A Digestion Spiked Blank and Spiked Sample  
Results for New RM: Results for Lead**

<b>Laboratory Performing Extraction</b>	<b>Lab C</b>
<b>Laboratory Performing Analysis</b>	<b>Lab C</b>
<b>3051A Digestion Spiked Blank and Spiked Sample Results for New RM: Lead</b>	
<b>Blank Spike Result (mg/L)</b>	9.63
<b>Blank Spike Percent Recovery</b>	96.3%
<b>Average (5) Result New RM (mg/L)</b>	6872.5
<b>New RM Matrix Spike Result (mg/L)</b>	8536.6
<b>New RM Matrix Spike Percent Recovery</b>	83.2%*

- The spike recovery value is unusable since the analyte concentration is disproportionate to the spike level. The recovery of the associated control sample (LCS or LFB) was acceptable.

**Appendix H  
Laboratory Submitted Study Results**

**Table 5. Laboratory, Instrument, Instrumental MDL, and 3051A Digestion  
Data Reporting Form for New RM: Results for Arsenic**

Laboratory Performing Extraction	Lab C			
Laboratory Performing Analysis	Lab C			
<b>Digestion Batch Results New RM: Arsenic</b>				
Instrument Type? (ICP-AES or ICP-MS)	ICP	Instrument Method Detection Limit (MDL) (ug/L)		
Digestion Date	05/02/12	60		
Digestion Arsenic Standard Manufacturer and Lot #	CPI	Lot# 11B077		
Analysis Date(s)	05/03/12			
Analysis Arsenic Standard Manufacturer and Lot #	CPI	Lot#	11B077	
Initial Calibration Verification Standard Source and Lot #	SPEX	Lot#	9-61-CR	
Interference Check Sample Source and Lot #	SPEX	Lot#	3-50YP	
Sample Name	Instrument result for the analytical solution (ug/L)	Dilution Factor	Final Instrumental result analytical solution (corrected for dilution) (ug/L)	Result in mg/Kg (corrected for 0.5g/100mL extraction)(i.e ug/L times 200/1000 = mg/kg)
EXAMPLE SOIL	35	10	350	70
Reagent Blank	<60	100	<6000	<600
New RM (Sample 1)	6206	100	620600	62060
New RM (Sample 2)	6734	100	673400	67340
New RM (Sample 3)	6550	100	655000	65500
New RM (Sample 4)	6393	100	639300	63930
New RM (Sample 5)	6312	100	631200	63120
Control Soil NIST SRM 2710a	7300	200	1460000	146000
Blank Spike	9054	1	9054	905
New RM Matrix Spike	13897	200	2779400	277940

**Appendix H  
Laboratory Submitted Study Results**

**Table 6. 3051A Digestion Spiked Blank and Spiked Sample  
Results for New RM: Results for Arsenic**

<b>Laboratory Performing Extraction</b>	<b>Lab C</b>
<b>Laboratory Performing Analysis</b>	<b>Lab C</b>
<b>3051A Digestion Spiked Blank and Spiked Sample Results for New RM: Arsenic</b>	
<b>Blank Spike Result (mg/L)</b>	9.05
<b>Blank Spike Percent Recovery</b>	90.5%
<b>Average (5) Result New RM (mg/L)</b>	643.9
<b>New RM Matrix Spike Result (mg/L)</b>	2779.4
<b>New RM Matrix Spike Percent Recovery</b>	106.7%

# Laboratory D

**Appendix H  
Laboratory Submitted Study Results**

**(1) EPA SOP EPA 9200.2-86 (Lead IVBA)**

Table 3. Laboratory, Instrument, Instrumental MDL, and IVBA Extraction Batch Data Reporting Form for New RM

Laboratory Performing Extraction	Lab D			
Laboratory Performing Analysis	Lab D			
<b>IVBA Extraction Batch Results New RM: Lead</b>				
Instrument Type? (ICP-AES or ICP-MS)	ICP-MS	Instrument Method Detection Limit (MDL) (ug/L)	0.031	
Extraction Date	5/7/12			
Extraction Lead Standard Manufacturer and Lot #	SPEX 11-116PB			
Analysis Date(s)	5/8/12			
Analysis Lead Standard Manufacturer and Lot #	SPEX 11-116PB			
Initial Calibration Verification Standard Source and Lot #	SPEX 20-140JB			
Interference Check Sample Source and Lot #	Environmental Express 0929914 + 1119513			
Sample Name	Instrument result for the analytical solution (ug/L)	Dilution Factor	Final Instrumental result analytical solution (corrected for dilution) (ug/L)	Result in mg/Kg (corrected for 1g/100mL extraction) (i.e ug/L times 100/1000 = mg/kg)
EXAMPLE SOIL		50		
Reagent Blank	0		0	NA
Bottle Blank	0		-2.3	NA
New RM (Extractor Position 1)	952	50	47624	4762
New RM (Extractor Position 2)	927	50	46389	4639
New RM (Extractor Position 3)	924	50	46221	4622
New RM (Extractor Position 4)	915	50	45759	4576
New RM (Extractor Position 5)	943	50	47199	4720
Control Soil NIST SRM 2710a	718	50	35947	3595
Blank Spike	200	50	10041	NA
New RM Matrix Spike	1056	50	52836	5284

**Appendix H  
Laboratory Submitted Study Results**

**Table 4. Lead Extraction Batch Spiked Blank and Spiked Sample Results for New RM**

<b>Laboratory Performing Extraction</b>	<b>Lab D</b>
<b>Laboratory Performing Analysis</b>	<b>Lab D</b>
<b>IVBA Extraction Batch Spiked Blank and Spiked Sample Results for New RM: Lead</b>	
<b>Bottle Blank Result (mg/L)</b>	0.0
<b>Blank Spike Result (mg/L)</b>	10
<b>Blank Spike Percent Recovery</b>	100
<b>Average (5) Result New RM (mg/L)</b>	46639
<b>New RM Matrix Spike Result (mg/L)</b>	52836
<b>New RM Matrix Spike Percent Recovery</b>	62

**Appendix H**  
**Laboratory Submitted Study Results**

**(2) SW-846 METHOD 3051A MICROWAVE ASSISTED ACID DIGESTION OF SEDIMENTS, SLUDGES, SOILS, AND OILS**

Table 3. Laboratory, Instrument, Instrumental MDL, and 3051A Digestion  
Data Reporting Form for New RM: Results for Lead

Laboratory Performing Extraction	Lab D			
Laboratory Performing Analysis	Lab D			
<b>Digestion Batch Results New RM: Lead</b>				
Instrument Type? (ICP-AES or ICP-MS)	ICP-MS	Instrument Method Detection Limit (MDL) (ug/L)	0.031	
Digestion Date	5/7/12			
Digestion Lead Standard Manufacturer and Lot #	SPEX 11-116PB			
Analysis Date(s)	5/8/12			
Analysis Lead Standard Manufacturer and Lot #	SPEX 11-116PB			
Initial Calibration Verification Standard Source and Lot #	SPEX 20-140JB			
Interference Check Sample Source and Lot #	Environmental Express 0929914 + 1119513			
Sample Name	Instrument result for the analytical solution (ug/L)	Dilution Factor	Final Instrumental result analytical solution (corrected for dilution) (ug/L)	Result in mg/Kg (corrected for 0.5g/50mL extraction)(i.e ug/L times 100/1000 = mg/kg)
Reagent Blank	0		0	NA
New RM (Sample 1)	1566	50	78303	7812
New RM (Sample 2)	1724	50	86215	8141
New RM (Sample 3)	1664	50	83192	8087
New RM (Sample 4)	1631	50	81542	7878
New RM (Sample 5)	1625	50	81256	7898
Control Soil NIST SRM 2710a	989	50	49429	4912
Blank Spike	215	50	10761	NA
New RM Matrix Spike	1742	50	87100	8534

**Appendix H  
Laboratory Submitted Study Results**

**Table 4. 3051A Digestion Spiked Blank and Spiked Sample  
Results for New RM: Results for Lead**

<b>Laboratory Performing Extraction</b>	<b>Lab D</b>
<b>Laboratory Performing Analysis</b>	<b>Lab D</b>
<b>3051A Digestion Spiked Blank and Spiked Sample Results for New RM: Lead</b>	
<b>Blank Spike Result (mg/L)</b>	10.7
<b>Blank Spike Percent Recovery</b>	108
<b>Average (5) Result New RM (mg/kg)</b>	7963
<b>New RM Matrix Spike Result (mg/kg)</b>	8534
<b>New RM Matrix Spike Percent Recovery</b>	57



**Appendix H  
Laboratory Submitted Study Results**

**Table 5. Laboratory, Instrument, Instrumental MDL, and 3051A Digestion  
Data Reporting Form for New RM: Results for Arsenic**

Laboratory Performing Extraction	Lab D			
Laboratory Performing Analysis	Lab D			
Digestion Batch Results New RM: Arsenic				
Instrument Type? (ICP-AES or ICP-MS)	ICP-MS	Instrument Method Detection Limit (MDL) (ug/L)		0.015
Digestion Date	5/7/12			
Digestion Arsenic Standard Manufacturer and Lot #	SPEX 11-116PB			
Analysis Date(s)	5/8/12			
Analysis Arsenic Standard Manufacturer and Lot #	SPEX 11-116PB			
Initial Calibration Verification Standard Source and Lot #	SPEX 20-140JB			
Interference Check Sample Source and Lot #	Environmental Express 0929914 + 1119513			
Sample Name	Instrument result for the analytical solution (ug/L)	Dilution Factor	Final Instrumental result analytical solution (corrected for dilution) (ug/L)	Result in mg/Kg (corrected for 0.5g/50mL extraction)(i.e ug/L times 100/1000 = mg/kg)
Reagent Blank	0		0	NA
New RM (Sample 1)	160	50	8025	800
New RM (Sample 2)	178	50	8913	841
New RM (Sample 3)	167	50	8399	816
New RM (Sample 4)	165	50	8276	799
New RM (Sample 5)	166	50	8302	806
Control Soil NIST SRM 2710a	311	50	15561	1546
Blank Spike	219	50	10963	NA
New RM Matrix Spike	381	50	19082	1869

**Appendix H  
Laboratory Submitted Study Results**

**Table 6. 3051A Digestion Spiked Blank and Spiked Sample  
Results for New RM: Results for Arsenic**

<b>Laboratory Performing Extraction</b>	<b>Lab D</b>
<b>Laboratory Performing Analysis</b>	<b>Lab D</b>
<b>3051A Digestion Spiked Blank and Spiked Sample Results for New RM: Arsenic</b>	
<b>Blank Spike Result (mg/L)</b>	10.9
<b>Blank Spike Percent Recovery</b>	110
<b>Average (5) Result New RM (mg/L)</b>	813
<b>New RM Matrix Spike Result (mg/L)</b>	1869
<b>New RM Matrix Spike Percent Recovery</b>	106

# Laboratory E

**Appendix H**  
**Laboratory Submitted Study Results**

**(1) EPA SOP EPA 9200.2-86 (Lead IVBA)**

Table 3. Laboratory, Instrument, Instrumental MDL, and IVBA Extraction Batch Data Reporting Form for New RM

Laboratory Performing Extraction	LAB E			
Laboratory Performing Analysis	LAB E			
<b>IVBA Extraction Batch Results New RM: Lead</b>				
Instrument Type? (ICP-AES or ICP-MS)	ICP-MS	Instrument Method Detection Limit (MDL) (ug/L)		0.03
Extraction Date	05/29/12			
Extraction Lead Standard Manufacturer and Lot #	VHG – lot # 101-0037			
Analysis Date(s)	06/11/12			
Analysis Lead Standard Manufacturer and Lot #	VHG – lot # 102-0115			
Initial Calibration Verification Standard Source and Lot #	VHG – lot # 011-0103			
Interference Check Sample Source and Lot #	VHG – lot #'s 102-0114 & 104-0052			
Sample Name	Instrument result for the analytical solution (ug/L)	Dilution Factor	Final Instrumental result analytical solution (corrected for dilution) (ug/L)	Result in mg/Kg (corrected for 1g/100mL extraction) (i.e ug/L times 100/1000 = mg/kg)
EXAMPLE SOIL	70	10	700	70
Reagent Blank	0.2	9.8	1.8	NA
Bottle Blank	0.2	9.9	1.6	NA
New RM (Extractor Position 1)	506.3	97.2	49209.2	4920.9
New RM (Extractor Position 2)	497.9	97.2	48394.6	4839.5
New RM (Extractor Position 3)	490.3	98.9	48487.8	4848.8
New RM (Extractor Position 4)	500.6	97.0	48565.9	4856.6
New RM (Extractor Position 5)	494.4	97.4	48159.8	4816.0
Control Soil NIST SRM 2710a	374.5	96.5	36149.8	3615.0
Blank Spike	19.9	98.1	1956.1	NA
New RM Matrix Spike	535.4	97.3	52085.4	5208.5

**Appendix H  
Laboratory Submitted Study Results**

**Table 4. Lead Extraction Batch Spiked Blank and Spiked Sample Results for New RM**

<b>Laboratory Performing Extraction</b>	LAB E
<b>Laboratory Performing Analysis</b>	LAB E
<b>IVBA Extraction Batch Spiked Blank and Spiked Sample Results for New RM: Lead</b>	
<b>Bottle Blank Result (mg/L)</b>	.002
<b>Blank Spike Result (mg/L)</b>	1.96 (adjusted to 10 in Table) CLJ
<b>Blank Spike Percent Recovery</b>	98.0 %
<b>Average (5) Result New RM (mg/L)</b>	48.6
<b>New RM Matrix Spike Result (mg/L)</b>	52.1
<b>New RM Matrix Spike Percent Recovery</b>	103.1 %

**Appendix H**  
**Laboratory Submitted Study Results**

**(2) SW-846 METHOD 3051A MICROWAVE ASSISTED ACID DIGESTION OF SEDIMENTS, SLUDGES, SOILS, AND OILS**

Table 3. Laboratory, Instrument, Instrumental MDL, and 3051A Digestion  
Data Reporting Form for New RM: Results for Lead

Laboratory Performing Extraction	LAB E			
Laboratory Performing Analysis	LAB E			
<b>Digestion Batch Results New RM: Lead</b>				
Instrument Type? (ICP-AES or ICP-MS)	ICP-MS	Instrument Method Detection Limit (MDL) (ug/L)		0.03
Digestion Date	06/07/12			
Digestion Lead Standard Manufacturer and Lot #	VHG – lot # 101-0037			
Analysis Date(s)	06/11/12			
Analysis Lead Standard Manufacturer and Lot #	VHG – lot # 102-0115			
Initial Calibration Verification Standard Source and Lot #	VHG – lot # 011-0103			
Interference Check Sample Source and Lot #	VHG – lot #'s 102-0114 & 104-0052			
<b>Sample Name</b>	<b>Instrument result for the analytical solution (ug/L)</b>	<b>Dilution Factor</b>	<b>Final Instrumental result analytical solution (corrected for dilution) (ug/L)</b>	<b>Result in mg/Kg (corrected for 0.5g/100mL extraction)(i.e ug/L times 200/1000 = mg/kg)</b>
EXAMPLE SOIL	35	10	350	70
Reagent Blank	0.5	9.2	4.6	NA
New RM (Sample 1)	729.9	93.0	67876.7	6787.7
New RM (Sample 2)	687.0	95.2	65428.5	6542.8
New RM (Sample 3)	715.9	93.4	66867.2	6686.7
New RM (Sample 4)	701.9	93.6	65663.4	6566.3
New RM (Sample 5)	698.0	93.6	65334.0	6533.4
Control Soil NIST SRM 2710a	496.8	91.3	45372.8	4537.3
Blank Spike	5.07	91.7	464.8	NA
New RM Matrix Spike	711.6	91.4	65066.9	6506.7

**Appendix H  
Laboratory Submitted Study Results**

**Table 4. 3051A Digestion Spiked Blank and Spiked Sample  
Results for New RM: Results for Lead**

<b>Laboratory Performing Extraction</b>	LAB E
<b>Laboratory Performing Analysis</b>	LAB E
<b>3051A Digestion Spiked Blank and Spiked Sample Results for New RM: Lead</b>	
<b>Blank Spike Result (mg/L)</b>	0.465
<b>Blank Spike Percent Recovery</b>	96.9 %
<b>Average (5) Result New RM (mg/L)</b>	66.2
<b>New RM Matrix Spike Result (mg/L)</b>	65.1
<b>New RM Matrix Spike Percent Recovery</b>	98.1 %

**Appendix H  
Laboratory Submitted Study Results**

**Table 5. Laboratory, Instrument, Instrumental MDL, and 3051A Digestion  
Data Reporting Form for New RM: Results for Arsenic**

Laboratory Performing Extraction	LAB E			
Laboratory Performing Analysis	LAB E			
<b>Digestion Batch Results New RM: Arsenic</b>				
Instrument Type? (ICP-AES or ICP-MS)	ICP-MS	Instrument Method Detection Limit (MDL) (ug/L)		0.02
Digestion Date	06/07/12			
Digestion Arsenic Standard Manufacturer and Lot #	VHG – lot # 101-0037			
Analysis Date(s)	06/11/12			
Analysis Arsenic Standard Manufacturer and Lot #	VHG – lot # 102-0115			
Initial Calibration Verification Standard Source and Lot #	VHG – lot # 011-0103			
Interference Check Sample Source and Lot #	VHG – lot #'s 102-0114 & 104-0052			
<b>Sample Name</b>	<b>Instrument result for the analytical solution (ug/L)</b>	<b>Dilution Factor</b>	<b>Final Instrumental result analytical solution (corrected for dilution) (ug/L)</b>	<b>Result in mg/Kg (corrected for 0.5g/100mL extraction)(i.e ug/L times 200/1000 = mg/kg)</b>
<b>EXAMPLE SOIL</b>	<b>35</b>	<b>10</b>	<b>350</b>	<b>70</b>
Reagent Blank	0.035	9.2	0.3	NA
New RM (Sample 1)	69.76	93.0	6487.3	648.7
New RM (Sample 2)	68.87	95.2	6559.0	655.9
New RM (Sample 3)	69.47	93.4	6488.7	648.9
New RM (Sample 4)	69.88	93.6	6537.3	653.7
New RM (Sample 5)	71.27	93.6	6671.0	667.1
Control Soil NIST SRM 2710a	144.7	91.3	13215.4	1321.5
Blank Spike	24.98	91.7	2290.2	NA
New RM Matrix Spike	92.99	91.4	8502.8	850.3



**Appendix H  
Laboratory Submitted Study Results**

**Table 6. 3051A Digestion Spiked Blank and Spiked Sample  
Results for New RM: Results for Arsenic**

<b>Laboratory Performing Extraction</b>	LAB E
<b>Laboratory Performing Analysis</b>	LAB E
<b>3051A Digestion Spiked Blank and Spiked Sample Results for New RM: Arsenic</b>	
<b>Blank Spike Result (mg/L)</b>	2.3 (adjusted to 10 in table) CLJ
<b>Blank Spike Percent Recovery</b>	95.8 %
<b>Average (5) Result New RM (mg/L)</b>	6.5
<b>New RM Matrix Spike Result (mg/L)</b>	8.5
<b>New RM Matrix Spike Percent Recovery</b>	120.7%

# Laboratory F

**Appendix H**  
**Laboratory Submitted Study Results**

**(1) EPA SOP EPA 9200.2-86 (Lead IVBA)**

Table 3. Laboratory, Instrument, Instrumental MDL, and IVBA Extraction Batch Data Reporting Form for New RM

Laboratory Performing Extraction	LAB F			
Laboratory Performing Analysis	LAB F			
<b>IVBA Extraction Batch Results New RM: Lead</b>				
Instrument Type? (ICP-AES or ICP-MS)	AES	Instrument Method Detection Limit (MDL) (ug/L)	20	
Extraction Date	5/4/2012			
Extraction Lead Standard Manufacturer and Lot #	Lot# 17-94PB	SPEX Certiprep 1000 mg/L Pb Std.		
Analysis Date(s)	5/9/2012			
Analysis Lead Standard Manufacturer and Lot #	Lot# 17-94PB	SPEX Certiprep 1000 mg/L Pb Std.		
Initial Calibration Verification Standard Source and Lot #	Lot# 24-134JB	SPEX Certiprep LPC Std1, 20 mg/L Pb. ICV, CCV prepared by diluting Std into 0.4M glycine to match matrix		
Interference Check Sample Source and Lot #	Lot# 37-29AS	SPEX Certiprep 5000 mg/L Al, Ca, Mg; 2000 mg/L Fe prepared by x10 dilution into 0.4M glycine and spiked with 5mg/L Pb		
Sample Name	Instrument result for the analytical solution(ug/L)mg/L	Dilution Factor	Final Instrumental result analytical solution(corrected for dilution)-(ug/L)mg/L	Result in mg/Kg (corrected for 1g/100mL extraction) (i.e ug/L times 100/1000 = mg/kg)
EXAMPLE SOIL	70	10	700	70
Reagent Blank	- 0.00387	1	- 0.00387	
Bottle Blank	- 0.00249	1	- 0.00249	
New RM (Extractor Position 1)	46.09	1	46.09	4609
New RM (Extractor Position 2)	46.04	1	46.04	4604
New RM (Extractor Position 3)	45.49	1	45.49	4549
New RM (Extractor Position 4)	45.63	1	45.63	4563
New RM (Extractor Position 5)	45.05	1	45.05	4505
Control Soil NIST SRM 2710a	34.00	1	34.00	3400
Blank Spike	11.31	1	11.31	1131
New RM Matrix Spike	55.66	1	55.66	5566

**Appendix H  
Laboratory Submitted Study Results**

**Table 4. Lead Extraction Batch Spiked Blank and Spiked Sample Results for New RM**

<b>Laboratory Performing Extraction</b>	<b>LAB F</b>
<b>Laboratory Performing Analysis</b>	<b>LAB F</b>
<b>IVBA Extraction Batch Spiked Blank and Spiked Sample Results for New RM: Lead</b>	
<b>Bottle Blank Result (mg/L)</b>	0.00249
<b>Blank Spike Result (mg/L)</b>	11.31
<b>Blank Spike Percent Recovery</b>	113%
<b>Average (5) Result New RM (mg/L)</b>	45.66
<b>New RM Matrix Spike Result (mg/L)</b>	55.66
<b>New RM Matrix Spike Percent Recovery</b>	100%

**Appendix H**  
**Laboratory Submitted Study Results**

**(2) SW-846 METHOD 3051A MICROWAVE ASSISTED ACID DIGESTION OF SEDIMENTS, SLUDGES, SOILS, AND OILS**

Table 3. Laboratory, Instrument, Instrumental MDL, and 3051A Digestion  
Data Reporting Form for New RM: Results for Lead

Laboratory Performing Extraction	LAB F			
Laboratory Performing Analysis	LAB F			
<b>Digestion Batch Results New RM: Lead</b>				
Instrument Type? (ICP-AES or ICP-MS)	AES	Instrument Method Detection Limit (MDL) (ug/L)	6.1	
Digestion Date	5/18/2012			
Digestion Lead Standard Manufacturer and Lot #	Lot# 17-94PB	SPEX Certiprep 1000 mg/L Pb Std.		
Analysis Date(s)	5/22/2012			
Analysis Lead Standard Manufacturer and Lot #	Lot# 17-94PB	SPEX Certiprep 1000 mg/L Pb Std.		
Initial Calibration Verification Standard Source and Lot #	Lot# 24-134JB	SPEX Certiprep LPC Std1, 20 mg/L Pb. ICV, CCV prepared by diluting Std to match sample matrix		
Interference Check Sample Source and Lot #	Lot#37-29AS	SPEX Certiprep 5000 mg/L Al, Ca, Mg; 2000 mg/L Fe prepared by x10 dilution into sample matrix and spike with 5 mg/L Pb (B)		
Sample Name	Instrument result for the analytical solution(ug/L)mg/L	Dilution Factor	Final Instrumental result analytical solution(corrected for dilution)(ug/L)mg/L	Result in mg/Kg (corrected for 0.5g/100mL extraction)(i.e ug/L times 200/1000 = mg/kg)
EXAMPLE SOIL	35	10	350	70
Reagent Blank	- 0.008072	10	- 0.08072	
New RM (Sample 1)	6.838	10	68.38	6838
New RM (Sample 2)	6.742	10	67.42	6742
New RM (Sample 3)	6.815	10	68.15	6815
New RM (Sample 4)	6.739	10	67.39	6739
New RM (Sample 5)	6.844	10	68.44	6844
Control Soil NIST SRM 2710a	5.491	10	54.91	5491
Blank Spike	1.041	10	10.41	
New RM Matrix Spike	7.561	10	75.61	

**Appendix H  
Laboratory Submitted Study Results**

**Table 4. 3051A Digestion Spiked Blank and Spiked Sample  
Results for New RM: Results for Lead**

<b>Laboratory Performing Extraction</b>	<b>LAB F</b>
<b>Laboratory Performing Analysis</b>	<b>LAB F</b>
<b>3051A Digestion Spiked Blank and Spiked Sample Results for New RM: Lead</b>	
<b>Blank Spike Result (mg/L)</b>	10.41
<b>Blank Spike Percent Recovery</b>	104
<b>Average (5) Result New RM (mg/L)</b>	67.96
<b>New RM Matrix Spike Result (mg/L)</b>	75.61
<b>New RM Matrix Spike Percent Recovery</b>	76.5

**Appendix H  
Laboratory Submitted Study Results**

**Table 5. Laboratory, Instrument, Instrumental MDL, and 3051A Digestion  
Data Reporting Form for New RM: Results for Arsenic**

Laboratory Performing Extraction	LAB F			
Laboratory Performing Analysis	LAB F			
<b>Digestion Batch Results New RM: Arsenic</b>				
Instrument Type? (ICP-AES or ICP-MS)	AES	Instrument Method Detection Limit (MDL) (ug/L)	1.3	
Digestion Date	5/18/2012			
Digestion Arsenic Standard Manufacturer and Lot #	Lot# 16-97AS	SPEX Certiprep 1000 mg/L Pb Std.		
Analysis Date(s)	5/22/2012			
Analysis Arsenic Standard Manufacturer and Lot #	Lot# 16-97AS	SPEX Certiprep 1000 mg/L As Std.		
Initial Calibration Verification Standard Source and Lot #	Lot# 24-134JB	SPEX Certiprep LPC Std1, 20 mg/L As. ICV, CCV prepared by diluting Std to match sample matrix		
Interference Check Sample Source and Lot #	Lot#37-29AS	SPEX Certiprep 5000 mg/L Al, Ca, Mg; 2000 mg/L Fe prepared by x10 dilution into sample matrix (A) and spike with 5 mg/L As (B)		
Sample Name	Instrument result for the analytical solution(ug/L)mg/L	Dilution Factor	Final Instrumental result analytical solution(corrected for dilution) (ug/L) mg/L	Result in mg/Kg (corrected for 0.5g/100mL extraction)(i.e ug/L times 200/1000 = mg/kg)
EXAMPLE SOIL	35	10	350	70
Reagent Blank	- 0.00009013	10	- 0.0009013	
New RM (Sample 1)	0.7760	10	7.760	776.0
New RM (Sample 2)	0.7890	10	7.890	789.0
New RM (Sample 3)	0.7695	10	7.695	769.5
New RM (Sample 4)	0.7648	10	7.648	764.8
New RM (Sample 5)	0.7977	10	7.977	797.7
Control Soil NIST SRM 2710a	1.684	10	16.84	168.4
Blank Spike	1.019	10	10.19	
New RM Matrix Spike	1.758	10	17.58	

**Table 6. 3051A Digestion Spiked Blank and Spiked Sample  
Results for New RM: Results for Arsenic**

**Appendix H  
Laboratory Submitted Study Results**

<b>Laboratory Performing Extraction</b>	<b>LAB F</b>
<b>Laboratory Performing Analysis</b>	<b>LAB F</b>
<b>3051A Digestion Spiked Blank and Spiked Sample Results for New RM: Arsenic</b>	
<b>Blank Spike Result (mg/L)</b>	10.19
<b>Blank Spike Percent Recovery</b>	102
<b>Average (5) Result New RM (mg/L)</b>	7.794
<b>New RM Matrix Spike Result (mg/L)</b>	17.58
<b>New RM Matrix Spike Percent Recovery</b>	97.9



# Laboratory G

**Appendix H  
Laboratory Submitted Study Results**

**(1) EPA SOP EPA 9200.2-86 (Lead IVBA)**

**Table 3. Laboratory, Instrument, Instrumental MDL, and IVBA Extraction Batch Data Reporting Form for New RM**

<b>Laboratory Performing Extraction</b>		Lab G		
<b>Laboratory Performing Analysis</b>		Lab G		
<b>IVBA Extraction Batch Results New RM: Lead</b>				
<b>Instrument Type? (ICP-AES or ICP-MS)</b>	ICP - AES	<b>Instrument Method Detection Limit (MDL) (ug/L)</b>	10.64 ug/L	
<b>Extraction Date</b>	4/23/2012			
<b>Extraction Lead Standard Manufacturer and Lot #</b>	Claritas Lot # 9-145CR			
<b>Analysis Date(s)</b>	4/24/2012			
<b>Analysis Lead Standard Manufacturer and Lot #</b>	CLaritas Lot # 9-145CR			
<b>Initial Calibration Verification Standard Source and Lot #</b>	Absolute Lot # 101110			
<b>Interference Check Sample Source and Lot #</b>	QATS Lot # 0503 & 0203			
<b>Sample Name</b>	<b>Instrument result for the analytical solution (ug/L)</b>	<b>Dilution Factor</b>	<b>Final Instrumental result analytical solution (corrected for dilution) (ug/L)</b>	<b>Result in mg/Kg (corrected for 1g/100mL extraction) (i.e ug/L times 100/1000 = mg/kg)</b>
EXAMPLE SOIL	70	10	700	70
Reagent Blank	<0			<0
Bottle Blank	<0			<0
New RM (Extractor Position 1)	45380			4538
New RM (Extractor Position 2)	44340			4434
New RM (Extractor Position 3)	45840			4584
New RM (Extractor Position 4)	45890			4589
New RM (Extractor Position 5)	46260			4626
Control Soil NIST SRM 2710a	33930			3393
Blank Spike	10180			1018
New RM Matrix Spike	57670			5767

**Appendix H  
Laboratory Submitted Study Results**

**Table 4. Lead Extraction Batch Spiked Blank and Spiked Sample Results for New RM**

<b>Laboratory Performing Extraction</b>	Lab G
<b>Laboratory Performing Analysis</b>	Lab G
<b>IVBA Extraction Batch Spiked Blank and Spiked Sample Results for New RM: Lead</b>	
<b>Bottle Blank Result (mg/L)</b>	<0
<b>Blank Spike Result (mg/L)</b>	10.2 mg/L
<b>Blank Spike Percent Recovery</b>	102%
<b>Average (5) Result New RM (mg/L)</b>	45.5 mg/L
<b>New RM Matrix Spike Result (mg/L)</b>	57.7 mg/L
<b>New RM Matrix Spike Percent Recovery</b>	122%

**Appendix H  
Laboratory Submitted Study Results**

**(2) SW-846 METHOD 3051A MICROWAVE ASSISTED ACID DIGESTION OF SEDIMENTS, SLUDGES, SOILS, AND OILS**

**Table 3. Laboratory, Instrument, Instrumental MDL, and 3051A Digestion  
Data Reporting Form for New RM: Results for Lead**

<b>Laboratory Performing Extraction</b>	Lab G			
<b>Laboratory Performing Analysis</b>	Lab G			
<b>Digestion Batch Results New RM: Lead</b>				
<b>Instrument Type? (ICP-AES or ICP-MS)</b>	ICP-AES	<b>Instrument Method Detection Limit (MDL) (ug/L)</b>		10.64 ug/L
<b>Digestion Date</b>	4/23/2012			
<b>Digestion Lead Standard Manufacturer and Lot #</b>	Claritas Lot # 9-145CR			
<b>Analysis Date(s)</b>	4/24/2012			
<b>Analysis Lead Standard Manufacturer and Lot #</b>	CLaritas Lot # 9-145CR			
<b>Initial Calibration Verification Standard Source and Lot #</b>	Absolute Lot # 101110			
<b>Interference Check Sample Source and Lot #</b>	QATS Lot # 0503 & 0203			
<b>Sample Name</b>	<b>Instrument result for the analytical solution (ug/L)</b>	<b>Dilution Factor</b>	<b>Final Instrumental result analytical solution (corrected for dilution) (ug/L)</b>	<b>Result in mg/Kg (corrected for 1.0g/50mL extraction)(i.e ug/L times 50/1000 = mg/kg)</b>
EXAMPLE SOIL	35	10	350	70
Reagent Blank	<0			<0
New RM (Sample 1)	133400			6670
New RM (Sample 2)	129400			6470
New RM (Sample 3)	132100			6605
New RM (Sample 4)	133400			6670
New RM (Sample 5)	132600			6630
Control Soil NIST SRM 2710a	103900			5195
Blank Spike	10120			506
New RM Matrix Spike	140000			7000

**Appendix H  
Laboratory Submitted Study Results**

**Table 4. 3051A Digestion Spiked Blank and Spiked Sample  
Results for New RM: Results for Lead**

<b>Laboratory Performing Extraction</b>	Lab G
<b>Laboratory Performing Analysis</b>	Lab G
<b>3051A Digestion Spiked Blank and Spiked Sample Results for New RM: Lead</b>	
<b>Blank Spike Result (mg/L)</b>	10.1 mg/L
<b>Blank Spike Percent Recovery</b>	101%
<b>Average (5) Result New RM (mg/L)</b>	132 mg/L
<b>New RM Matrix Spike Result (mg/L)</b>	140 mg/L
<b>New RM Matrix Spike Percent Recovery</b>	80%

**Appendix H  
Laboratory Submitted Study Results**

**Table 5. Laboratory, Instrument, Instrumental MDL, and 3051A Digestion  
Data Reporting Form for New RM: Results for Arsenic**

<b>Laboratory Performing Extraction</b>	Lab G			
<b>Laboratory Performing Analysis</b>	Lab G			
<b>Digestion Batch Results New RM: Arsenic</b>				
<b>Instrument Type? (ICP-AES or ICP-MS)</b>	ICP - AES	<b>Instrument Method Detection Limit (MDL) (ug/L)</b>		9.09 ug/L
<b>Digestion Date</b>	4/23/2012			
<b>Digestion Arsenic Standard Manufacturer and Lot #</b>	Claritas Lot # 9-145CR			
<b>Analysis Date(s)</b>	4/24/2012			
<b>Analysis Arsenic Standard Manufacturer and Lot #</b>	Claritas Lot # 9-145CR			
<b>Initial Calibration Verification Standard Source and Lot #</b>	Absolute Lot # 101110			
<b>Interference Check Sample Source and Lot #</b>	QATS Lot # 0503 & 0203			
<b>Sample Name</b>	<b>Instrument result for the analytical solution (ug/L)</b>	<b>Dilution Factor</b>	<b>Final Instrumental result analytical solution (corrected for dilution) (ug/L)</b>	<b>Result in mg/Kg (corrected for 1g/50mL extraction)(i.e ug/L times 50/1000 = mg/kg)</b>
<b>EXAMPLE SOIL</b>	<b>35</b>	<b>10</b>	<b>350</b>	<b>70</b>
<b>Reagent Blank</b>	<0			<0
<b>New RM (Sample 1)</b>	14290			714.5
<b>New RM (Sample 2)</b>	13990			699.5
<b>New RM (Sample 3)</b>	14240			712.0
<b>New RM (Sample 4)</b>	14110			705.5
<b>New RM (Sample 5)</b>	15040			752.0
<b>Control Soil NIST SRM 2710a</b>	30100			1505
<b>Blank Spike</b>	9990			499.5
<b>New RM Matrix Spike</b>	24280			1214

**Appendix H  
Laboratory Submitted Study Results**

**Table 6. 3051A Digestion Spiked Blank and Spiked Sample  
Results for New RM: Results for Arsenic**

<b>Laboratory Performing Extraction</b>	Lab G
<b>Laboratory Performing Analysis</b>	Lab G
<b>3051A Digestion Spiked Blank and Spiked Sample Results for New RM: Arsenic</b>	
<b>Blank Spike Result (mg/L)</b>	9.99 mg/L
<b>Blank Spike Percent Recovery</b>	99.9%
<b>Average (5) Result New RM (mg/L)</b>	14.3 mg/L
<b>New RM Matrix Spike Result (mg/L)</b>	24.3 mg/L
<b>New RM Matrix Spike Percent Recovery</b>	100%

# Laboratory H



**Appendix H**  
**Laboratory Submitted Study Results**

**(1) EPA SOP EPA 9200.2-86 (Lead IVBA)**

Table 3. Laboratory, Instrument, Instrumental MDL, and IVBA Extraction Batch Data Reporting Form for New RM

Laboratory Performing Extraction		Lab H		
Laboratory Performing Analysis		Lab H		
<b>IVBA Extraction Batch Results New RM: Lead</b>				
Instrument Type? (ICP-AES or ICP-MS)	ICP-AES	Instrument Method Detection Limit (MDL) (ug/L)	4.0	
Extraction Date	5/14/2012			
Extraction Lead Standard Manufacturer and Lot #	Inorganic Ventures CGPB1-1			
Analysis Date(s)	5/15/2012			
Analysis Lead Standard Manufacturer and Lot #	Inorganic Ventures CGPB1-1			
Initial Calibration Verification Standard Source and Lot #	Spex 43-47AS			
Interference Check Sample Source and Lot #	Inorganic Ventures E2-MEB348035 and E2-MEB399019			
Sample Name	Instrument result for the analytical solution (ug/L)	Dilution Factor	Final Instrumental result (corrected for dilution) (ug/L)	Result in mg/Kg (corrected for 1g/100mL extraction) (i.e ug/L times 100/1000 = mg/kg)
EXAMPLE SOIL	70	10	700	70
Reagent Blank	4.51	1	4.51	0.451
Bottle Blank	4.13	1	4.13	0.413
New RM (Extractor Position 1)	44210	1	44210	4314
New RM (Extractor Position 2)	43580	1	43580	4285
New RM (Extractor Position 3)	43000	1	43000	4267
New RM (Extractor Position 4)	45000	1	45000	4393
New RM (Extractor Position 5)	43790	1	43790	4310
Control Soil NIST SRM 2710a	33860	1	33860	3332
Blank Spike	9835	1	9835	983.5
New RM Matrix Spike	53240	1	53240	5218

**Appendix H**  
**Laboratory Submitted Study Results**

**Table 4. Lead Extraction Batch Spiked Blank and Spiked Sample Results for New RM**

<b>Laboratory Performing Extraction</b>	<b>Lab H</b>
<b>Laboratory Performing Analysis</b>	<b>Lab H</b>
<b>IVBA Extraction Batch Spiked Blank and Spiked Sample Results for New RM: Lead</b>	
<b>Bottle Blank Result (mg/L)</b>	0.00413
<b>Blank Spike Result (mg/L)</b>	9.835
<b>Blank Spike Percent Recovery</b>	98.35
<b>Average (5) Result New RM (mg/L)</b>	43.91
<b>New RM Matrix Spike Result (mg/L)</b>	53.24
<b>New RM Matrix Spike Percent Recovery</b>	92.2

**Appendix H**  
**Laboratory Submitted Study Results**

**(2) SW-846 METHOD 3051A MICROWAVE ASSISTED ACID DIGESTION OF SEDIMENTS, SLUDGES, SOILS, AND OILS**

Table 3. Laboratory, Instrument, Instrumental MDL, and 3051A Digestion  
Data Reporting Form for New RM: Results for Lead

Laboratory Performing Extraction	Lab H			
Laboratory Performing Analysis	Lab H			
<b>Digestion Batch Results New RM: Lead</b>				
Instrument Type? (ICP-AES or ICP-MS)	ICP-AES	Instrument Method Detection Limit (MDL) (ug/L)	1.5 ug/L (0.30 mg/kg)	
Digestion Date	5/7/2012			
Digestion Lead Standard Manufacturer and Lot #	Inorganic Ventures CGPB1-1			
Analysis Date(s)	5/15/2012			
Analysis Lead Standard Manufacturer and Lot #	Inorganic Ventures CGPB1-1			
Initial Calibration Verification Standard Source and Lot #	Spex 43-47AS			
Interference Check Sample Source and Lot #	Inorganic Ventures E2-MEB348035 and E2-MEB399019			
Sample Name	Instrument result for the analytical solution (ug/L)	Dilution Factor	Final Instrumental result analytical solution (corrected for dilution) (ug/L)	Result in mg/Kg (corrected for 0.5g/100mL extraction)(i.e ug/L times 200/1000 = mg/kg)
EXAMPLE SOIL	35	10	350	70
Reagent Blank	-1.88	2	-3.76	-0.376
New RM (Sample 1)	31280	2	62560	6246
New RM (Sample 2)	32580	2	65160	6513
New RM (Sample 3)	32310	2	64620	6471
New RM (Sample 4)	32750	2	65500	6538
New RM (Sample 5)	33690	2	67380	6737
Control Soil NIST SRM 2710a	25910	2	51820	5181
Blank Spike	5237	2	10474	1047
New RM Matrix Spike	36600	2	73200	7313

**Appendix H  
Laboratory Submitted Study Results**

**Table 4. 3051A Digestion Spiked Blank and Spiked Sample  
Results for New RM: Results for Lead**

Laboratory Performing Extraction	Lab H
Laboratory Performing Analysis	Lab H
<b>3051A Digestion Spiked Blank and Spiked Sample Results for New RM: Lead</b>	
Blank Spike Result (mg/L)	10.474
Blank Spike Percent Recovery	105
Average (5) Result New RM (mg/L)	65.044
New RM Matrix Spike Result (mg/L)	73.2
New RM Matrix Spike Percent Recovery	81.2

**Appendix H  
Laboratory Submitted Study Results**

**Table 5. Laboratory, Instrument, Instrumental MDL, and 3051A Digestion  
Data Reporting Form for New RM: Results for Arsenic**

Laboratory Performing Extraction	Lab H			
Laboratory Performing Analysis	Lab H			
Digestion Batch Results New RM: Arsenic				
Instrument Type? (ICP-AES or ICP-MS)		Instrument Method Detection Limit (MDL) (ug/L)	2.35 (0.47 mg/kg)	
Digestion Date	5/7/2012			
Digestion Arsenic Standard Manufacturer and Lot #	Inorganic Ventures CGAS1-1			
Analysis Date(s)	5/15/2012			
Analysis Arsenic Standard Manufacturer and Lot #	Inorganic Ventures CGAS1-1			
Initial Calibration Verification Standard Source and Lot #	Spex 43-47AS			
Interference Check Sample Source and Lot #	Inorganic Ventures E2-MEB348035 and E2-MEB399019			
Sample Name	Instrument result for the analytical solution (ug/L)	Dilution Factor	Final Instrumental result analytical solution (corrected for dilution) (ug/L)	Result in mg/Kg (corrected for 0.5g/100mL extraction)(i.e ug/L times 200/1000 = mg/kg)
EXAMPLE SOIL	35	10	350	70
Reagent Blank	-0.12	2	-0.24	-0.024
New RM (Sample 1)	3744	2	7488	747.6
New RM (Sample 2)	3654	2	7308	730.5
New RM (Sample 3)	3706	2	7412	742.2
New RM (Sample 4)	3957	2	7914	790.0
New RM (Sample 5)	3841	2	7682	768.0
Control Soil NIST SRM 2710a	7886	2	15772	1577
Blank Spike	5193	2	10386	1039
New RM Matrix Spike	8638	2	17276	1726

**Appendix H**  
**Laboratory Submitted Study Results**

**Table 6. 3051A Digestion Spiked Blank and Spiked Sample  
Results for New RM: Results for Arsenic**

<b>Laboratory Performing Extraction</b>	<b>Lab H</b>
<b>Laboratory Performing Analysis</b>	<b>Lab H</b>
<b>3051A Digestion Spiked Blank and Spiked Sample Results for New RM: Arsenic</b>	
<b>Blank Spike Result (mg/L)</b>	10.386
<b>Blank Spike Percent Recovery</b>	104
<b>Average (5) Result New RM (mg/L)</b>	7.561
<b>New RM Matrix Spike Result (mg/L)</b>	17.276
<b>New RM Matrix Spike Percent Recovery</b>	97.1