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AN EVALUATION OF PCB TESTING CONDUCTED AT THE ALLENDALE ELEMENTARY SCHOOL PITTSFIELD, MA

FINAL REPORT

October 2007

Prepared by:

Environmental Toxicology Program Bureau of Environmental Health Massachusetts Department of Public Health 250 Washington Street Boston, Massachusetts 02108





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October 24, 2007

Mr. Dean Tagliaferro U.S. Environmental Protection Agency 10 Lyman St Pittsfield, MA 01201

Dear Mr. Tagliaferro,

The Massachusetts Department of Public Health, Bureau of Environmental Health (MDPH/BEH) is announcing the release of the final report, *An Evaluation of PCB Testing Conducted at the Allendale Elementary School, Pittsfield, MA*. This final report incorporates responses to more than sixty public comments that were received after the release of the draft report on October 25, 2006. It is important to note that no comments were received that resulted in changes to the draft conclusions of the report that were stated last fall. Specifically, based on the Department's evaluation of the indoor environmental testing and blood samples analyzed for serum PCB levels, the MDPH/BEH determined that results did not appear to reveal unusual opportunities for PCB exposures to the Allendale School community and that the levels reported for indoor air in the school were below health-based screening values. Results of the blood serum analyses for all adults and children who participated showed generally low PCB levels. The U.S. Centers for Disease Control and Prevention (CDC) reviewed the data as well and concluded that serum testing results were low and PCB congener patterns were consistent with data seen in the general U.S. population.

The MDPH/BEH appreciates the patience of the Pittsfield City Council, concerned residents, and in particular, the Allendale Elementary School community, as the Department worked to provide responses to the public comments received and to complete the final report. The final report is available on the MDPH/BEH web site (http://www.mass.gov/dph/environmental_health) under Environmental Health Investigations or by calling 800-240-4266 (617-624-5757).

Sincerely,

mleg Blanchet

Meg Blanchet, Assistant Director Environmental Toxicology Program Bureau of Environmental Health (BEH)

Cc: Suzanne K. Condon, Associate Commissioner, Director, BEH Martha J. Steele, Deputy Director, BEH

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BACKGROUND

The purpose of this report is to provide a comprehensive evaluation of the results of the follow-up surface wipe, unit ventilator filter, carpet surface dust, vacuum bag dust, indoor air testing/analysis for polychlorinated biphenyl compounds (PCBs) at the Allendale Elementary School in Pittsfield, Massachusetts (see Figure 1). In addition to indoor environmental sampling, PCB serum testing of students, parents, faculty and staff of the Allendale School was offered as a service to the Allendale School community in response to requests from some members of the Allendale School community for these tests.

The Massachusetts Department of Public Health, Bureau of Environmental Health (formerly known as the Center for Environmental Health), Environmental Toxicology Program (MDPH/BEH/ETP), in collaboration with the Pittsfield Board of Health, first conducted indoor environmental testing at the school in November and December 2005. At that time, all samples (a total of 88 samples of surface dust, indoor air, and unit ventilator filter, as well as two outdoor air samples) showed no detectable levels of PCBs. The samples were analyzed using U.S. Environmental Protection Agency (USEPA) methods, which measure Aroclors. Aroclor is the industrial trade name for commercially produced mixtures of PCBs. Subsequent to this effort, MDPH/BEH and other local and state agencies involved with the GE Site learned of two filter samples reportedly taken from the school by a community resident and analyzed by the State University of New York at Albany (SUNY). SUNY used a different analytical technique (congener-specific) than the MDPH contract laboratory and reported the presence of low level PCBs in the samples. Congeners are single, unique compounds within PCBs (ATSDR 2000). In order to best address continuing concerns, MDPH/BEH agreed to conduct additional sampling at the school in collaboration with all involved parties.

INTRODUCTION

MDPH/BEH formed an indoor environmental testing work group comprising members of the: Housatonic River Initiative (HRI); Allendale School Task Force; SUNY; Spectrum

Analytical, Inc. (SAI); Southwest Research Institute (SWRI); Allendale Elementary School; Pittsfield Board of Health; and MDPH/BEH Environmental Toxicology Program. USEPA provided technical assistance.

The workgroup developed a detailed protocol that included descriptions and rationale behind the types of samples to be collected, their location, collection and analysis methods, chain of custody, quality assurance/quality control (QA/QC), and data evaluation (see Appendix A). Three formal meetings and several conference calls amongst various workgroup members were held between January and May 2006. The draft protocol was released by MDPH/BEH in May 2006 for public comment. MDPH/BEH received eight sets of comments, which were reviewed and discussed among several workgroup members prior to the commencement of sampling. A formal response to these comments can be found in Appendix A.

Concurrent with protocol development for indoor environmental testing, MDPH/BEH also requested analytic laboratory assistance from the U.S. Centers for Disease Control and Prevention (CDC) and the MDPH State Laboratory Institute in developing a protocol for conducting serum PCB testing offered to the Allendale School community. Through the winter and early spring of 2006, MDPH/BEH staff developed or compiled the following materials:

- A summary of the PCB serum testing protocol that included a description of topics to be included in the questionnaire and a proposed interpretation of the results;
- The Consent Form for both an adult participant and a parent on behalf of their child participant;
- CDC's blood sample collection and handling protocol;
- CDC's analytic method for analyzing PCBs in serum;
- CDC's PCB sections of the Third National Report on Human Exposure to Environmental Chemicals (2005)

In April 2006 MDPH/BEH formed a Health and Medical Peer Review Team (HMPRT) comprising environmental health physicians/experts to review and comment on these materials. The HMPRT was also provided with selected articles in the most current literature on PCBs. MDPH/BEH received comments from the HMPRT, which were reviewed and incorporated into the project summary and consent documents. A formal response to these comments can be found as part of Appendix B.

INDOOR ENVIRONMENTAL TESTING

METHODS

Sample Collection

Sample collection began on Monday, June 12, 2006, the last week of the school year. The nearby General Electric disposal site was operational and receiving waste, thereby reflecting conditions that would maximize the ability to detect PCBs that might be present. Additionally, the weather was favorable for maximizing the potential for PCB volatilization from the GE disposal area (i.e., increasingly warmer, dry, and windy weather preceded by a period of rain). The weather station at Pittsfield Municipal Airport reported daily showers from the previous Wednesday, June 7th to Saturday, June 10th, with high temperatures ranging from 57° to 64° F and daily rainfall ranging from a trace to 0.4 inches. The weather began clearing on Sunday, June 11th, and continued into Monday, June 12th, with partly sunny conditions, the temperature reaching 73° F, and average wind speed of 7 miles per hour (mph) out of the west-northwest, which is the direction from the disposal site towards the school (see Figure 1) (www.wundergound.com 2006).

On Monday, June 12th, Environmental Compliance Services, Inc. (ECS) of Agawam, Massachusetts, collected the surface wipe, unit ventilator filter and carpet dust samples. Accompanying the ECS staff were Elaine Krueger, Director of the BEH/ETP, Michael Celona, Senior Environmental Analyst in the BEH/ETP, and Mr. Geoff Coelho, the

Allendale Elementary School science teacher. Dr. Phil Adamo, Chairman of the Pittsfield Board of Health, was also present for a portion of the sampling. Samples were collected according to the protocol, with one exception. The surface wipe sample from the gymnasium was originally to be collected from above a hanging ceiling light. To obtain this sample it was planned to use a hydraulic lift. However, due to an inability to get the lift to operate, the surface wipe sample was collected, with the use of a ladder, from a windowsill approximately 10 feet from the gymnasium floor. As discussed in the protocol, six additional surface wipe samples were to be collected from locations chosen by Mr. Coelho during the sampling round. The six locations were the following: the ceiling vent in the Health Office; inside classroom #24's unit ventilator in an area where air pools before being filtered; the ceiling fan blade in classroom #27; the storage bin cover in classroom #24; the ceiling pipe in classroom #23; and the top of a VCR in classroom #28. All of these locations contained visible dust and were inaccessible to the students. Most of the locations were also inaccessible to staff without the use of a ladder. See Table 1 for a list of the sample locations and Figure 2 for the school floor plan.

The collection of the air samples and vacuum bag dust samples also began on June 12th. The air samples were collected over two 24-hour periods (i.e., Monday-Tuesday and Tuesday-Wednesday). Although the Protocol discusses only one vacuum cleaner, the school uses two vacuum cleaners to vacuum the school. Therefore, two vacuum bag samples were collected on Friday, June 16th, after the vacuum cleaners were used to vacuum the entire school during the week. Chain of custody on the vacuum cleaners was maintained throughout the week by ECS staff.

Due to damage to the air and unit ventilator filter sample containers during shipping, the unit ventilator filter samples and the air samples were subsequently re-collected. The unit ventilator filters were re-sampled from the same two classrooms (#21 and #24) on Wednesday, June 14th. The air samples were collected over two 24-hour periods beginning on Thursday, June 22 (i.e., Thursday-Friday and Friday-Saturday). On Thursday, June 22nd, the nearby General Electric disposal site was operational and receiving waste. According to the weather station at Pittsfield Municipal Airport, the

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weather on Monday, June 19 and Tuesday June 20 consisted of temperatures of 86° F and 77° F and rainfall of 0.45 and 0.52 inches, respectively. The weather cleared on Wednesday, June 21, with a temperature of 75° F and an average wind speed of 5 mph out of the west-southwest direction. The weather on Thursday, June 22nd and Friday, 23rd consisted of temperatures of 81° F both days, very small amounts of precipitation (0.02 and 0.07 inches, respectively) and an average wind speed of zero mph (highest wind speeds of 8 mph and 9 mph, respectively, out of the southwest-west). On Saturday, June 24th, the temperature was 73° F, approximately 0.4 inches of rain fell, and the average wind speed was 1 mph out of the west-northwest (<u>www.wundergound.com</u> 2006).

Because three different laboratories were analyzing samples, some additional sample preparation or collection was required prior to the laboratories beginning their analysis. By medium, these additional steps are briefly described below:

- Wipes: For each wipe sample location, three co-located wipe samples were taken with each sample sent to a different laboratory. The three co-located samples were taken side-by-side (but not over the previously wiped area) at the designated sample location.
- Air: All air samples were sent to SWRI for extraction into a solution and then split such that each of the three laboratories had a portion of the extracted solution.
- Unit ventilator filters: Three clippings (from edges) from each filter sampled were collected, with one clipping sent to each of the laboratories.
- Carpet dust: Three samples from separate 25 square foot sections of the carpet were collected. Each lab received one of the three samples.
- Vacuum bag dust: Samples were sent to SWRI for extraction and then split into samples for each laboratory to analyze.

Sample Analysis

Prior to the three laboratories (SAI, SWRI, and SUNY) processing the samples, their analytical methods and quality assurance/quality control procedures (standard operating procedures or SOPs) were reviewed by the USEPA Office of Environmental Measurement and Evaluation, Quality Assurance Unit in Chelmsford, MA. The Unit provided comments on the laboratory's methods and procedures. SAI analyzed samples for Aroclors, SUNY for congeners and SWRI for both Aroclors and congeners. Aroclors are mixtures of congeners; there are 209 individual PCB congeners. For example, Aroclor 1242 comprises approximately 100 individual congeners. Different Aroclors have different congener compositions and fractions (by weight) of congeners.

SAI analyzed samples for Aroclors using USEPA Method TO-4A (air, carpet dust) and USEPA Method SW846;8082 (vacuum bag dust, wipes, vent filters). SWRI analyzed samples for Aroclors using EPA Method 8082 and congeners using a modified USEPA Method TO-4A, and SUNY analyzed for congeners using a method based on two research papers published by SUNY (DeCaprio et al. 2000, 2005). The Aroclor analyses targeted seven Aroclors, while the congener analyses targeted 101 congeners (see details in protocol contained in Appendix A). For more information on congeners contained within Aroclors and their fraction (by weight) in Aroclor mixtures, please see: www.epa.gov/toxteam/pcbid/aroclor comp.htm.

SWRI performed the GC/MS analysis for the 101 targeted congeners generally as described in TAP [test/analytical procedure] 01-0408-0491. Four 13C12 labeled congeners were added to the extract as internal standards prior to GC/MS selected ion monitoring (SIM) analysis. The labeled tetra congener was used as the internal standard for the di, tri and tetra congeners. The labeled penta and hexa congeners were used as the internal standards for the penta and hexa congeners respectively. The labeled hepta congener was used as the internal standard for the internal standard for the penta and hexa congeners respectively. The labeled hepta

¹ Determination of Pesticides, PCB Congeners, Phthalates, PBDEs and PAHs by GC/MS

Relative response factors based on the internal standards were generated for the 101 target analytes and were used for quantitation in the samples.

As part of the QA/QC protocol developed prior to the start of sampling, SWRI and SUNY agreed to analyze a dust sample from the National Institute of Standards and Technology (NIST) that contained known quantities of certain congeners. The purposes of this step were to determine the comparability of the SWRI and SUNY analyses and assess how closely their results matched with the known quantities of congeners in the NIST sample. Results of these analyses are contained in Appendix D.

Methods for Initial Screening of Results

Health assessors use a variety of health-based screening values, called comparison values, to help decide whether compounds detected in environmental samples might need further evaluation. These comparison values include cancer risk evaluation guides (CREGs) and environmental media evaluation guides (EMEGs), which are values that have been scientifically peer reviewed or derived using scientifically peer-reviewed values and published by the U.S. Agency for Toxic Substances and Disease Registry (ATSDR). CREG values provide information on the potential for carcinogenic effects, while EMEG values are used to evaluate the potential for noncancer health effects. Chronic EMEGs correspond to exposures lasting one year or longer in a residential setting. CREG values are derived assuming a lifetime of daily exposure (i.e., 70 years) in a residential setting.

If the concentration of a compound exceeds its comparison value, adverse health effects are not necessarily expected. Rather, these comparison values help in selecting compounds for further consideration. For example, if the concentration of a chemical in a medium (e.g., air) is greater than the CREG for that medium, the potential for exposure to the compound should be further evaluated for the specific situation to determine whether cancer health effects might be possible. Conversely, if the concentration is less than the CREG, it is unlikely that exposure would result in cancer health effects.

For surface wipe samples, ATSDR has no comparison values but the USEPA has a regulatory clean-up standard of 10 micrograms PCB per 100 square centimeters (10 μ g /100 cm²) for wipes collected from indoor residential surfaces that have been affected by a spill of a low-concentration PCB mixture (40 Code of Federal Regulations 761.125). In addition, the California Department of Toxic Substance Control has published a recommended clean-up guideline for PCBs on surface areas in schools of 0.1 μ g/100 cm². This recommended standard for California is intended to be protective of short and long term exposures involving dermal contact and incidental ingestion (CDTSC 2003).

Results for dust samples from carpet and the vacuum cleaners were compared to ATSDR comparison values or regulatory standards for residential soils. As is discussed in the sampling protocol, there are no available comparison values for carpet surface and vacuum cleaner bag dust. Thus, MDPH compared these dust samples to comparison values and regulatory standards for residential soil as an initial screening method to be conservative.

The ATSDR comparison values for PCBs in residential soils are 1 milligram per kilogram (mg/kg) (chronic EMEG for children), 10 mg/kg (chronic EMEG for adults), and 0.4 mg/kg (CREG) (the International Agency for Research on Cancer has classified PCBs as "probable human carcinogens" based on sufficient evidence of carcinogenicity in animals and limited evidence in humans) (ATSDR 2000). The chronic EMEG was derived by ATSDR based on a study that found immunological effects (e.g., decrease in antibodies, change in lymphocyte T-cell subsets, and decreasing trends in lymphocyte proliferation and phagocytic activity of peripheral blood monocyte) in adult monkeys that were exposed to Aroclor 1254 (Tryphonas et al., 1989, 1991 as cited in ATSDR 2000). The MDEP residential soil standard under the Massachusetts Contingency Plan (MCP) is 2 mg/kg. For air samples, results were compared with the ATSDR comparison value of 0.01 microgram per cubic meter of air (μ g/m³) (CREG).

There are no available comparison values for PCBs in vent filter samples. These results were qualitatively evaluated by reviewing information on other sample results from the same rooms.

If any sample had at least one detectable Aroclor or congener and other Aroclors or congeners that were not detected in the sample, the total PCB concentration of the sample was calculated in two ways. The first way was to assume all non-detected Aroclors or congeners in the samples to be present in the sample at a concentration of one-half the detection limit (see Table 2 for detection limits). The second method was to assume all non-detectable Aroclors or congeners not to be present (this method is how the laboratories, including SUNY, reported their data).

RESULTS

Surface Wipe Samples

For the Aroclor analysis, SAI did not detect any PCBs in any of 27 surface wipe samples, while SWRI detected PCBs in one of 27 samples [Three co-located samples were taken from 27 locations – hence, a total of 81 wipe samples were taken.] The one detection was reported by the laboratory to be a concentration of 0.144 ug/100 cm², or 0.294 ug/100 cm² assuming non-detected Aroclors were present at ½ the detection limit (Tables 3b and 4). This sample was from a windowsill and was mentioned earlier in this report. The windowsill had a large amount of dust on it, was located about 10 feet from the floor in the gymnasium, and was inaccessible without a ladder. If we assumed all non-detected Aroclors in this sample as zero, the total PCB concentration would be 0.144 μ g/100 cm², which was below the USEPA cleanup standard (10 μ g/100 cm²) and slightly above the California cleanup guideline (0.1 μ g/100 cm²) (Table 4).

Using the congener method, two of the 27 samples analyzed by SWRI showed the presence of PCBs. The sample from the same location on the gymnasium windowsill discussed above had a PCB concentration of 0.280 μ g/100 cm², similar to what SWRI

found with the Aroclor method of analysis, assuming all non-detected congeners were present at one-half the detection limit (Tables 3b and 4). Again, this was well below the EPA cleanup standard and slightly above the California cleanup guideline. One other surface wipe sample taken from a vent in the ceiling of the health office had a PCB concentration (congener method; SWRI) of 0.259 μ g/100 cm², again assuming all nondetected congeners were present at one-half the detection limit. SUNY detected a concentration of 0.0218 μ g/100cm² in a wipe sample from this location, well below both EPA and California cleanup levels. If we assume that the congeners that were not detected in these samples were at zero, then the SWRI concentrations in the gymnasium and ceiling vent samples were 0.070 and 0.006 μ g/100 cm², respectively, which are again below both cleanup levels. The nurse reported that the ceiling vent had not been cleaned for a considerable period of time and was visibly dusty and dirty (see picture). As mentioned SWRI did not detect PCB congeners in any of the other wipe samples from throughout the school.

It should be noted that the SWRI samples of the gymnasium windowsill and health office ceiling vent had between 75 and 100 percent of the detected congeners flagged with a "J". A "J" flag is a quality assurance/quality control designation that indicates that the constituent is present in the sample but the concentration lies somewhere below the method detection limit but above the lower calibration limit.

SUNY, which had a detection limit more than 10 times lower than SWRI for congener analyses of wipe samples (0.00028 vs. 0.005 μ g/100 cm², respectively), detected a maximum total congener concentration in any wipe sample of 0.0467 μ g/100 cm², below both the California and USEPA cleanup levels.

Air Samples

Air samples were collected in two indoor locations and one outdoor location for comparison, with samples collected on two different days for a total of six samples. Using the Aroclor methods, no indoor or outdoor air sample exceeded the ATSDR

comparison value (0.01 μ g/m³) (maximum concentration of 0.00333 μ g/m³; 0.00291 μ g/m³ if non-detected Aroclors are assumed to be zero) (Tables 3b and 4).

Since December 2005, the USEPA has been conducting routine ambient (outdoor) air monitoring at two locations on the Allendale School property, as well as at a location on the perimeter of the Hill 78 site located closest to the school (the "northwest" location; see Figure 3). PCBs are analyzed using the Aroclor method. Results from USEPA testing conducted at the time of the MDPH sampling effort in June 2006 revealed a concentration of 0.0071 μ g/m³ (Hill 78 perimeter) and 0.0037 μ g/m³ at each of the two Allendale School property locations (samples collected on June 22-23, 2006). These results were higher than those measured in the MDPH sample in the school and importantly were below the ATSDR comparison value. These results were flagged with a "J" value and reflect the sum of only detected Aroclors. USEPA treats non-detected Aroclors as zero when summing the concentrations.

Using the congener method, SUNY had detections in one of two samples taken from classroom number 28 and one outdoor air sample taken for comparison at concentrations of approximately the ATSDR comparison value of 0.01 μ g/m³ (0.0114 μ g/m³ and 0.0117 μ g/m³, respectively) (Tables 3b and 4). If we assume non-detects are zero, the concentrations remained similar (0.0112 μ g/m³ and 0.0116 μ g/m³). For both of these samples, SWRI detected concentrations at least ten times lower than the SUNY results using the congener method (0.001 and 0.0007 μ g/m³ for indoor and outdoor air, respectively). The SWRI air concentrations were thus below the ATSDR comparison value.

It is important to note that SUNY also had detections of PCB congeners in the air method and field blank samples that were part of the QA/QC protocol. Method blanks are used to test for sample contamination resulting from laboratory methods. They consisted of new cartridges sent directly from the ECS office in Agawam and were not handled until they were received and processed by SWRI, which then sent aliquots to SAI and SUNY. SUNY detected concentrations of 0.610 and 0.832 μ g/mL PCB congeners, assuming all

non-detected congeners were present at one-half the detection limit. Units reported for blank detections are not the same as for the air samples because the blank analyses do not have associated air volumes.

A field blank essentially involves sample collection equipment going through all the sampling and shipping procedures as the same equipment being used to collect the environmental sample (in this case, air samples) except that for the blank sampling equipment, no environmental samples are taken. Thus, a field blank is designed to determine whether improper handling of sampling equipment in the field or during shipment may result in contamination that may not originate from the environmental media being sampled. SUNY detected concentrations of 1.029 and 1.885 μ g/mL PCB congeners, assuming all non-detected congeners were present at one-half the detection limit.

SUNY therefore found PCB congeners in all four blanks, while the other laboratories did not report detections of either congeners or Aroclors in any of their blanks. Although it is possible that the cartridges themselves may contain PCB congeners as an unintended contaminant, it is also possible that there were QA/QC issues involving the analytical work at SUNY that resulted in detections in blanks. Thus, the PCB concentrations detected by SUNY may be overestimates of what was actually in the indoor or outdoor air.

Dust Samples

Three carpet dust samples were collected from the school with one sample going to each of the three labs, and dust samples from two separate vacuum bags were also collected and analyzed for PCBs. Any detections using either Aroclor or congener methods for dust were below ATSDR comparison values (Tables 3a, 3b and 4). Using the Aroclor method, SWRI detected no PCBs in the carpet dust sample, while SAI detected one Aroclor (Aroclor 1260) at a concentration of 0.0592 mg/kg. If we assume the non-detected Aroclors were present at one-half the detection limit, then the concentration of

total Aroclors would be 0.238 mg/kg, which is less than the ATSDR comparison value of 0.4 mg/kg for residential soil. Using the congener method and assuming non-detects are present at one-half the detection limit, the total concentration of PCBs in the SUNY sample was 0.111 mg/kg, while the total concentration of PCBs in the SWRI sample was 0.526 mg/kg. If the non-detected congeners were assumed to be zero, the concentrations of PCB congeners were 0.0762 mg/kg (SWRI) and 0.107 mg/kg (SUNY), both below the ATSDR comparison value (Tables 3b and 4). SWRI flagged nearly all of its congener detects with a "J" flag, meaning the detections were estimates somewhere below the method detection limit.

For the vacuum bag dust samples and Aroclor method, SWRI did not detect PCBs (detection limit of about 0.1 mg/kg), while SAI detected one Aroclor (i.e., Aroclor 1260) at 0.285 and 0.292 mg/kg, both below the ATSDR comparison value of 0.4 mg/kg. If we recalculate the total PCB concentrations using one-half the detection limit for each Aroclor, then the total PCB concentrations for the vacuum bag dust samples would be 1.28 and 1.29 mg/kg, above the ATSDR comparison value but less than the MDEP residential soil standard of 2 mg/kg.

For the congener analysis, SWRI detected 0.502 mg/kg and 0.534 mg/kg in vacuum bag dust samples (assuming half of the detection limit for all non-detected congeners), while SUNY detected a maximum of 0.0709 mg/kg in these samples. If we assume non-detects are zero, all samples analyzed by the congener method were well below the ATSDR comparison value of 0.4 mg/kg PCBs (0.0601 and 0.0687 mg/kg for SWRI) (Tables 3b and 4).

As with the carpet dust sample, SWRI flagged nearly all of its congener detections in vacuum bag dust samples with a "J" flag indicating uncertainty in the actual concentrations that lie somewhere below the method detection limit and the lower calibration limit.

SWRI and SUNY analyzed a NIST dust sample for 30 congeners that were known to be present at certified concentrations ranging from 0.00414 to 0.0402 mg/kg. SWRI detected 24 of the 30 congeners with an average recovery rate of 95%. The recovery rate is the concentration of the congeners detected by the laboratory divided by the known concentration multiplied by 100. SUNY detected 29/30 congeners, with an average recovery rate of 43 %. These results suggest that SWRI analyses for congeners were more accurate than SUNY's results.

Unit Ventilator Filter Samples

Using the Aroclor method, SAI did not detect PCBs in the two unit ventilator samples. SWRI detected 0.224 and 0.255 μ g PCBs per sample (Aroclor method) assuming Aroclors not detected are present at one-half the detection limit. If we assume nondetects are zero, the results are 0.0743 and 0.105 μ g PCBs per sample (Tables 3b and 4).

Similarly, using the congener method, both SUNY and SWRI detected the presence of PCBs, with SUNY detecting 0.0519 and 0.0786 μ g PCBs per sample (μ g/sample), while SWRI detected 0.259 and 0.278 μ g/sample, similar to their Aroclor analyses. These results assume congeners are present at one-half the detection limit. If we assume non-detects are zero, the results are 0.0467 and 0.0742 μ g/sample for SUNY and 0.0288 and 0.0907 μ g/sample for SWRI (Tables 3b and 4). About 75 percent of the detected congeners were reported by SWRI with a "J" flag.

As previously stated, there are no available guidance levels for PCBs in filter samples. Other types of samples taken in these classrooms and the results of these samples are as follows:

Air: Two indoor air samples (taken on different days) were taken in Classroom
21. The maximum concentration detected by any lab for any method was 0.007
ug/m3, or below ATSDR comparison value of 0.01 ug/m³.

• Surface wipes: Four wipe samples were taken from Classroom 24. SAI and SWRI reported non-detect for all of these samples, using either the Aroclor (SAI and SWRI) or congener method (SWRI). SUNY, using the congener method, found that three of four surface wipes from this classroom had detections, with a maximum concentration of 0.0139 ug/100cm², well below both the California and USEPA cleanup levels (0.1 and 10 ug/m100 cm², respectively).

Thus, no samples in these classrooms exceeded any available screening or cleanup levels. As an additional evaluation of the filter results from these rooms, we calculated an estimate of PCBs in the filter sample per kg filter weight to provide some level of comparison to the ATSDR comparison value for soil (given as mg PCBs per kg soil weight). Although neither SUNY nor SWRI reported the filter weights for the filter sample, it is reasonable to assume their filter samples were similar to those recorded by SAI which reported filter sample weights of 2 grams. The SWRI results are estimated to range from 0.112 to 0.128 mg PCBs/ kg filter and from 0.130 to 0.138 mg PCBs/ kg filter (Aroclor and congener methods, respectively). The SUNY analyses are estimated to be 0.0262 and 0.0400 mg PCBs/ kg filter. If we assume zero for the non-detected Aroclors or congeners, the maximum estimated concentration from the filters using either method is 0.0525 mg/kg. All of these concentrations are lower than the ATSDR comparison value of 0.4 mg/kg.

SUNY had detections in the unit ventilator filter blank sample. The other two laboratories did not have detections in the filter blank sample. Although the unit ventilator filters themselves may contain PCB congeners as an unintended contaminant, it is also possible that there may be QA/QC issues involving the analytical work at SUNY that resulted in detections in blanks. Thus, the PCB concentrations detected by SUNY may overestimate what was actually in unit ventilators.

DISCUSSION

Results of indoor environmental testing at the Allendale School revealed that most samples (93 of 98) taken did not exceed or were similar to any available health-based comparison values or surface cleanup standards or guidance used to initially screen the results (assuming non-detected Aroclor or congener were $\frac{1}{2}$ detection limit). If all non-detected congener or Aroclors were assumed to be zero, then all but one sample were below or similar to any available comparison value or cleanup guidance. The one exception was a wipe sample from the gymnasium windowsill, where the concentration slightly exceeded the California cleanup guidance (0.144 µg/wipe vs. 0.1 µg/wipe CA Guideline). The following sections further evaluate the data and compare results with available information from the scientific literature.

Surface Wipe Samples

No PCBs were detected (using the Aroclor method) in any wipe sample analyzed by SAI. One wipe sample analyzed by SWRI for Aroclors detected the presence of PCBs that was below the USEPA cleanup standard for surfaces or slightly above the California cleanup guidance. None of the other 26 wipe samples analyzed from throughout the school by SWRI using the Aroclor method had detectable PCBs. Results analyzed using the congener method assuming that all non-detects were present at one-half the detection limit revealed a similar concentration in the gym windowsill sample, as well as a similar concentration in a ceiling vent sample taken in the health office. However, when determining total congener concentrations assuming non-detected congeners were zero, the total congener concentrations were well below available cleanup standards or guidance.

The wipe samples taken from the gymnasium windowsill and ceiling vent had visible dust layers. Given the inaccessibility of these areas to students and staff and that the results of all other surface wipe samples throughout the school were either non-detect or below available guidelines or standards, it is not expected that opportunities for

exposures to PCBs in surface wipe samples from the school would result in any health concerns. Using the most conservative exposure scenario available, i.e., if the maximum concentration detected in a wipe sample was readily accessible on surfaces throughout the school on a daily basis for six years for children or 30 years for adults, opportunities for exposures to PCBs would not be expected to result in health concerns (see Appendix E for calculations).

It is important to note that surface wipe samples are generally taken to help determine where more aggressive cleaning may be necessary, not to assess health risks, as no comparison values are available. The EPA and California cleanup levels cited here are useful to help determine the need for more aggressive cleaning in the school. While most results of wipe samples were non-detect, certain areas (e.g., the windowsills and other areas not cleaned on a routine basis) should be inspected and cleaned with greater frequency.

Air Samples

No indoor or comparison outdoor air sample collected at the school and analyzed using the Aroclor method exceeded the ATSDR comparison value. Low levels of PCBs were detected in the samples, but given that PCB concentrations in air tend to be higher in warmer months than in colder months, which was an important reason to target this particular time of year for this sampling effort, these results are not remarkable. Using the congener method, SUNY detected a concentration in one indoor air sample and one comparison outdoor sample similar to the ATSDR comparison value (0.01 μ g/m³). However, SUNY also detected PCBs in QA/QC blank samples for air, and hence air results may be overestimated given detections in blank samples. SWRI detected congener-specific results about 10 times lower than the SUNY results (and hence, less than screening values). In addition, SWRI qualified about half of the detected congeners as "J" flags, or estimated values somewhere below the method detection limit.

As previously discussed, ambient (i.e., outdoor) air samples taken by the USEPA at the same time period (June 22-23) as the MDPH samples from two locations on the Allendale School property revealed higher PCB concentrations (Aroclor method) than the MDPH outdoor air samples.

As mentioned earlier, PCB concentrations in air are generally higher in the summer months than in the winter months (ATSDR 2000). This has been observed in numerous other sampling events in Pittsfield. As part of the GE site evaluation and clean-up and in order to establish outdoor background concentrations of PCBs in the Pittsfield area, an ambient air monitoring station at Berkshire Community College (BCC) was established in the mid-1990s and located approximately five miles west of the GE sites and the Allendale Elementary School. According to USEPA staff, this site was established as a background location based on discussions at a public meeting held at the time considering wind patterns and that there was no known PCB contamination on the west side of Pittsfield (USEPA, 2007). Sampling was conducted at BCC during several months in 1991, 1992, 1993, 1995, and 1996. Overall, 48 samples were collected and analyzed for PCB Aroclors. Fifteen of the 27 results taken in warmer months (i.e., mid-May to mid-September) had PCB detections, with a mean concentration of 0.001 μ g/m³. Four of 21 results taken in cooler months had PCB detections, with a mean concentration of 0.0004 µg/m³ (MDPH 2003). The SAI and SWRI results for air samples were consistent with the BCC background levels for warmer months.

More recently, USEPA has been collecting air samples at two locations outside the Allendale School and analyzing them for PCB Aroclors. From December 6, 2005, to August 30, 2006, USEPA has collected 102 samples. Fifty-four of the 68 samples (or approximately 80%) taken from May 16-September 29 had PCB detections, with a mean concentration equivalent to the samples at BCC { $0.0016 \ \mu g/m^3$ (range of detections from 0.0003 to 0.0059 $\ \mu g/m^3$)}. Nine of the 34 samples (or 26%) taken between December 6, 2005 and May 12, 2006 had PCB detections, with a mean concentration of 0.00046 $\ \mu g/m^3$ (range of detections from 0.00015 to 0.0009 $\ \mu g/m^3$).

Under the most conservative exposure scenario available, i.e., if the maximum concentration detected in an air sample was present at the school on a daily basis, for six years for children or 30 years for adults, opportunities for exposure to PCBs would not be expected to result in health concerns (see Appendix E for calculation).

To better interpret indoor air results, MDPH also evaluated available information regarding studies that have measured PCBs in indoor air. A recent evaluation of indoor air concentrations of PCBs at the New Bedford High School was conducted in April 2006 (Beta 2006). Six indoor air and two outdoor background samples were taken and analyzed for over 200 congeners. To compare with results from Allendale, we calculated the concentrations in the New Bedford data of the congeners that were analyzed at the Allendale Elementary School². These concentrations ranged from 0.000098 to 0.051 μ g/m³, with an average of 0.020 μ g/m³ (Beta 2006). The two outdoor background samples for New Bedford were 0.00087 and 0.0010 μ g/m³. The source of the PCBs was hypothesized to be building materials in the school. These results are 30-50 fold higher than observed in Allendale samples based on SWRI congener specific methods.

In the MMR school mentioned previously, in addition to wipe samples, indoor air samples were also collected (EH8E 1995a; 1995b). The average of six samples taken in three rooms in the school in September 1995 revealed an average of 1.44 μ g/m³, with a range of 1.02-2.87 μ g/m³. As noted previously, the likely source of PCBs at the school was determined to be the presence of building materials containing PCBs.

A study was conducted comparing PCB congener concentrations in air from houses near New Bedford Harbor and houses located a distance from the Harbor in southeastern Massachusetts. Portions of the New Bedford Harbor were classified as a National Priority List site in 1983 due to PCB contamination. The New Bedford area study found indoor air concentrations in houses located near New Bedford Harbor ranging from

² Note that since SWRI analyzed for more congeners than SUNY, the congeners selected from the New Bedford High School data are those analyzed for by SWRI. Summing the congeners that SUNY analyzed for produces very slightly lower concentrations.

 $0.0079 \ \mu g/m^3$ to $0.061 \ \mu g/m^3$, with a geometric mean of $0.018 \ \mu g/m^3$. Houses located a distance from New Bedford Harbor had concentrations ranging from $0.0052 \ \mu g/m^3$ to $0.051 \ \mu g/m^3$, with a geometric mean of $0.0052 \ \mu g/m^3$ (Vorhees et al 1997). These concentrations were again higher than those detected in indoor air samples from the Allendale School, demonstrating the ubiquitousness of PCBs in the environment in general.

A study conducted in North Carolina, whose purpose was to establish background indoor concentrations of contaminants, such as PCBs, found concentrations of PCBs in one group of child care centers ranging from 0.0571 to 0.246 μ g/m³ with an arithmetic mean of 0.0704 μ g/m³ and in another group of child care centers ranging from 0.00872 μ g/m³ to 0.258 μ g/m³ with an arithmetic mean of 0.0604 μ g/m³ (Wilson et al. 2001, Wilson 2006). Another study that analyzed indoor air in several office buildings, laboratories, and houses in a part of the United Kingdom for PCB congeners found concentrations ranging from 0.0011 μ g/m³ to 0.069 μ g/m³, with a mean concentration of 0.009 μ g/m³ (Currado and Harrad 1998). The indoor air samples at the Allendale School were within or less than the concentrations detected in the North Carolina and Great Britain studies.

A study conducted under contract to MDPH examined contaminants, including three PCB congeners (#52, 105, and 153) in indoor air and dust in 120 Cape Cod houses found detectable concentrations of at least one of the three congeners in indoor air from 38 of the houses. In all cases these results exceeded indoor air congener results for the Allendale School For example, congener #52 was detected in 37 of the Cape Cod houses at concentrations ranging from 0.000686 μ g/m³ to 0.0247 μ g/m³, with a mean concentration of 0.00414 μ g/m³. SUNY detected congener #52 in the four indoor air samples at concentrations ranging from 0.000213 μ g/m³ to 0.000356 μ g/m³, with a mean concentration of 0.00285 μ g/m³. SWRI detected congeners #52+69 (SWRI reported both congeners at a combined concentration) in the four indoor air samples at concentrations ranging from 0.0000500 μ g/m³ to 0.0000730 μ g/m³, with a mean concentration of 0.0000595 μ g/m³. The maximum detected congener values from inside

the Allendale School do not exceed the minimum concentrations detected in the houses on Cape Cod.

Carpet and Vacuum Bag Dust

Results from carpet and vacuum bag dust samples revealed some samples using either the Aroclor or congener method and assuming non-detection at ¹/₂ the detection limit that exceeded the ATSDR comparison value of 0.4 mg/kg but all samples were less than the regulatory residential soil standard of 2 mg/kg. If we assume non-detected Aroclors or congeners are zero, neither carpet nor vacuum bag dust samples exceeded the ATSDR comparison value of using either analytic techniques.

If we assume a maximum concentration of 0.526 mg/kg (based on calculating all nondetects at one-half the detection limit), daily exposure to children for 6 years or to adults for 30 years, opportunities for exposures to PCBs would not be expected to result in health concerns (see Appendix E for calculations).

MDPH also evaluated the scientific literature for information on indoor dust measurements in other studies. Two studies from New Bedford and North Carolina analyzed PCB concentrations in carpets. The New Bedford study found a geometric mean concentration of 1.4 mg/kg in houses near the Harbor and 0.69 mg/kg in houses located a distance from the Harbor (Vorhees et al. 1999). Another study, analyzing dust on the classroom floors of several child care centers in North Carolina for various compounds, including 20 PCB congeners, found levels of PCBs in one group of four child care centers ranging from 0.143 to 2.76 mg/kg with an arithmetic mean of 1.05 mg/kg and in another group of seven child care centers ranging from 0.072 to 28.2 mg/kg with an arithmetic mean of 7.69 mg/kg (Wilson et al. 2001). The location and/or possible effect of nearby contaminated sites are not mentioned in the study. By comparison, the results of carpet sampling at Allendale Elementary School showed maximum PCB concentrations in carpet of 0.526 mg/kg (assuming no detection = ½ detection limit),

which was lower than the geometric mean concentration found in New Bedford Harbor homes located farthest from the source of PCB contamination in the harbor.

In the North Carolina study mentioned above, the contents of the vacuum cleaner from one group of three child care centers and a second group of four child care centers were analyzed for 20 PCB congeners. These vacuum cleaners were owned and operated by the child care centers and were operated for one month before the vacuum bag was removed and its contents analyzed. The concentration in the vacuum bag from the first group of child care centers ranged from 0.139 to 1.99 mg/kg with a mean of 0.785 mg/kg and from the second group of child care centers ranged from 0.120 to 3.15 mg/kg with a mean of 2.45 mg/kg (Wilson et al. 2001). By comparison, the results of vacuum bag sampling at Allendale Elementary School, which were analyzed for 101 congeners, showed maximum PCB congener concentrations of 0.534 mg/kg. The maximum PCB Aroclor concentration was 1.29 mg/kg (assuming non-detects = $\frac{1}{2}$ detection limit), or 0.292 mg/kg (assuming non-detects = zero). These concentrations are within the range found in the North Carolina study.

The MDPH-sponsored study on Cape Cod found detectable concentrations of at least one of the PCB congeners (#52, 105, and 153) in indoor dust from 22 of the 120 houses included in the study. The dust was collected by vacuuming the surfaces of rugs, floors, upholstery, furniture, ceiling fans, and windowsills (Rudel et al. 2003). Similar to the air results, the maximum detected congener values from inside the Allendale School do not exceed the minimum concentrations detected in the Cape Cod houses. For example, congener #153 was detected in 19 of the Cape Cod houses at concentrations ranging from 0.0754 mg/kg to 35.3 mg/kg, with a mean concentration of 4.74 mg/kg. SUNY detected congener #153 in the carpet dust sample at a concentration of 0.00341 mg/kg and in the vacuum bag samples at concentrations of 0.00258 mg/kg and 0.00297 mg/kg. SWRI detected congener #153 in the carpet dust sample at a concentration of 0.0073 mg/kg and in the vacuum bag samples at concentrations of 0.0086 mg/kg and 0.01 mg/kg. All of these concentrations are lower than the detected concentrations in the Cape Cod homes.

Results from the analysis of the NIST dust sample revealed that SWRI detected fewer congeners (24/30) than SUNY (29/30) but, for those congeners detected by both labs, SWRI reported more accurate concentrations and greater percent recoveries. SUNY detected most congeners, but the recovery rates were considerably lower than SWRI, suggesting that SUNY's reported concentrations were not as accurate as SWRI's reported concentrations.

Unit Ventilator Filters

SAI did not detect PCBs in the unit ventilator samples (Aroclor method). SUNY and SWRI detected low concentrations of PCBs in the filter samples. Other media sampled in these rooms (air, surface wipes) were below any available guidelines or cleanup levels. When converting the amount of PCBs detected in the vent filter samples, the estimated concentrations in mg/kg were less than guidelines for mg/kg in soil. Thus, the filters did not appear to contain an unusual amount of PCBs, nor did the rooms in which the filter samples were taken from have other types of samples with any PCB detections above available guidance or cleanup levels.

PCB SERUM TESTING

MDPH/BEH collaborated with the CDC and the MDPH State Laboratory Institute to develop a protocol for serum PCB testing and then offered this testing to members of the Allendale School community upon request. In addition, some other residents of Pittsfield, including former students at the Allendale, requested to participate in this testing. MDPH/BEH agreed to accommodate these requests. The following sections summarize the methods and results of this effort.

METHODS

Consent Form

In order to collect blood samples, MDPH required that each participant (or parent, in the case of children) sign a consent form. MDPH/BEH developed a consent form specifically for this testing effort. The consent forms were adapted from similar consent forms previously used for participants in PCB blood testing in Berkshire County and elsewhere in Massachusetts and were reviewed and approved by the MDPH Institutional Review Board (IRB). The consent form was also approved by the MDPH Office of General Counsel and reviewed by the MDPH Health and Medical Peer Review Team. A copy of the consent form is contained in Appendix C.

Questionnaire

MDPH/BEH has developed questionnaires used in many other PCB investigations for obtaining information on risk factors that are known to or may affect serum PCB levels. For this project, previously used questionnaires were adapted to gather information that included the following: age, gender, residential history (including duration of residence), usual occupation, occupation associated with use of PCBs, company, duration, number of years attending or working at Allendale School, locations in the school where most time was spent for up to each of the last seven school years (if applicable), time spent indoors and outdoors during the school day, fish consumption in general, freshwater fish consumption (how obtained, source, Housatonic River fish), change in fishing/fish consumption habits, fiddlehead fern gathering/consumption, recreational areas and types of activities in Pittsfield area (camping, playgrounds, dirt biking, etc), hunting/wildlife consumption (type of prey, how often), gardening (type), playing in dirt or grass at current address, farm residence, open ended question on any other contact with PCBs, breast feeding and duration (for child participant), number of prior children breast fed (for adult female parent), lifestyle risk factors (e.g., smoking). The questionnaires were administered in two parts; the more lengthy first part was administered over the phone

before the blood draw and the second part was administered at the time of the blood draw. The second part of the questionnaire included questions relevant to the blood draw (i.e. weight and height) as well as questions which required the participant to view a map of the Allendale School.

Notification of PCB Testing Offer

On April 11, 2006, students, parents, faculty, and staff were sent a letter from the MDPH/BEH with an offer to conduct serum PCB sampling should any member of the Allendale community want such biologic testing. The MDPH/BEH, in partnership with the Pittsfield Board of Health, held an informational meeting on May 2, 2006, to discuss the MDPH/BEH PCB blood testing offer with members of the Allendale School community and answer any questions that people may have had prior to the actual blood testing.

Phlebotomy and Laboratory Training

MDPH contracted with Berkshire Medical Center to provide phlebotomy services to those individuals who responded to the offer to conduct serum PCB testing. BMC has provided these types of services for a number of MDPH projects in Berkshire County involving serum PCB measurements since 1995. Training for BMC staff on proper collection, preparation, and shipping procedures was provided by MDPH State Laboratory Institute staff on May 4, 2006 and May 19, 2006. Protocol specific supplies and equipment were provided by both the CDC and the MDPH State Laboratory Institute.

Sample Transport

The BMC laboratory performed blood collection processes and prepared the samples for shipping. Samples were placed on dry ice and transported by a MDPH/BEH staff member from Berkshire Medical Center to the MDPH SLI for inventory and storage until

all serum samples were collected. Once all samples were collected, they were shipped overnight to the U.S. CDC in Atlanta, Georgia.

Sample Analysis

The blood testing methodology used for the biomonitoring portion of this project is a congener-specific analysis as described in the Third National Report on Human Exposure to Environmental Chemicals published by CDC in July 2005. The Third National Report presents biomonitoring exposure data for 116-148 environmental chemicals including PCBs for the civilian, non-institutionalized U.S. population over the period 1999-2002 and is a nationally representative survey from the National Health and Nutrition Examination Survey (NHANES). While children under 12 are not included in the CDC's PCB blood analyses, children are included for the 12-19 year old age group. For the 2001-2002 NHANES, 758 children in the 12-19 age category were tested for serum PCB levels.

The analytic laboratory methods used by the CDC for the serum samples from the Allendale School community are the most up to date congener specific methods available (CDC Method HRGC/ID-HRMS, No.28). Method detection limits for PCB serum analysis are congener specific and may vary between samples, largely due to variations in sample volume (USCDC, 2005). According to the CDC the method detection limits for NHANES III range from 10.5 - 32.4 ppb (lipid-adjusted) and are typical for most methods using about 1mL of sample (USCDC, 2005). The Allendale School serum collection resulted in analysis of 2mL samples and the congener specific detection limits for these samples are approximately 10 times lower than those reported in NHANES III, i.e., 0.7 to 2.9 ppb. Similarly, detection limits for NHANES III based on whole weight basis ranged from approximately 0.01 - 0.04 ppb, while the detection limits for the Pittsfield samples ranged from approximately 0.005 - 0.02 ppb, or approximately half of NHANES. Table 5 lists method detection limits for each congener analyzed (lipid-adjusted).

The CDC analyzed serum samples for 36 congeners (including 2 pairs of co-congeners reported together) that are known to be detected in the serum of the general U.S. population and consistent with congeners analyzed in the ongoing NHANES study. The final list of 36 congeners was derived from the most recent NHANES data collection period of 2003/2004, which is not yet published or available.

Serum PCBs concentrations in the Pittsfield participants were compared with data from the 2001-2002 NHANES. CDC reported that the most appropriate way to compare the data is to take the most common 15 congeners identified in 2001-2002 NHANES that were also identified in Pittsfield participants and compare those. These congeners are 52, 74, 99, 105, 118, 138/158, 146, 153, 156, 170, 180, 187, 194, 199, and 204. CDC also reported that all of these congeners had at least a 95th percentile value from the NHANES data [A 95th percentile value means 95% of the population surveyed had serum PCB concentrations at or below this value]. For these total PCB summary calculations, non-detects were treated as the method detection limit divided by the square root of two.

NHANES reports serum PCB congener results by whole weight (ng/g of serum) and lipid-adjusted (ng/g of lipid) values for the 50th, 75th, 90th, 95th percentiles as well as calculating a geometric mean when statistically possible. The whole weight serum values (ng/g serum) reported by the CDC can be converted to ng/mL of serum by multiplying by the average density of serum samples, 1.026 g/mL. CDC also reported that the sum of the congeners by whole weight basis most closely approximate what had been previously reported in the scientific literature based on Aroclor methods.

Historically, CDC and most researchers have conducted serum PCB testing as a whole weight as µg PCBs/L blood. However, today with advances in laboratory analytical capabilities, serum PCBs are increasingly being reported using lipid-adjustment results. PCBs are associated with fatty (lipid) fractions in the blood and tend to concentrate in these fatty or lipid fractions. Hence, lipid-adjusted concentrations are numerically higher than whole weight values due to PCBs concentrating in fatty tissue. Also, lipid-adjusted values take into account differences between people in terms of lipids in the blood. For

example, if two people had the same whole weight value for serum PCBs but one had twice the concentration of lipids in the blood then the lipid-adjusted values would be half of the other one.

The Pittsfield results were also compared with available data in the scientific literature, particularly for children. These data include summary data from the 2000 ATSDR. Toxicologic Profile for PCBs, as well as studies from the Netherlands, Germany, and Alabama that included PCB serum results from children.

RESULTS

The offer to test the Allendale School community for PCB serum levels resulted in 32 participants ranging in age from 8 to 59 years. Samples were taken by BMC staff from May 31, 2006, through July 27, 2006. All samples were shipped to CDC via overnight mail on August 16, 2006.

Participation in PCB serum testing included current Allendale School students, Allendale School staff, and other concerned area residents, including former Allendale students and those living near or parents of children attending the Allendale School. Samples were collected for 14 children (ages 8-19 years) and 18 adults (ranging from 20-59 years of age). Among the children were 7 current Allendale students, 5 former Allendale students and two others. Among adults, four current Allendale School staff participated in serum PCB testing. A summary of the participant distribution by age and gender is included in Table 6.

PCB congener results were reported by the CDC on a serum whole weight (ppt, pg/g) and lipid-adjusted basis (ppb, ng/g) consistent with reporting results in NHANES. PCB congeners 138,158 and congeners 196,203 are co-congeners that cannot be separated by this methodology and are reported together. PCB congener 18 was not reported because one or more of the quality assurance/quality control (QA/QC) parameters did not meet the

specified criteria. CDC reports that this is a common result for congener 18 for all labs and that this congener is a minor contributor to total serum PCBs.

Serum PCBs in Children Ages 8-19 Years Old

A total of 14 children participated in the serum PCB testing effort. Seven children were current Allendale students, ages 8-10 years. The median (or 50th percentile) of total PCBs, (15 congeners, whole weight) for the current Allendale students was 0.117 ppb (Table 9). This compares to the NHANES 50th percentile value (12-19 year olds) of 0.345 ppb. All seven of the current Allendale students (ages 8-10 years) had serum PCB levels below the CDC 50th percentile (for children ages 12-19 years). (See Table 9).

As previously discussed, current analytical methods result in lipid-adjusted serum PCB concentrations as well. Lipid-adjusted concentrations are reported in ng PCBs/g lipid (fat) in the blood. PCBs preferentially store in fatty tissue and hence lipid-adjusted concentrations will be higher numbers reflecting the fact that PCBs are more concentrated in fatty tissue. For example, one of the Pittsfield child participants had a serum PCB level of 0.124 ppb (whole weight) and 31.4 ppb (lipid-adjusted). Both of these values are well below the corresponding 50th percentile for NHANES.

For the seven current Allendale students (ages 8-10 years) the median lipid-adjusted total PCB concentrations in serum, based on summing 15 congeners, was 25.2 ppb. This compares to the NHANES 50th percentile of 71.8 ppb (12-19 year olds). All seven students were below the NHANES 50th percentile for 12-19 year olds.

The other seven children (non-current Allendale students) were ages 12-19 years, or the same age cohort for which CDC has comparison data from NHANES. Among the seven were five former Allendale students. Median (50th percentile) serum PCB levels (15 congeners) were 0.141 ppb (whole weight). This compares to the NHANES 50th percentile of 0.345 ppb (12-19 year olds). Five of these children had serum PCB

concentrations lower than the 50th percentile value from NAHNES for this age group, while the remaining two were less than the 90th percentile value.

Lipid-adjusted results based on the sum of 15 congeners for these seven children (noncurrent Allendale students) showed a median of 26.2 ppb. The NHANES lipid-adjusted 50th percentile for this age group was 71.8 ppb. All individuals were less than the NHANES 90th percentile value (113.7 ppb, with a confidence interval of 103.5 - 133.1 ppb) for lipid-adjusted serum PCB concentrations.

Serum PCBs in Adults (Ages 20 or more years)

A total of 18 adults participated in the serum PCB testing. Of these, four were current Allendale School staff, 6 were parents of current students at the school, and the remainder were individuals living near the school or elsewhere in Pittsfield or neighboring communities. The median serum PCB level in adults (summing all 15 congeners) was 0.918 ppb (whole weight) (Table 8). This compares to the NHANES 50th percentile value in ages 20+ of 1.062 ppb. The median serum PCB level for the four Allendale staff was 1.618 ppb, or above the NHANES 50th percentile (1.062) but below the 75th percentile (1.883 ppb). As with the results for children, all adult participants had serum PCB levels less than the NHANES 90th percentile value.

Lipid-adjusted summary data showed similar results. The median serum PCB concentration (15 congeners) was 176.1 ppb. This compares with the NHANES 50^{th} percentile (lipid-adjusted) of 168.5 ppb. CDC also reported a 95 percent confidence interval around the 50^{th} percentile of 154.7 - 184.2 ppb. The 95 percent confidence interval is the range of estimated values that have a 95% probability of including the true 50^{th} percentile value for the population. Thus, because the median for Pittsfield adults was within the confidence interval of the NHANES 50^{th} percentile serum PCB level, the Pittsfield participants had serum PCB levels consistent with the general US adult population. For the current Allendale staff, the median lipid-adjusted value was 263.1 ppb, or between the NHANES 50^{th} percentile (the latter is 291.8 ppb).

As is well established in the scientific literature, serum PCB levels are higher as age increases. Although numbers were small, this trend was also observed among the Pittsfield adult participants. Three individuals in the age range of 20-39 showed median (whole weight) total PCB concentration (15 congeners) of 0.698 ppb; nine individuals ages 40-49 showed a median concentration of 0.831 ppb; and six individuals ages 50-59 showed a median concentration of 1.554 ppb.

To further address questions about the four Allendale staff mentioned above, MDPH further evaluated the serum PCB data for the current Allendale staff. The median serum PCB level for the four Allendale staff (1.618 ppb) was above the NHANES 50th percentile (1.062 ppb) but below the 75th percentile (1.883 ppb). MDPH evaluated the length of employment for each of the four participants by serum PCB level. While there were too few individuals to evaluate any meaningful trends, there are some observations that may be useful in considering any possible association between the school and individual serum PCB levels. That is, the individual with the lowest level worked there for more than a decade, while the individual with the highest level worked there for the least amount of time. If the school was the major source of PCBs, we would expect higher levels in individuals working longest at the school. It is also worth noting here that the CDC concluded that "Results of the analyses in the Pittsfield participants revealed that the Pittsfield participants showed low PCB levels on either a whole weight basis or on a lipid-adjusted congener basis as compared with the third National Report data."

Comparison of Congener Detections

In this report, data have been provided on total PCBs based on summing 15 congeners tested for. Figure 4 shows the distribution of detection frequencies of 31 of the 35 congeners analyzed by CDC in the Pittsfield adult participants. These frequencies are also provided in Figure 5 for adults from the NHANES data. The congener patterns observed for Pittsfield and NHANES are similar, suggesting similarities with what is found in the US population. In addition, in response to discussions held with the

HMPRT, we asked CDC whether congener patterns in Pittsfield differed from those they typically see in the U.S. population. CDC noted that their review did not reveal any unusual patterns among the Pittsfield participants to suggest that exposures that may have led to any evidence of PCBs in blood samples are different than the U.S. population (see Appendix F).

Figure 4 also shows that the 15 congeners selected by CDC for comparing Pittsfield with NHANES data, are indeed the most prevalent congeners found in the general U.S. population.

DISCUSSION

<u>Children</u>

Results of the serum PCB testing for Pittsfield children (n=14) show that participants, especially the seven current Allendale students (ages 8-10 years), had low levels when compared with national data from NHANES (for children ages 12-19 years), as provided by CDC. The Allendale students (n=7) had a median serum PCB level (whole weight) of 0.117ppb, well below the NHANES value of 0.345 ppb. Similar results were seen for lipid-adjusted serum PCBs. Similar to the Allendale students, children that were not currently students (n=7, ages 12-19 years) at the Allendale also had median serum PCB levels lower than comparable NHANES data.

In addition to NHANES data, there have been a limited number of scientific publications documenting serum PCB testing results in children and these are summarized here for comparison to the Pittsfield results. In a study that included Dutch children ages 3.5 years old considered to have "background" levels of exposure to PCBs, serum samples were analyzed for four specific congeners: 118, 138, 153, and 180 (Lanting et al. 1998). Results from 298 children showed a 50th percentile concentration of 0.4 ppb and 95th percentile concentration of 1.9 ppb (units of μ g/L). Among the Pittsfield children closest in age (the seven Allendale children ages 8-10 years), the 50th percentile value of the sum of these four congeners (plus 158, which was reported as part of the 138/158 pair) was
0.0758 ppb with a maximum of 0.137 ppb, or less than the 50^{th} percentile from the Dutch study.

In a study of German children ages 7-10 years old in 18 German townships (Karmaus et al., 2005), serum PCB concentrations were measured for the following congeners: 101, 118, 138, 153, 170, 180, 183, and 187. The geometric mean PCB levels (μ g/L) were as follows:

- 7 year olds (n=153): 0.54 ppb
- 8 year olds (n=160): 0.47 ppb
- 9-10 year olds (n=12): 0.33 ppb

Summarizing these same congeners (plus 158, as this was analyzed with congener 138) in the current Allendale students (age 8-10 years), the maximum total PCBs for these congeners was 0.156 ppb with a geometric mean of 0.0840 ppb, all well below the concentrations reported in Karmaus et. al.

Finally, CDC conducted a study of children in Anniston, Alabama (Orloff et al. 2003). This community was the site of a plant that formerly manufactured PCBs from 1935 to the 1970s. Serum samples were analyzed for 37 PCB congeners. A total of 37 children (ages 1-16 years) participated in this study. The total PCB concentration ranged from non-detect to 4.6 ppb (μ g/L) (whole weight). The mean concentration in children was 0.37 ppb, while the median was non-detect (detection limit < 1 ppb). Assuming the PCB concentrations in the samples without detectable PCBs to be one-half the detection level, the mean and median concentrations in children were calculated to be 1.59 and 1.10 ppb, respectively (Orloff et al. 2003). The total serum PCB median concentration among the 14 Pittsfield children (sum of all 35 congeners), assuming non-detected congeners as the detection limit divided by the square root of two, was 0.170 ppb, well below the Anniston children median. [If the Allendale results were calculated assuming all non-detects were equal to one-half the detection limit, the median concentration for Allendale would have been even lower.]

<u>Adults</u>

Adult participants had serum PCB levels consistent with NHANES data. The median serum PCB level was 0.918 (whole weight) versus the comparable NHANES level of 1.062 ppb. All adult participants had serum PCB levels less than the NHANES 90th percentile.

The ATSDR Toxicologic Profile for PCBs (2000) reports that serum PCB levels have been declining in the U.S. population. They report on more recent studies of nonoccupationally exposed populations that do not consume fish from PCB-contaminated waters (ATSDR 2000). Geometric mean serum PCB levels in these populations ranged from 0.9 to 1.5 ppb (μ g/L), with a range among individuals in these populations of 0.46 to 9.5 ppb (ATSDR 2000). Among the adult participants in Pittsfield, the geometric mean serum PCB level (all congeners, whole weight) was 1.150 ppb, with a maximum concentration of 3.595 ppb. CDC reported their whole weight PCB concentrations on a ng/g basis. Converting to μ g/L, for comparison to the ATSDR reported data, results in a geometric mean of 1.18 ppb, with a max of 3.688 ppb. Thus, the Pittsfield adult participants had serum PCB levels consistent with data cited in ATSDR 2000.

MDPH further evaluated information related to the individual with the highest detected serum level, 3.595 ppb (sum of all congeners, whole weight). This individual reported occupational exposure, was in the oldest age group of all participants (>50 years old), reported living in a residence near GE, and reported being a fish eater. Hence, although the serum PCB level in this individual is still consistent with national data (reported as the sum of 15 congeners), the individual did report several factors that likely contributed to his PCB exposure (e.g., age, occupational exposure).

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CONCLUSIONS /

Results from indoor environmental and serum PCB testing at the Allendale School did not appear to reveal unusual opportunities for PCB exposures to the Allendale School community or to other participants in the serum PCB testing. Although PCBs were detected in some indoor environmental samples, with one exception (of 98 samples), no detection of either Aroclors or congeners exceeded any available screening guideline or regulatory standards. The one exception was a slight exceedance (0.144 μ g/wipe) for a sample taken from a windowsill in the gymnasium located 10 feet above floor level. This sample result was, however, well below the USEPA cleanup standard for determining whether more aggressive cleaning may be needed for a surface.

Although 5 of 98 samples slightly exceeded at least one available guideline assuming non-detectable Aroclors or congeners were assumed to be present at one-half the detection limit, these concentrations under the most conservative exposure assumptions would not be expected to result in health effects. In addition, levels detected in the Allendale School were generally lower than those reported in other studies in Massachusetts, North Carolina, and Great Britain for indoor environments, including schools, day care centers, and homes. Finally, levels detected for indoor air in the school were below health-based screening values and consistent with historical data that show that PCBs are more frequently detected during warmer months in outdoor air samples at concentrations slightly higher than during colder months of the year.

Serum PCB testing conducted using state-of-the-art analytical techniques by the U.S. CDC showed that the current Allendale students (participants ages 8-10 years) were well below available national data for children ages 12-19 years old. In addition, comparison with available data for children in the scientific literature also revealed that the Allendale children had lower serum PCB levels compared to those reported in the literature. Adult participants, including current Allendale School staff, also showed typical serum PCB levels based on the national NHANES data, including the fact that there was a trend of serum PCB levels increasing with age, a well-established trend for serum PCBs. The

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median concentration in Pittsfield adults (0.918 ppb whole weight) was less than the comparable NHANES value (1.062 ppb) and all adults had levels within the NHANES 90th percentile (3.099 ppb whole weight).

CDC reported that "Results of the analyses of the Pittsfield participants revealed that the Pittsfield participants showed low PCB levels on either a whole weight basis or on a lipid-adjusted congener basis as compared with the third National Report data." In addition, CDC evaluated the congener pattern seen in the Pittsfield participants and reported the following: "This review did not reveal any unusual patterns among the Pittsfield participants to suggest that exposures that may have led to any evidence of PCBs in blood samples are different from the U.S. population." Given the small numbers of participants, the MDPH/BEH could not speak conclusively about PCB serum levels for those who were not actually tested, however the environmental data would suggest that elevated serum PCB levels would not be likely based on attendance/occupation associated with the Allendale School.

RECOMMENDATIONS

The following recommendations are provided:

- 1. MDPH/BEH recommends that more aggressive cleaning of surfaces not routinely cleaned (e.g., windowsills) be undertaken and regularly conducted.
- 2. MDPH/BEH will respond to any public comments received on this public comment release report and prepare a final report that includes responses to all comments received.
- 3. At the request of the Pittsfield Board of Health and/or community residents MDPH/BEH will evaluate any ambient air results of testing being conducted by the US EPA that may be of concern.

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Sample Type	Locations
	Classroom 19, Classroom 23,
	Classroom 24, Classroom 27,
	Classroom 28, Classroom 32,
Surface Wipe	Hallway Outside Classroom 15,
$(\mu g/100 \text{ cm}^2)$	Hallway Outside Classroom 21,
	Hallway Outside Classroom 29,
	Hallway Outside Gymnasium,
	Health Office, Gymnasium
Air	Classroom 21, Classroom 28,
$(\mu g/m^3)$	Outside between Classrooms 23 and 24
Carpet Surface Dust (mg/kg)	Classroom 19
Vacuum Bag (mg/kg)	Entire School
Unit Ventilator Filter	Classroom 21, Classroom 24

Table 1: Sample Locations for Environmental Testing

Sample Type	Analysis	Laboratory	Method Detection Limit
		SAI	0.05
Surface Wipe	Aroclor –	SWRI	0.050
(µg/wipe)	G	SUNY	0.00028
	Congener –	SWRI	0.0050
	A	SAI	0.000011
Air	Aroclor	SWRI	0.000067
$(\mu g/m^3)$	Comment	SUNY	0.0000093
	Congener	SWRI	0.0000058
	Aroclor	SAI	0.047
Carpet Surface Dust	Arocior	SWRI	0.100
(mg/kg)	Conconor	SUNY	0.00025
	Congener –	SWRI	0.0050
	Aroclor	SAI	0.250
Vacuum Bag	ATOCIOI	SWRI	0.0994
(mg/kg)	Congener	SUNY	0.00025
	Congener –	SWRI	0.0098
	Aroclor	SAI	0.103
Unit Ventilator	AIOCIOI	SWRI	0.050
Filter		SUNY	0.00013
(mg/kg)	Congener	SWRI	0.0025

Table 2: Method Detection Limits for Environmental Testing

Agency	Soil	Surface wipe	Air
ATSDR	CREG = 0.4 mg/kg		CREG = $0.01 \mu g/m^3$
	Chronic EMEG =1 mg/kg for children; 10 mg/kg for adults		one of onor µg/m
MDEP	2 mg/kg		
EPA		10 µg/100cm ²	
California		$0.1 \mu g / 100 cm^2$	

Table 3a: Environmental PCB Guidelines Used by MDPH to Evaluate Data

CREG- Cancer Risk Evaluation Guide EMEG- Environmental Media Evaluation Guide

Location	Analysis	Laboratory	QA/QC Notes	Result (ND=0)	Result (half ND)	Comparison Values
Air (µg/m3)						
	Angelen	SAI		0.000566	0.000654	
D 01/45	Aroclor	SWRI		0.00181	0.00220	CREG-0.01 µg/m ³
Room 21 (A)	0	SUNY		0.00688	0.00704	CREG-0.01 µg/m
	Congener	SWRI	50% J	0.000450	0.000934	
		SAI		0.000588 .	0.000676	
D 01 (D)	Aroclor	SWRI		0.00180	0.00219	
Room 21 (B)	0	SUNY		0.00491	0.00507	
	Congener	SWRI	40% J -	0.000450	0.000964	
	Aroclor	SAI		0.000590	0.00069	
D 00 (A)		SWRI		0.00232	0.00277	
Room 28 (A)		SUNY		0.0112	0.0114	
	Congener	SWRI	50% J	0.000988	0.00135	
	4 1	SAI		0.000642	0.000734	
D 00 (D)	Aroclor	SWRI	•	0.00291	0.00333	
Room 28 (B)	0	SUNY		0.00864	0.00879	
	Congener	SWRI	40% J	0.00109	0.00148	
		SAI		0.000412	0.000496	
	Aroclor	SWRI		0.00143	0.00181	
Outside (A)		SUNY		0.0116	0.0117	
	Congener	SWRI	70% J	0.000392	0.000707	

Table 3b: Sample Results for Environmental Testing

Location	Analysis	Laboratory	QA/QC Notes	Result (ND=0)	Result (half ND)	Comparison Values
	Aroclor	SAI		0.000341	0.000425	
Outside (B)		SWRI		0.00152	0.00190	
	Canadanan	SUNY		0.00779	0.00793	
	Congener	SWRI	70% J	0.000432	0.000751	
	Aroclor	SAI		ND (0.020 µg/PUF)	ND (0.020 µg/PUF)	
Field Blank 1		SWRI		ND (0.020 µg/PUF)	ND (0.020 µg/PUF)	
Field Blank I	Canada	SUNY	Detections in field blank	1.600 μg/mL	1.885 μg/mL	
	Congener	SWRI		ND (0.0002 µg/SPL)	ND (0.0002 µg/SPL)	
	Aroclor	SAI		ND (0.020 µg/PUF)	ND (0.020 µg/PUF)	
Field Blank 2		SWRI		ND (0.020 µg/PUF)	ND (0.020 µg/PUF)	
Field Blank 2	Canada	SUNY	Detections in field blank	0.699 μg/mL	1.029 μg/mL	
	Congener	SWRI		ND (0.0002 µg/SPL)	ND (0.0002 µg/SPL)	
	Aroclor	SAI		ND (0.020 µg/PUF)	ND (0.020 µg/PUF)	
Matrix Blank 1		SWRI		ND (0.020 µg/PUF)	ND (0.020 µg/PUF)	
Maulx Blank I	Canada	SUNY	Detections in matrix blank	0.247 μg/mL	0.610 μg/mL	
	Congener	SWRI		ND (0.0002 µg/SPL)	ND (0.0002 µg/SPL)	
	Aroclor	SAI		ND (0.020 µg/PUF)	ND (0.020 µg/PUF)	
Moterin Disals 2		SWRI		ND (0.020 µg/PUF)	ND (0.020 µg/PUF)	
Matrix Blank 2	0	SUNY	Detections in matrix blank	0.505 μg/mL	0.832 μg/mL	
	Congener	SWRI		ND (0.0002 µg/SPL)	ND (0.0002 µg/SPL)	
urface Wipes (µg/wij	pe)					
Room 19 (A)	Aroclor	SAI		ND (0.05)	ND (0.05)	CDTSC- 0.1 µg/wip

Location	Analysis	Laboratory	QA/QC Notes	Result (ND=0)	Result (half ND)	Comparison Value
		SWRI		ND (0.05)	ND (0.05)	
	0	SUNY		0.000150	0.0126	
	Congener	SWRI		ND (0.005)	ND (0.005)	
	Aroclor	SAI		ND (0.05)	ND (0.05)	
Room 19 (B)		SWRI		ND (0.05)	ND (0.05)	
K00III 19 (D)	Canaanan	SUNY		ND (0.00014)	ND (0.00014)	
	Congener	SWRI		ND (0.005)	ND (0.005)	
	Aroclor	SAI		ND (0.05)	ND (0.05)	
Room 19 (C)		SWRI		ND (0.05)	ND (0.05)	
Koom 19 (C)	Congener	SUNY		0.000230	0.0127	
	Congener	SWRI		ND (0.005)	ND (0.005)	
	Aroclor	SAI		ND (0.05)	ND (0.05)	
Room 19 (D)		SWRI		ND (0.05)	ND (0.05)	
Koom 19 (D)	Congener	SUNY		0.000130	0.0126	
	Congener	SWRI		ND (0.005)	ND (0.005)	and the second second
	Aroclor	SAI		ND (0.05)	ND (0.05)	
Room 24 (A)		SWRI		ND (0.05)	ND (0.05)	
100111 24 (A)	Concense	SUNY		0.00188	0.0139	
	Congener	SWRI		ND (0.005)	ND (0.005)	
Room 24 (B)	Aroclor	· SAI		ND (0.05)	ND (0.05)	
		SWRI		ND (0.05)	ND (0.05)	
	Congener	SUNY		ND (0.00014)	ND (0.00014)	

Location	Analysis	Laboratory	QA/QC Notes	Result (ND=0)	Result (half ND)	Comparison Value
		SWRI		ND (0.005)	ND (0.005)	
	Aroclor	SAI		ND (0.05)	ND (0.05)	
Room 24 (C)		SWRI		ND (0.05)	ND (0.05)	
K0011 24 (C)	Conconor	SUNY		0.00020	0.0126	
	Congener	SWRI		ND (0.005)	ND (0.005)	
	Aroclor	SAI		ND (0.05)	ND (0.05)	
Room 24 (D)		SWRI		ND (0.05)	ND (0.05)	
K00III 24 (D)	Congonar	SUNY		0.00011	0.0126	
	Congener	SWRI		ND (0.005)	ND (0.005)	
	Aroclor	SAI		ND (0.05)	ND (0.05)	
Room 28 (A)		SWRI		ND (0.05)	ND (0.05)	
K00III 28 (A)	Canada	SUNY		0.00350	0.0153	
	Congener	SWRI		ND (0.005)	ND (0.005)	
	Aroclor	SAI		ND (0.05)	ND (0.05)	
Room 28 (B)		SWRI		ND (0.05)	ND (0.05)	
K00III 28 (B)	Congener	SUNY		0.00030	0.0127	
	Congener	SWRI		ND (0.005)	ND (0.005)	
	Aroclor	SAI		ND (0.05)	ND (0.05)	
Room 28 (C)		SWRI		ND (0.05)	ND (0.05)	
	Conserve	SUNY		0.00030	0.0127	
	Congener	SWRI		ND (0.005)	ND (0.005)	
Room 28 (D)	Aroclor	SAI		ND (0.05)	ND (0.05)	

Location	Analysis	Laboratory	QA/QC Notes	Result (ND=0)	Result (half ND)	Comparison Value
		SWRI		ND (0.05)	ND (0.05)	
	Canana	SUNY		0.0010	0.0134	
	Congener	SWRI		ND (0.005)	ND (0.005)	
	Aroclor	SAI		ND (0.05)	ND (0.05)	
Room 32 (A)		SWRI		ND (0.05)	ND (0.05)	
R00III 52 (A)	Canaanan	SUNY		0.00070	0.0130	
	Congener	SWRI		ND (0.005)	ND (0.005)	
	Aroclor	SAI		ND (0.05)	ND (0.05)	
Room 32 (B)		SWRI		ND (0.05)	ND (0.05)	
K00III 52 (B)	Congener	SUNY		0.00108	0.0134	
	Congener	SWRI		ND (0.005)	ND (0.005)	Service Service
	Aroclor	SAI		ND (0.05)	ND (0.05)	1.
Room 32 (C)		SWRI		ND (0.05)	ND (0.05)	
R0011 52 (C)	Congener	SUNY		0.00197	0.0141	
	Congener	SWRI		ND (0.005)	ND (0.005)	
	Aroclor	· SAI		ND (0.05)	ND (0.05)	
Room 32 (D)		SWRI		ND (0.05)	ND (0.05)	1990 - A.
(D)	Congonor	SUNY		0.00028	0.0127	
	Congener	SWRI		ND (0.005)	ND (0.005)	
Hall (A)	Aroclor	SAI		ND (0.05)	ND (0.05)	
		SWRI		ND (0.05)	ND (0.05)	
	Congener	SUNY		0.00009	0.0126	

Location	Analysis	Laboratory	QA/QC Notes	Result (ND=0)	Result (half ND)	Comparison Values
		SWRI		ND (0.005)	ND (0.005)	
	Aroclor	SAI		ND (0.05)	ND (0.05)	
Hall (B)		SWRI		ND (0.05)	ND (0.05)	
Hall (B)	Concensor	SUNY		0.00009	0.0126	
	Congener	SWRI		ND (0.005)	ND (0.005)	
	Aroclor	SAI		ND (0.05)	ND (0.05)	
Hall (C)		SWRI		ND (0.05)	ND (0.05)	
Hall (C)	Congener	SUNY		ND (0.00014)	ND (0.00014)	
	Congener	SWRI		ND (0.005)	ND (0.005)	
	Aroclor	SAI		ND (0.05)	ND (0.05)	
Hall (D)		SWRI		ND (0.05)	ND (0.05)	
Hall (D)	Congener	SUNY		0.00023	0.0127	
		SWRI		ND (0.005)	ND (0.005)	
	Aroclor	SAI		ND (0.05)	ND (0.05)	
Gymnasium		SWRI		0.144	0.294	
Windowsill	Canganan	SUNY		0.04003	0.0467	
	Congener	SWRI	75% J	0.0702	0.280	· · · · · · · · · · · · · · · · · · ·
	Aroclor	SAI		ND (0.05)	ND (0.05)	
Health Office Vent		SWRI		ND (0.05)	ND (0.05)	
ricann Onice vent	Conserve	SUNY		0.0132	0.0218	
	Congener	SWRI	100% J	0.0061	0.259	
Room 23 Pipe	Aroclor	SAI		ND (0.05)	ND (0.05)	

Location	Analysis	Laboratory	QA/QC Notes	Result (ND=0)	Result (half ND)	Comparison Values
		SWRI		ND (0.05)	ND (0.05)	
	C	SUNY		ND (0.0125)	ND	
	Congener	SWRI		ND (0.005)	ND (0.005)	
	Aroclor	SAI		ND (0.05)	ND (0.05)	
Room 24 Storage Bin Cover		SWRI		ND (0.05)	ND (0.05)	
Cover	0	SUNY		0.00154	0.0135	
	Congener	SWRI		ND (0.005)	ND (0.005)	
	Aroclor	SAI		ND (0.05)	ND (0.05)	
Room 24 Unit		SWRI		ND (0.05)	ND (0.05)	
Ventilator	Congener	SUNY		0.00137	0.0135	
		SWRI		ND (0.005)	ND (0.005)	
	Aroclor	SAI		ND (0.05)	ND (0.05)	
Room 27 Ceiling Fan		SWRI		ND (0.05)	ND (0.05)	
Blade	Congener	SUNY		0.00193	0.0139	
	Congener	SWRI		ND (0.005)	ND (0.005)	
	Aroclor	SAI		ND (0.05)	ND (0.05)	
Room 28 VCR		SWRI		ND (0.05)	ND (0.05)	
Room 28 VCR	C	SUNY		0.00214	0.0142	
	Congener	SWRI		ND (0.005)	ND (0.005)	All States and
Field Blank	Aroclor	SAI		ND (0.05)	ND (0.05)	
		SWRI		ND (0.05)	ND (0.05)	
	Congener	SUNY		ND (0.00014)	ND (0.00014)	

Location	Analysis	Laboratory	QA/QC Notes	Result (ND=0)	Result (half ND)	Comparison Values
		SWRI		ND (0.005)	ND (0.005)	
Carpet Dust (mg/kg	g)					
State State	Aroclor	SAI		0.0592	0.238	Chronic Child EMEG
Room 19		SWRI		ND (0.100)	ND (0.100)	(for soil)- 1 mg/kg Chronic Adult EMEC
Room 19	Congener	SUNY		0.108	0.111	(for soil)- 10 mg/kg
	Congener	SWRI	100% J	0.0762	0.526	CREG (for soil)- 0.4 mg/kg
Vacuum Bags (mg/	kg)					
	Aroclor	SAI		0.285	1.285	Chronic Child EMEG
Bag 1		SWRI		ND (0.099)	ND (0.099)	(for soil)- 1 mg/kg Chronic Adult EMEC
Dag I	Congonar	SUNY		0.0513	0.0559	(for soil)- 10 mg/kg
	Congener	SWRI	80% J	0.0687	0.534	CREG (for soil)- 0.4 mg/kg
	Aroclor	SAI		0.292	1.292	
Bag 2		SWRI		ND (0.099)	ND (0.099)	
Dag 2	Congener	SUNY		0.0666	0.0709	
	Congener	SWRI	90% J	0.0601	0.502	
Unit Ventilator Filt	ters (µg/vent)					
	Aroclor	SAI		ND (99.8)*	ND (99.8)*	
Room 21		SWRI		0.0743	0.224	
Room 21	Concerner	SUNY		0.0467	0.0519	N/A
	Congener	SWRI	100% J	0.0288	0.259	
Room 24	Aroclor	SAI		ND (106)*	ND (106)*	
		SWRI		0.105	.0.255	

Location	Analysis	Laboratory	QA/QC Notes	Result (ND=0)	Result (half ND)	Comparison Values
	Congener	SUNY		0.0742	0.0786	
	Congener	SWRI	80% J	0.0907	0.278	
	Aroclor	SAI		ND (87)*	ND (87)*	
Field Blank		SWRI		ND (0.05)	ND (0.05)	
Field Blank	Congener	SUNY	Detections in field blank	0.00294	0.0155	
	Congener	SWRI		ND (0.005)	ND (0.005)	
NIST SRM (mg/kg)						
Prepared Sample		SUNY	81 congeners detected	0.39	0.40	
	Congener	SWRI	41 congeners detected; 60% J	0.4463	0.7521	

SAI reported filters in µg/kg.

Sample Type	Analysis	Detects/Samples	QA/QC Notes	Max *	Max**	Screening Value	
Surface Wipe	A-SAI	0/27	None	the set are too are	AND 160 100 100 100		
$(\mu g/100 \text{ cm}^2)$	A-SWRI	1/27	None	0.144	0.294	0.1 (CDTSC)	
	C-SWRI	2/27	90% J	0.07	0.28	10 (USEPA)	
	C- SUNY	23/27	None	0.040	0.047	IU (USEPA)	
				Lagard			
Indoor Air	A-SAI	4/4	None	0.00064	0.00073		
$(\mu g/m^3)$	A-SWRI	4/4	None	0.00291	0.00333	0.01 (ATSDR	
	C-SWRI	4/4	50% J	0.00109	0.00148	CREG)	
	C- SUNY	4/4	Blank detects	0.0112	0.0114		
	1			1000011	0.00050		
Outdoor Air	A-SAI	2/2	None	0.00041	0.00050		
$(\mu g/m^3)$	A-SWRI	2/2	None	0.00091	0.0019	0.01 (ATSDR CREG)	
	C-SWRI	2/2	50% J	0.00041	0.00075		
	C-	2/2	Blank	0.0116	0.0117		
	SUNY		detects				
Carpet Dust	A-SAI	1/1	None	0.059	0.238	0.4 (ATSDR CRE	
(mg/kg)	A-SWRI	0/1	None			1 (ATSDR Child	
(C-SWRI	1/1 .	90% J	0.076	0.526	EMEG)	
	C- SUNY	1/1	None	0.108	0.111	10 (ATSDR Adu EMEG) 2 (MDEP)	
Vacuum Dust	A-SAI	2/2	None	0.292	1.29	0.4 (ATSDR CRE	
(mg/kg)	A-SAI A-SWRI	0/2	None	0.292	1.47	1 (ATSDR Child	
(mg/kg)	C-SWRI	2/2	90% J	0.0687	0.534	EMEG)	
	C-SWRI C-	2/2	None	0.066	0.07	10 (ATSDR Adu	
	SUNY	LIL	None	0.000	0.07	EMEG)	
	SUNT			1		2 (MDEP)	
		0/0					
Unit Ventilator Filter	A-SAI	0/2	None	0.105	0.055		
(µg/sample)	A-SWRI	2/2	None	0.105	0.255	NUA	
	C-SWRI	2/2	75% J	0.0907	0.268	N/A	
	C- SUNY	2/2	None	0.0742	0.08		

Table 4: Summary of Indoor Environmental Testing Results

A-SAI = Aroclor analysis by SAI; also A-SWRI C-SWRI = congener analysis by SWRI; also C-SUNY Max* = maximum concentration assuming non-detected Aroclors or congeners were zero.

Max** = maximum concentration assuming non-detected Aroclors or congeners were present at ½ detection limit. CREG = Cancer Risk Evaluation Guide

Adult/Child EMEG = Environmental Media Evaluation Guide for Adult/Children (non-cancer effects) J = Estimated concentration below the method detection limit

CDC METHOD DETECTION LIMITS* PPB (Lipid Adjusted)					
		NHANES III	PITTSFIELD		
	PCB Congener	(All Ages)	(8-59yo)		
1	PCB18	NR	NR		
2	PCB28	32.4	2.9		
3	PCB52	12.4	1.4		
4	PCB49	NR	1.4		
5	PCB44	NR	0.7		
6	PCB74	10.5	0.7		
7	PCB66	12.4	1.4		
8	PCB101	10.5	0.7		
9	PCB99	10.5	0.7		
10	PCB87	10.5	0.7		
11	PCB110	10.5	0.7		
12	PCB118	10.5	0.7		
13	PCB105	10.5	0.7		
14	PCB151	10.5	0.7		
15	PCB149	10.5	0.7		
16	PCB146	10.5	0.7		
17	PCB153	10.5	0.7		
18	PCB138-158	10.5	0.7		
19	PCB128	10.5	0.7		
20	PCB167	10.5	0.7		
21	PCB156	10.5	0.7		
22	PCB157	10.5	0.7		
23	PCB178	10.5	0.7		
24	PCB187	10.5	0.7		
	PCB183	10.5	0.7		
	PCB177	. 10.5	0.7		
27	PCB172	10.5	0.7		
	PCB180	10.5	0.7		
29	PCB170	10.5	0.7		
30	PCB189	10.5	0.7		
31	PCB199	10.5	0.7		
32	PCB196-203	10.5	0.7		
33	PCB195	28.1	0.7		
34	PCB194	10.5	0.7		
35	PCB206	28.1	0.7		
36	PCB209	NR	0.7		

Table 5 (Serum PCB Testing)

NR= Not Reported

*Detection limits vary with samples size. Maximum detection limits among the samples analyzed are reported in this table for both NHANES and Pittsfield.

A	*Currently Affiliated with AS			Not Currently Affiliated with AS			Total Participants		
Age (years)	Male	Female	Subtotal	Male	Female	Subtotal	Male	Female	Totals
0-19	4	. 3	7	2	5	· . 7	6	8	14
20-59	2	8	10	5	· 3	8	7	11	18
Total	6	11	17	7	8	15	13	19	32

Table 6: Distribution of Participants (Serum PCB Testing)

* Students, Parents, Staff

SUM OF CONGENERS (15) 52, 74, 99, 180, 187, 194,	
Pittsfield Serum Samples n=14 (AGES 8-19)	NHANES Serum Samples (AGES 12-19)
SUM OF PCBs - WI	hole Weight (ppb)
MEDIAN	MEDIAN/50th Percentile
0.121	0.345
SUM OF PCBs - Lip	oid Adjusted (ppb)
25.7	71.8

Table 7: Summary of Total PCB Concentrations for Children

SUM OF CONGENERS (15) 52, 74, 99, 180, 187, 194,	
Pittsfield Serum Samples n=18 (AGES 20-59)	NHANES Serum Samples (AGES 20+)
SUM OF PCBs - W	hole Weight (ppb)
MEDIAN	MEDIAN/50th Percentile (CI)
0.918	1.062 (0.968,1.177)
SUM OF PCBs - Lip	pid Adjusted (ppb)
176.1	168.5 (154.7,184.2)

Table 8: Summary of Total PCB Concentrations for Adults

CI = Confidence Interval

Table 9: Comparison of Total* PCB Serum Levels for Current Allendale Students

CHILDREN Serum PCB Levels	ALLEN MED		NHANES MEDIAN/50 TH PERCENTILE		
	WHOLE WEIGHT	LIPID ADJUSTED	WHOLE WEIGHT	LIPID ADJUSTED	
Current Allendale Students 8-10yo. (n=7)	0.117 ppb	25.2 ppb			
Allendale graduates and other community members 12-19yo (n=7)	0.141 ppb	26.2 ppb	0.345 ppb (0.340,0.362)	, 71.8 ppb (69.1,74.2)	

*The total of 15 most frequently detected PCB congeners in the population.

The upper and lower confidence intervals for serum PCB levels are depicted in parenthesis in the column labeled NHANES MEDIAN/50th PERCENTILE.









Figure 4



Serum PCB Congener Patterns for Allendale Community

Figure 5





APPENDIX A: INDOOR ENVIRONMENTAL TESTING PROTOCOL

PROTOCOL FOR INDOOR ENVIRONMENTAL SAMPLING ALLENDALE ELEMENTARY SCHOOL PITTSFIELD, MASSACHUSETTS

Allendale Indoor PCB Environmental Sampling Workgroup

October 2006

INTRODUCTION

In order to address concerns about exposure opportunities to polychlorinated biphenyls (PCBs) at the Allendale Elementary School, the Allendale Indoor PCB Environmental Sampling Workgroup (workgroup) was formed. The workgroup is composed of representatives from the MA Department of Public Health's Center for Environmental Health Environmental Toxicology Program (MDPH/BEH/ETP); MA Department of Environmental Protection's Bureau of Waste Site Cleanup (MDEP); Pittsfield Board of Health; State University of New York at Albany's Institute for Health and the Environment (SUNY); Spectrum Analytical, Inc. (SAI); Southwest Research Institute (SWRI); Allendale Elementary School; the Housatonic River Initiative; and the Allendale School Task Force. The U.S. Environmental Protection Agency (USEPA) is attending meetings and providing technical assistance to the workgroup. MDEP has been informed and has discussed the sampling activities with BEH/ETP. The charge of the workgroup is to develop an indoor environmental sampling and analysis plan for this follow-up effort at the school.

GOAL

The overall goal of the proposed sampling effort is to determine whether PCBs are present in the indoor environment of the Allendale Elementary School in areas where children, faculty, and staff may have opportunities for exposure; and to determine if health concerns are present and whether follow-up activities are warranted.

OBJECTIVES

Specific objectives are to collect and analyze samples for PCBs utilizing both congener specific and Aroclor based standard methods. These samples will include: indoor air (with an outdoor comparison sample), surface wipes, carpet surface dust, vacuum bag dust, and unit ventilator filters. The sampling objectives will serve to address several questions, including the following:

- 1. Are detectable concentrations of PCBs present in the indoor environment of the school?
- 2. Are detectable concentrations of PCBs present in areas of the school accessible to students and staff?
- 3. If PCBs are present in the school, could the concentrations present exposure opportunities or health concerns?

LOCATION OF SAMPLES

Air Samples:

<u>Location of Samples</u>: Samples will be collected from inside the school in, or in the vicinity of classrooms #21 and 28 and outside the school in the building nook between classrooms #23 and 24 (see Figure).

<u>Rationale</u>: Classroom #21 is the middle classroom in the new building wing and #28 is the middle classroom in the original building, both of which face the back of the school and the GE disposal area. Thus, both classrooms are representative of the two wings of the building and are on the side of the school that faces the landfills. Classroom #28 has water damaged ceiling tiles, which could provide an entry point for unfiltered outside air (MDPH 2005). Wind coming from the direction of the GE disposal area will likely pool in the area where the two wings of the school meet, which is between classrooms #23 and 24. Sampling at this location would likely constitute the highest PCBs concentrations, if any, near the school.

Wipe Samples:

Location of Samples: Wipe samples will be taken to obtain a representative picture of possible concentrations of PCBs on frequently and infrequently touched hard surfaces inside the school. Samples will be collected from selected classrooms on the rear-side of the building, which faces the GE disposal site, as well as hallway corridors, the gymnasium, and several locations to be chosen on the day of sampling. In selected classrooms, samples will be collected from one windowsill and a wall on the opposite side of the classroom from the windows, representing frequently touched areas, and a window pane and the top shelf of a bookshelf, representing infrequently touched areas. The selected classrooms are #19, 24, 28, and 32. Four wipe samples will be collected from the two corridors that span the length of the two building wings. The wipe samples will be collected from one location on each side of the corridor, in each wing, above the area that is normally cleaned (approximately five feet). Wipe samples will be collected from the top of 1 or 2 hanging ceiling lights (depending on accessibility) that are located in the gymnasium. The hallway corridor and gymnasium samples represent infrequently touched surfaces (see Figure). Finally, several wipe samples will be collected from locations chosen during the sampling event.

<u>Rationale:</u> PCBs that could potentially enter the school through the air could potentially be bound to dust particles and settle onto surfaces. Therefore, collecting wipe samples from specific locations within the school will provide information on whether PCBs are present. Specific classrooms within the school were chosen based on information gathered during the MDPH/BEH site visit in November 2005 and contained in the MDPH/BEH Emergency Response/Indoor Air Quality Program's report, "Indoor Air Quality Assessment: Allendale Elementary School." All of the classrooms were chosen because they face the Hill 78 disposal area, they represent classrooms distributed along the entire length of the building facing the Hill 78 disposal area, and they have waterstained ceiling tiles. The water stains could be from leaks in the roof, which are a potential route for outside air to enter the classroom without passing through the unit ventilator filters (MDPH 2005). Open classroom windows and doors, possibly during warmer months or to let children outside, are also routes for unfiltered air to enter the classrooms. The ceiling lights in the gymnasium represent an area that is likely infrequently touched, except for an occasional light bulb change. Some work group members expressed concern that dust from the lights could become airborne or fall to the gymnasium floor during times of building activity/vibration.

Unit Ventilator Filter Samples:

Location of Samples: Each unit ventilator has three filters which lie in a row parallel to each other (i.e., the air passes through this row of filters) (MDPH 2005). For consistency, samples of unit ventilator filters will be collected from the center filter in each unit ventilator. Samples will be collected from classrooms # 21 and 24 (see Figure). These classrooms face the back of the school and the Hill 78 disposal area.

<u>Rationale</u>: Each occupied classroom within the school is provided heat and outside air by a unit ventilator. The unit ventilators intake air from both outside and from inside the room, mix it, and then vent it into the room. During cooler months, the unit ventilator heats the air before venting it into the room. During warmer months, the unit ventilator provides a source of outside air. Before air is vented into the room, it passes through a filter, which is intended to capture dust particles. The unit ventilator filters capture dust particles before they enter the classroom as they draw air in from the outside and they also capture dust particles that are inside the classroom by recirculating classroom air. PCBs that have attached to dust particles may become trapped in the filters. The classrooms were chosen because they face the Hill 78 disposal area. Classroom #24 was specifically selected because it is located in the building nook, where the two wings of the school meet. It is theorized that wind blowing from the Hill 78 disposal area towards the school would pool in the nook due to the shape of the building.

Carpet Surface Dust Samples:

Location of Samples: Samples will be collected from classroom #19, a kindergarten room (see Figure).

Rationale:

- 1. Carpet can retain dust on its surface as individuals walk on it and airborne particles settle onto it.
- 2. Individuals can come into contact with this dust while touching or playing on the carpet.
- 3. The classroom faces the GE disposal area, and has a water damaged ceiling tile, which could provide an entry point for unfiltered outside air (MDPH 2005).

Vacuum Bag Sample:

Location of Sample: Dust samples will be collected from a vacuum cleaner that is operated throughout the entire school.

<u>Rationale</u>: Dust settles throughout the entire school. The vacuum cleaner collects dust that settles on the floor, along with any possible PCBs. Sampling the contents of the vacuum cleaner bag will provide information on whether PCBs are present in the floor dust.

TIMING OF SAMPLE COLLECTION

<u>Description:</u> Samples will be collected during the week of June 12, 2006 (see Sample Packaging and Transport/Chain of Custody section for information on sample possession). The vacuum cleaner bag sample will be collected after the vacuum cleaner has been used for one school week (i.e., 5 days). Carpet surface dust, wipe and unit ventilator filter samples will be collected during a single school day during that week. Air samples will begin to be collected during the same school day as carpet surface dust, wipe, and unit ventilator filters (the machines run for 24 hours). Air samples need to be collected during active operation of the landfills and on a warm dry weather day, preferably after a period of wet weather. Air samples will be collected during two distinct sampling rounds on two different days.

<u>Rationale</u>: PCBs can become airborne through a process of volatilization. This process can be increased when PCB-contaminated soil dries, as more PCBs enter the atmosphere (ASTDR 2000). The months of May and June typically involve periods of wet weather, followed by periods of dry, warm weather. Measurements of PCBs during this time period would likely be representative of the highest rates of PCB volatilization from Hill 78.

SAMPLE COLLECTION METHODS

Air Samples:

<u>Description</u>: All samples will be collected by a trained technician with Environmental Compliance Services (ECS), an environmental consulting firm. Samples will be collected by following USEPA Method TO-4A. This method involves using a high volume sampler, which is a box-like structure that contains a motor and a cartridge, to collect and filter air onto a sorbent cartridge for 24 hours. The cartridge is then placed into a sterile glass jar, which is placed in a cooler.

<u>Rationale</u>: USEPA Method TO-4A is the standard method for collecting and analyzing air samples for PCBs. One sample will be collected from each location for each of the two sampling rounds and split by SWRI after being extracted into a solution. Co-located samples cannot be collected due to the logistics of collecting the air samples (e.g., shipping the equipment, running several loud machines in classrooms during the school day). The analytical methods require that the samples be cooled after collection and prior to analysis.

Wipe Samples:

<u>Description</u>: All samples will be collected by a trained technician with ECS. Wipe samples will be collected using a National Institute for Occupational Safety and Health (NIOSH) surface wipe method. This method involves wetting an absorbent pad with hexane, wiping a 10 centimeter x 10 centimeter area horizontally, vertically, horizontally again, and placing the pad in a sterile glass jar. Three co-located samples will be collected from each sample location (i.e., samples will be collected from an area adjacent to each other). The jars will be placed into a cooler.

<u>Rationale</u>: ECS technicians have been trained to collect environmental samples, including wipe samples. The SAI method ensures that any PCBs will become attached to the absorbent pad. This method is similar to a USEPA Collection method included in the Toxic Substances Control Act regulations (40 Code of Federal Regulations 761.123). In order to produce three samples from each sample location for the three laboratories to analyze, co-located samples will be collected. Three samples cannot be collected from the same location because the sample collection method is intended to remove all possible PCBs from the location after the first wipe.

Unit Ventilator Filter Samples:

<u>Description</u>: All samples will be collected by a trained technician with ECS. Samples will be collected by using the following method: using sterile gloves and a pair of scissors, a 1"x 10" section of the middle filter will be removed from three edges of the unit ventilator filter and placed into separate sterile glass jars. The scissors will be wiped with hexane between samples. The jars will be placed into a cooler.

<u>Rationale</u>: The unit ventilators contain three filters, which are installed with metal spaces that prevent air from bypassing the filters (MDPH 2005). Due to this design, the air should have an equal probability of passing through each of the filters. The middle filter and clippings from the three edges were chosen simply to be consistent. One clipping will be analyzed by each of the three laboratories. While there is no available USEPA sample collection method for unit ventilator filter samples, the analytical methods require that the samples be cooled after collection and prior to analysis.

Carpet Surface Dust Samples:

<u>Description</u>: All samples will be collected by a trained technician with ECS. The samples will be collected according to a method developed by the American Society for Testing and Materials International (ASTM) (i.e., ASTM D5438-00). The carpet will be divided into quadrants and a sample will be collected from three of the quadrants. The samples will be placed into a cooler.

<u>Rationale</u>: In order to produce three samples for the three laboratories to analyze, the carpet needs to be divided into sections. Three samples cannot be collected from the

same location on the carpet because the sample collection method is intended to remove all possible surface dust after the first vacuuming. The analytical methods require that the samples be cooled after collection and prior to analysis.

Vacuum Bag Sample:

<u>Description</u>: The school vacuum cleaner will be operated in a normal fashion by the custodian. At the end of the week, the vacuum bag will be removed from the vacuum, placed into a cooler, and sent overnight delivery to SWRI, where the dust will be separated for the three laboratories, according to the previously agreed upon SOPs.

<u>Rationale</u>: The custodian vacuums the school daily. The purpose of sampling the vacuum cleaner bag is to determine the levels of PCBs that may be present in the dust throughout the school. ECS staff will retain chain of custody of the vacuum at all times during the test week.

SAMPLE PACKAGING AND TRANSPORT/CHAIN OF CUSTODY

<u>Description</u>: ECS staff will maintain possession of the samples during and after collection. ECS will maintain possession of the school's vacuum cleaner when it is not in use and will be present when it is in use. Depending upon the time when sampling is completed, the samples may be stored in a refrigerator at ECS in Agawam (i.e., if the sampling is completed after the closing of mail facilities). ECS will package the samples into coolers and overnight deliver them to SWRI in San Antonio, Texas and SUNY in Albany, New York and deliver them to SAI in Agawam, Massachusetts. SWRI will receive carpet surface dust, vacuum bag dust, wipe, unit ventilator filter, and air samples. SAI and SUNY will receive carpet surface dust, wipe, and unit ventilator filter samples. SWRI will process the air and vacuum bag dust samples in order to extract any PCBs into a solution, which will be split into four aliquots. SWRI will ship an aliquot to SAI and SUNY, analyze one aliquot, and hold onto an aliquot for QA/QC purposes (e.g., in case an aliquot is lost during shipping).

<u>Rationale</u>: For chain of custody purposes, it is important that ECS and the respective laboratories maintain possession of the samples during and after sample collection. The analytical methods require that the samples be cooled after collection and prior to analysis.

SAMPLE ANALYSIS METHODS

All laboratories must follow detailed standard operation procedures (SOPs) that are agreed to prior to the start of sampling.

<u>Description</u>: Following their SOPs for sample preparation for PCB analysis, SWRI, SAI, and SUNY will first process the samples into a solution in order to extract any PCBs.
<u>Air and Vacuum Cleaner Bag Dust:</u> SWRI will be analyzing air and vacuum cleaner bag dust samples for both particle-phase and vapor-phase PCB Aroclors and congeners using a modified USEPA Method TO-4A. SWRI will be analyzing vacuum cleaner bag dust samples for both PCB Aroclors and congeners using USEPA Method TO-4A. SAI will be analyzing air and vacuum cleaner bag dust samples for PCB Aroclors using USEPA Method TO-4A. SUNY will be analyzing air and vacuum cleaner bag dust samples for PCB congeners using a method based on two published research papers: DeCaprio et al. 2000, 2005.

<u>Carpet Surface Dust, Wipes and Unit Ventilator Filters:</u> SWRI will be analyzing carpet surface dust, wipe, and unit ventilator filter samples for PCB Aroclors and congeners using a modification of USEPA Method TO-4A. SAI will be analyzing carpet surface dust, wipe, and unit ventilator filter samples for PCB Aroclors using USEPA Method SW846: 8082. SUNY will be analyzing carpet surface dust, wipe, and unit ventilator filter samples for PCB Aroclors using USEPA Method SW846: 8082. SUNY will be analyzing carpet surface dust, wipe, and unit ventilator filter samples for PCB congeners using a method based on two published research papers: DiCaprio et al. 2000, 2005.

<u>Rationale:</u> Three different laboratories (i.e., Spectrum Analytical Laboratory, SUNY Institute for Health and Environment Laboratory, and Southwest Research Institute) will be analyzing samples in order to address questions raised about previous testing that analyzed separate samples from the school for Aroclors and congeners and reported different results. USEPA Method SW846: 8082, 1668A, and TO-4A are certified by the USEPA. USEPA Method SW846: 8082 uses gas chromatography/mass spectrometry to detect PCBs, USEPA Method TO-4A uses gas chromatography/multi-detector detection, and the modified USEPA Method TO-4A uses gas chromatography/mass spectrometry. SUNY bases their method from two published research papers, which describe the method for congener analysis in detail. This method uses parallel dual-column gas chromatography with electron capture detection.

ANALYTES

<u>Aroclors</u>: Aroclor is the industrial trade name for commercially produced mixtures of PCBs used in the manufacturing of electrical equipment at GE. The mixtures consist of varying amounts of chlorine, which are signified by the last two digits of their names. For example, Aroclor 1254 contains approximately 54% chlorine by weight, while Aroclor 1260 contains approximately 60% chlorine by weight. The exception is Aroclor 1016, which contains approximately 41% chlorine by weight (ATSDR 2000).

The samples will be analyzed for seven specific Aroclor mixtures: 1016, 1221, 1232, 1242, 1248, 1254, and 1260. These are the Aroclors that the USEPA Method SW846: 8082 has been tested for (USEPA 1996).

<u>Congeners</u>: Congeners are single, unique compounds within PCBs (ASTDR 2000). While there are a total of 209 different congeners, most are not commonly detected (McFarland and Clarke 1989). Based on a review of published literature on congeners detected in house dust and indoor air (e.g., Currado and Harrad 1998; Kohler et al. 2002; MacLeod 1981; Vorhees et al. 1997, 1999; Wallace et al. 1996; Wilson et al. 2001) their percent makeup in the above listed Aroclors (Camann et al. 2002; Camann et al. 2001; Levin et al. 2002; Rudel et al. 2003; Wolff et al. 1997), and the congeners that were previously analyzed for by SUNY, the samples will be analyzed for 101 specific congeners: #1, 3, 4+2, 10, 7, 9, 6, 8, 19, 13, 18, 15, 17, 24+27, 32+16, 29, 26, 25, 31, 28, 33, 53, 51, 22, 45, 46, 52, 49, 47+59, 44, 42, 71, 64, 40, 67, 63, 74, 70, 66, 95, 91, 56, 92, 84, 90+101, 99, 83, 97, 87, 85, 136, 110, 77, 151, 144, 147+109, 123+149, 118, 134, 114, 146, 153, 132, 105, 141, 179, 137, 176, 130, 164+163+138, 158, 129, 187, 183, 128, 185, 174, 177, 171, 156, 201, 172, 180, 200, 170, 190, 199, 203, 196, 195, 194, 206. This list of PCB congeners includes the 18 PCB congeners which comprise at least 5% by weight of several Aroclor mixtures; many of them are prevalent in several of the Aroclor mixtures (ATSDR 2000, Camann 2006). This congener list also represents the full range of lower to higher chlorinated congeners.

QA/QC PROCEDURES

<u>Laboratory Control Sample</u>: The accuracy of the laboratory analysis will be checked by having the laboratories analyze spiked sample media. An unused media for each sample type (i.e., cotton wipe, unit ventilator filter, and air cartridge filter) will be shipped to each laboratory. The laboratories will spike the media with a known PCB Aroclor or congener and then analyze the sample for it. This will provide percent recovery.

<u>Matrix Duplicate</u>: An intra-laboratory split sample which is used to document the precision of a method in a given sample matrix.

<u>Method Blanks</u>: Sample contamination resulting from the laboratory analytical methods will be checked by method blanks. Method blanks consist of an analyte-free matrix to which all reagents are added in the same volumes or proportions as used in sample processing. The method blank will be carried through the complete sample preparation and analytical procedure.

<u>Standard Reference Materials</u>: The comparability of the congener laboratory results will be checked by having SWRI and SUNY analyze standard reference materials (SRMs). SRMs are produced by the National Institute of Standards and Technology (NIST) and are certified to contain a specific amount of a substance. SRMs for Aroclor analysis are no longer produced. The congener SRMs will help in comparing data from SWRI and SUNY.

<u>Surrogates</u>: A surrogate is an organic compound that is similar to the target analyte (i.e., PCBs) in chemical composition and behavior in the analytical process, but which is not normally found in environmental samples.

<u>Trip Blanks</u>: Field sampling methods will be checked by collecting a trip blank using precleaned sample containers provided by ECS. Trip blanks are used to assess field introduced PCB contamination into samples. Air and wipe trip blanks will comprise the sampling media placed in the sample container. Unit ventilator filter trip blanks will be comprised of clippings from new unit ventilator filters placed in the sample container. Trip blanks will be packaged at the school, like the regular samples, to capture any fieldintroduced PCBs. Trip blanks for each medium will be collected and analyzed by each laboratory.

DATA EVALUATION

The data will be evaluated by the BEH/ETP using methodologies consistent with readily available guidance or methods, and consistent with evaluations contained in the public health assessments produced by MDPH (MDPH 2003a-h).

Air Samples

Description: The air samples will be evaluated by using health-based screening values, such as the CREG, that have been scientifically peer reviewed or derived using scientifically peer reviewed values and published by ATSDR. If a concentration of PCB exceeds its screening value, adverse health effects are not necessarily expected. Rather, the concentration can be further evaluated for the specific situation (e.g., outdoor sample, classroom sample) to determine whether health effects might be possible. In addition to screening, the results from the air samples will also be used to calculate a lifetime average daily intake, which takes into account certain assumptions, such as the age specific average weight of the person, air intake rate [e.g., 10 cubic meters per day (m^{3}/day) for child, 15.2 m³/day for adult], and the length of time in the building (e.g., 6 hours/day for 180 days/year for child, 8 hours/day for 180 days/year for adult). ATSDR has not developed an MRL for inhalation because of a lack of sufficient data on which to base an MRL. In lieu of this, the air results will be compared to the lowest level that adverse health effects (LOAEL) have been observed in animal studies (LOAEL = 3 $\mu g/m^3$) for evaluating the risk of adverse noncancer health effects. The air sample results will also be compared to background values reported in previously published studies that evaluated PCB concentrations in air (e.g., ATSDR 2000, Vorhees et al. 1997).

<u>Rationale:</u> MDPH/BEH/ETP traditionally uses both quantitative and qualitative approaches to evaluating results. Examples of this can be seen in the Discussion sections of the public health assessments MDPH/BEH/ETP has written for the GE sites. These can be found on the MDPH website (<u>www.mass.gov/dph/ceh</u>), at the Berkshire Atheneum, or by calling MDPH to request a copy (617-624-5757).

Carpet Surface Dust, Vacuum Cleaner Bag Dust, and Wipe Samples

<u>Description</u>: Carpet surface dust, vacuum cleaner bag dust, and wipe samples measure the possible concentration of PCBs in the dust and residue on a specific surface. Individuals (e.g., students, staff) that come into contact with PCBs that are in the dust and residue could potentially ingest them or the PCBs could be absorbed through their skin. The results from these samples will be used to calculate a lifetime average daily intake, which takes into account certain assumptions, such as the average weight of children and adults (e.g., 35 kilograms for child, 70 kilograms for adult), the amount of total soil adhered [e.g., 525 milligram per day (mg/day) for child, 326 mg/day for adult], and the length of time in the building (e.g., 6 hours/day for 180 days/year for child, 8 hours/day for 180 days/year for adult). The lifetime average daily intake can be compared to standard comparison or screening values such as the ATSDR Minimum Risk Level (MRL), which is 0.00002 milligrams per kilogram per day [milligram per kilogram per day (mg/kg/day)] for chronic oral exposure. The MRL is an estimate of daily human exposure to a substance (e.g., PCBs) that is likely to be without an appreciable risk of adverse noncancer health effects over a specified duration of exposure. MRLs are derived from no-observed-adverse-effect-levels (NOAELs) or lowest-observed-adverseeffect-levels (LOAELs) from either human or animal studies. For cancer effects, estimated intake can be compared to Cancer Risk Evaluation Guides (CREGs). CREGs are derived assuming a lifetime of exposure in a residential setting. While there is not a CREG for the ingestion of PCBs in dust, there is a CREG of 0.4 mg/kg for the ingestion of PCBs in soil. These comparison values are intended to be used as guidance. It is also important to emphasize that exposures to children should be prevented or minimized to the extent possible.

With regard to skin contact with PCB dust from surfaces, the Exposure Factors Handbook has summarized literature for children on this topic (USEPA 1997). In general, the major factors that affect opportunities for exposure via skin contact (e.g., PCBs from surface wipe samples from indoor environments) are: how much PCB is in contact with the skin; the potential amount taken in by ingestion or skin absorption the amount of skin surface area exposed; and the duration of exposure. It is important to note that not all of the compounds (e.g., PCBs) found in a layer of dust/dirt on the skin surface may be taken into the body by ingestion or skin absorption. However, in many cases assumptions can be made to estimate what the upper limit of ingestion/absorption may be so as to know whether there is a reason to be concerned about health impacts. We know that a number of factors influence how much dust/dirt adheres to skin. Increased dust/dirt moisture levels, hand contact, and outdoor activities, particularly with wet soil contact (e.g., wetlands, riverbanks) will lead to greater dust/dirt adherence to skin. The wipe sample results will also be compared to values reported in previously published studies that evaluated PCB concentrations in dust (e.g., ATSDR 2000, Vorhees et al. 1999).

<u>Rationale:</u> There is little information available on federal or state guidelines or standards for evaluating PCB carpet surface dust, vacuum cleaner bag dust, or wipe sample results for human health purposes. The only formal guidelines that were found were a USEPA clean-up standard of 10 micrograms PCB per 100 square centimeters (10 μ g/100 cm²) for wipes collected from indoor residential surfaces that have been affected by a spill of a low-concentration PCB mixture (40 Code of Federal Regulations 761.125) and a recommended clean-up standard of 0.1 μ g/100 cm² developed by the California Department of Toxic Substance Control for PCB contamination in schools resulting from lighting retrofits (CDTSC 2003). Exposure will be estimated and compared to the MRL, NOAELs, and LOAELs. We will approach the interpretation of these samples utilizing a standard approach as described in the equation in the Exposure Factors Handbook (see Attachment) and guidance for ATSDR public health assessments.

Unit Ventilator Filter Samples

<u>Description</u>: The unit ventilator filter samples will be qualitatively evaluated by reviewing information on all other sample results and such factors as weather, location, etc. and by qualitatively comparing these results to indoor classroom results.

<u>Rationale:</u> There are no available federal or state guidelines or standards for evaluating PCB unit ventilator filter sample results for human health purposes. The unit ventilators are designed to transport outside air into the classroom, filter it, and to re-circulate the air once it is inside. As such, they can capture particulates with PCBs and hence provide a qualitative indicator of the presence of PCBs in fugitive dust. However, there is no direct exposure to the filters themselves (not accessible except occasionally to maintenance staff). For that reason, measurements in carpet surface dust, wipe, vacuum cleaner bag dust, and air samples are more important in evaluating exposure risks because individuals can come into contact with PCBs in those media. Results from unit ventilator filter samples may provide an understanding of potential exposure opportunities from particulate matter containing PCBs over extended periods of time.

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FIGURE



ATTACHMENT

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6. DERMAL ROUTE

Dermal exposure can occur during a variety of activities in different environmental media and microenvironments (U.S. EPA, 1992). These include:

- Water (e.g., bathing, washing, swimming);
- Soil (e.g., outdoor recreation, gardening, construction);
- Sediment (e.g., wading, fishing);
- Liquids (e.g., use of commercial products);
- Vapors/fumes (e.g., use of commercial products); and
- Indoors (e.g., carpets, floors, countertops).

The major factors that must be considered when estimating dermal exposure are: the chemical concentration in contact with the skin, the potential dose, the extent of skin surface area exposed, the duration of exposure, the absorption of the chemical through the skin, the internal dose, and the amount of chemical that can be delivered to a target organ (i.e., biologically effective dose) (see Figure 6-1). A detailed discussion of these factors can be found in Guidelines for Exposure Assessment (U.S. EPA, 1992a).

This chapter focuses on measurements of body surface areas and various factors needed to estimate dermal exposure to chemicals in water and soil. Information concerning dermal exposure to pollutants in indoor environments is limited. Useful information concerning estimates of body surface area can be found in "Development of Statistical Distributions or Ranges of Standard Factors Used in Exposure Assessments" (U.S. EPA, 1985). "Dermal Exposure Assessment: Principles and Applications (U.S. EPA, 1992b), provides detailed information concerning dermal exposure using a stepwise guide in the exposure assessment process.

The available studies have been classified as either key or relevant based on their applicability to exposure assessment needs and are summarized in this chapter. Recommended values are based on the results of the key studies. Relevant studies are presented to provide an added perspective on the state of knowledge pertaining to dermat exposure factors. All tables and figures presenting data from these studies are shown at the end of this chapter.

6.1. EQUATION FOR DERMAL DOSE

The average daily dose (ADD) is the dose rate averaged over a pathway-specific period of exposure expressed as a daily dose on a per-unit-body-weight basis. The ADD is used for exposure to chemicals with non-carcinogenic non-chronic effects. For

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compounds with carcinogenic or chronic effects, the lifetime average daily dose (LADD) is used. The LADD is the dose rate averaged over a lifetime.

For dermal contact with chemicals in soil or water, dermally absorbed average daily dose can be estimated by (U.S. EPA, 1992b):

 ADD
 SAcord X EV X ED X EF X SA

 ADD
 BW X AT

 ADD
 average daty dose (mg/kg-day);

 DAcord X
 absorbed dose per event (mg/kg-day);

 DAcord X
 event frequency (eventskay);

 EV
 event frequency (eventskay);

 ET
 exposure duration (vears);

 EF
 exposure duration (vears);

 EA
 exposure duration (vears);

 BW
 averaging free (days) an proceerbogenb effects, AT = ED and for cardinogenc effects, AT = 70 years to 23,550 days.

This method is to be used to calculate the absorbed dose of a chemical. Total body surface area (SA) is assumed to be exposed for a period of time (ED).

For dermal contact with water, the DA_{event} is estimated with consideration for the permeability coefficient from water, the chemical concentration in water, and the event duration. The approach to estimate DA_{event} is different for inorganic and organic compounds. The nonsteady-state approach to estimate the dermally absorbed dose from water is recommended as the preferred approach for organics which exhibit octanol-water partitioning (U.S. EPA, 1992b). First, this approach more accurately reflects normal human exposure conditions since the short contact times associated with bathing and swimming generally mean that steady state will not occur. Second, the approach accounts for uptake that can occur after the actual exposure event due to absorption of residual chemical trapped in skin tissue. Use of the nonsteady-state model for organics has implications for selecting permeability coefficient (K_p) values (U.S. EPA, 1992b). It is recommended that the traditional steady-state approach be applied to inorganics (U.S. EPA, 1992b). It is recommended that the traditional steady-state approach be applied to inorganics (U.S. EPA, 1992b). Detailed information concerning how to estimate absorbed dose per event (DA_{event}) and K_p values can be found in Section 5.3.1 of "Dermal Exposure Assessment: Principles and Applications" (U.S. EPA, 1992b).

For dermal contact with contaminated soil, estimation of the DA_{crent} is different from the estimation for dermal contact with chemicals in water. It is based on the concentration of the chemical in soil, the adherence factor of soil to skin, and the absorption fraction. Information for DA_{crent} estimation from soil contact can be found in U.S. EPA (1992b), Section 6.4.

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APPENDIX B: SUMMARY PROTOCOL FOR TESTING SERUM PCBs

PROJECT SUMMARY AND ENCLOSURES

Introduction/Background

The primary purpose of this document is to provide the protocol/rationale for interpreting serum PCB results of faculty/staff and students who attend the Allendale School in Pittsfield. PCB serum testing of students, parents, faculty and staff of the Allendale School is being offered as a service to the Allendale School community in response to public concerns related to the General Electric (GE) disposal site (i.e. Hill 78 and Building 71) located in an area adjacent to the school.

The disposal site receives PCB waste materials from the clean-up of the GE sites in Pittsfield. The Massachusetts Department of Public Health, Center for Environmental Health. (MDPH/BEH) has completed eight public health assessments for the GE sites (the public health assessment for Hill 78 Landfill Area conducted by the MDPH/BEH is enclosed for your information). The remedial work being carried out by GE contractors is under the oversight of the U.S. Environmental Protection Agency (EPA) under a Consent Decree agreed to by EPA, the Massachusetts Department of Environmental Protection (MDEP), the City of Pittsfield, and GE in 2001. The disposal site consists of two landfills, one lined landfill (i.e. Building 71 area) for higher level waste (e.g. PCB materials over 50 ppm, liquid wastes), and one landfill that is not lined (i.e. Hill 78 Landfill Area) for lower level waste (i.e. PCBs equal to or less than 50 ppm). Hill 78 Landfill Area was preexisting and historically received PCB waste materials at concentrations higher than the current 50ppm limit and other hazardous wastes. Systematic field sampling is done to determine the level of contamination. The remediation activities began around 2000. According to EPA the Building 71 Landfill is expected to reach full capacity this year (2006) and to have its final cap installed by 2007. EPA expects the Hill 78 Landfill to reach capacity in 2008 and have its final cap installed by 2009. Disposal activities occur during times of the year when the ground is not frozen (e.g. March/April through November/December). There are specific work practices and monitoring requirements in place under the Consent Decree. EPA has recently enhanced these monitoring efforts.

Monitoring results (i.e. ambient air monitoring including a new air monitor at Allendale School) conducted along the perimeter of the disposal site have averaged non-detectable or below health risk based criteria established by EPA. Soil sampling on the school playground and in the crawlspace under the school was also conducted by EPA and DEP in the fall of 2005. Results were non-detectable or have averaged below health risk based criteria established by EPA or MDEP.

In November/December 2005, MDPH/BEH hired independent contractors and sampled indoor air (and one outdoor air for background comparison), surface dust wipes, and air vent filter samples (103 total samples) in response to concerns related to the potential for site contaminants to enter the indoor environment. All samples were non-detectable for PCBs.

Concurrent with these indoor environmental tests, a local advocacy group also collected two filter samples from the Allendale School and they were found to have low levels of PCBs based upon congener analyses. MDPH/BEH, in collaboration with a work group, is designing and implementing follow-up indoor environmental monitoring for June 2006, a time of year when the weather is warmer and the disposal site is active.

Serum PCB Testing

With regard to implementation of serum PCB testing, Berkshire Medical Center (BMC) will be providing phlebotomy services. BMC has provided these types of services for a number of MDPH projects in Berkshire County involving serum PCB measurements since 1995. Training on the proper collection, preparation, and shipping of blood samples will be provided to the BMC staff by MDPH State Laboratory Institute (MDPH/SLI) staff. The U. S. Centers for Disease Control and Prevention (CDC) in Atlanta has agreed to perform all analyses using a congener specific method as published in the Third National Report on Human Exposure to Environmental Chemicals in July 2005 (we have enclosed the summary and PCB chapters of this report for your information). The CDC has informed us that they are ready to begin receiving samples for analyses in May 2006.

The Third National Report presents biomonitoring exposure data for 116 environmental chemicals including PCBs for the civilian non-institutionalized U. S. population over the period 2001-2002 and is a nationally representative survey. The serum collection procedures were supplied by the CDC laboratory. Copies of the Blood Collection for Serum PCBs (supplies, procedures, flow chart for the phlebotomists), the CDC Method Summary, and the CDC Laboratory Procedure Manual for PCBs and Persistent Pesticides are enclosed. Analysis at CDC will be performed by high-resolution gas chromatography/isotope dilution high-resolution mass spectrometry (HRGC/ID-HRMS). Thirty-eight PCB congeners will be quantified according to the current CDC procedure that is being implemented for the Fourth National Report. Hence, the samples will be analyzed by state-of-the-art instrumentation and methodologies.

With regard to obtaining consent and important supplementary information to aid in the interpretation of results, we have enclosed copies of two consent forms, one to be signed by a parent for their child; the other to be signed by adult participants. These consent forms have been adapted from similar consent forms we have previously used for participants in PCB blood testing in Berkshire County and elsewhere in Massachusetts and that have been reviewed and approved by our Institutional Review Board (IRB). This version of the updated consent form was also recently approved by the MDPH Office of General Counsel.

With regard to obtaining supplementary (exposure) information, MDPH/BEH traditionally uses a standard questionnaire for obtaining information on risk factors that are known to or may affect serum PCBs levels. The two questionnaires (adult, child) will include questions on the following: age, gender, residential history (including duration of residence), usual occupation, occupation associated with use of PCBs, company, duration, number of years attending or working at Allendale School, locations in the school where most time was spent for up to each of the last seven school years (if applicable), time spent indoors and outdoors during the school day, fish consumption in general, freshwater fish consumption (how obtained, source, Housatonic River fish), change in fishing/fish consumption habits, fiddlehead fern gathering/consumption, recreational areas and types of activities in Pittsfield area (camping, playgrounds, dirt biking, etc), hunting/wildlife consumption (type of prey, how often), gardening (type), playing in dirt or grass at current address, farm residence, open ended question on any other contact with PCBs, breast feeding and duration (for child participant), number of prior children breast fed (for adult female parent), lifestyle risk factors (e.g., smoking). The questionnaires will be administered in two parts; the more lengthy first part with be administered over the phone before the blood draw and the second part will be administered at the time of the blood draw. The second part of the questionnaire includes questions relevant to the blood draw (i.e. weight and height) as well as questions which will require the participant to view a map of the Allendale School.

Interpretation of Serum PCB Analyses

With regard to the interpretation of results, an important observation that has been made by public health researchers including MDPH/BEH is that serum PCB levels generally increase with age. Younger people have very low to (in many cases) non-detectable levels. The enclosures related to the PCB testing from the Third National Report clearly support this trend. Adult participants will be compared to their respective age/race group in the Third National Report. The considerable background information provided in the questionnaires for each participant will aid in understanding both individual and group results.

With regard to children under the age of 12, the older children in this age group would be expected to be similar to the 12 to 19 year olds included in the Third National Report. Younger participants would be expected to demonstrate lower serum PCB results (possibly nondetectable), but the responses to the questionnaire (e.g., dietary exposure, history of breast feeding) will provide important supplemental information. Research on the world literature to identify other groups of children who have been tested for PCBs is enclosed. While interesting, a number of these populations are confounded by industrial exposure, accidental poisoning, known high fish or blubber consumption, and other environmental sources of PCB exposure. The enrollment date into some studies is very long ago (e.g., 1960s, 1970s, 1980s) and CDC has told us that levels have dropped considerably, perhaps up to 80% since the 1980s. Thus, many of these cohorts are not necessarily optimal in establishing background levels today. The Dutch and German studies have more recent recruitment and provide information for younger children. The Faroese children (and mothers) are heavy consumers of blubber and fish so their levels would be expected to be higher. The Anniston, Alabama, study is interesting because it is very recent, there are young children included, and the laboratory methods are identical to those being used for the Allendale School community.

Laboratory methodology is important in measuring concentrations. Detection limits and quantification of varying numbers of congeners differ across studies (e.g., the Dutch studies use 4 congeners). Because children are normally so low, many researchers like to use the known higher more persistent congeners (e.g., PCB-153) as they are more reliably detected and measured in young children. When comparing studies with different congeners with regard to levels in the population, some researchers have picked one common congener (e.g., PCB-153) that is known to be usually the highest, while others choose all of the congeners that the studies being compared have in common to determine which study population has "higher" levels. We prefer the latter approach. Thus, in the approach to interpreting results from the Allendale School children that were common with any comparison study (e.g., the Third National Report, the Anniston study). We will then sum the concentrations of the congeners common to both the Allendale children and the comparison study to determine whether the Allendale children were higher or lower than the comparison study. Finally, we will qualitatively compare congener

patterns from the chromatograms to observe any patterns in the Allendale participants that may be different from the patterns CDC typically observes based on general dietary exposure in the U.S.

As noted earlier, each participant will be evaluated on a case by case basis. MDPH/BEH will work closely with CDC to interpret any findings (particularly as it relates to the child participants). MDPH/BEH in collaboration with CDC will decide whether either individual or group findings need further follow-up investigation based on review of all of the information.

Health and Medical Peer Review Team (MDPH/HMPRT): Summary of responses to comments for documents relating to PCB blood testing for the Allendale School.

Comments from HMPRT Member 1:

1. Comment:

<u>Methodology</u>- Would like clarification, difficulty determining minimal detection limits (or Limits of Detection-LOD) in information provided.

o LOD vary for different congeners

• Units vary from lab to lab

• Based on attached articles: Detection limits must be pretty low- at least in the .01-.04 ppb (whole weight basis) to detect various congeners.

Response:

The published analyses from the Centers for Disease Control and Prevention (CDC) Third National Report on Human Exposure to Environmental Chemicals (NHANES III, July 2005) have congener specific LOD's. There are also individual LOD's for each sample, largely due to the sample volume available for analysis being different for each sample. A higher sample volume results in a lower LOD and a better ability to detect low levels, as stated in NHANES III, appendix A. The CDC is conducting the analysis for the serum samples from the Allendale School Community using the most up to date congener specific methods that they use for the ongoing NHANES sampling; therefore there will be no lab to lab unit discrepancies when comparing Allendale School test results and results from this report for interpretation.

According to the CDC, the method detection limits ranging from 0.01-0.04 ppb (whole weight, g/g) are typical for most methods using about 1 mL of sample. The Allendale School serum collection will result in analyses of 2 mL samples. In general, the CDC's PCB congener specific detection limits for these samples should be approximately half or .005-0.020 ppb (whole weight).

2. Comment:

<u>Proposed Interpretation</u>- Need to use correct language for conveying interpretation of results to parents, to give results some meaning.

Response:

MDPH will be working closely with the CDC to interpret the findings. Comparisons will be made to the information from the Third National Report on Human Exposure to Environmental Chemicals (NHANES, July 2005) and other literature sources to properly convey this information to the participant in a letter clearly informing them of where their levels fall (if detected) in comparison with others of the same age and gender.

3. Comment:

Additional Articles for interpreting results:

• Relationship of Lead, Mercury, Mirax, Dichlorodiphenyldichloroethylene, Hexachlorobenzene, and Polychlorinated Biphenyls to timing of Menarche Among Akwesasne Mohawk Girls. Denham, M et al., Pediatrics 2005;115;e127-e134.

• Organochlorines, Lead, and Mercury in Akwesasne Mohawk Youth. Schell, L. et al. Environmental Health Perspectives Vol. 111 Num 7 June 2003, 954 – 961.

Response:

The literature on PCB's is voluminous; we provided only a sample of some of the recent publications. Thank you for providing these references with regard to blood concentrations of youth ages 10-17 years old, we will add them to our background information.

Comments from HMPRT Member 2:

4. Comment:

<u>Project Summary</u>, Paragraph 2: I think it is important to remind stakeholders that the newer Building 71 OPCA is lined and that the Hill 78 OPCA, which was pre-existing, is not lined. Although, Hill 78 now only receives PCB waste <50ppm, it did historically receive PCB waste at significantly higher concentrations. It also contains other hazardous wastes that have not been as well quantified or monitored.

Response:

This information has been added to the project summary Introduction/Background paragraph 2 and has been communicated at numerous public meetings. While this information regarding Hill 78 being unlined is accurate, it is important to note that the gradient is toward the River not toward the School.

5. Comment:

<u>Consent Forms</u>: I think it's important to share with both parents of children and adults the following. Maybe the best format to do this in would be a **Frequently Asked Questions** format.

a) That you are doing congener specific testing and that you will be looking for congener patterns that may be different from patterns CDC typically observes based on general dietary exposure in the United States.

The Allendale community has been very sensitized to this issue and will be reassured that you are aware that patterns secondary to non-dietary exposures may differ and that they may also be different from the original aroclor mixtures used at the site.

b) That each participant will be evaluated on a case-to-case basis, and that MDPH/BEH will work closely with the CDC to interpret any findings, particularly as they relate to child participants.

c) That MDPH/BEH in collaboration with the CDC will review the test results and decide whether individual or group findings need further follow-up investigation.

d) That a copy of the blood tests results, summary of the questionnaire, and the interpretation will be provided to the participant (or parent of the participant if the participant is a minor) and ONLY them.

If they want to share this information with their individual health care providers, it will be completely up to them. Other parties including but not limited to insurers, employers, school administration, city officials, will NOT receive any information that could be linked to individual participants.

The blood tests results will not be included in individual patients' medical charts unless the participant specifically chooses to share results with their health care provider and specifically requests that a copy be included in their records.

It will also be important for participants and health care providers to have MDPH contact information should they have additional questions or concerns.

Response:

(a) The letter from the MDPH, which went home with students and staff of the Allendale School, informed the Allendale School Community that we are conducting congener specific analysis. We will continue to stress this in all our communication initiatives. (b and c) Language contained in the letter to the Allendale School Community and the consent form express the use of the questionnaire to collect information and conveys the collaboration of the MDPH and the CDC regarding the interpretation of individual results as they relate to children. (d and e) The consent form states that the information provided and the blood test results will be treated as confidential information and will not be published or shared with anyone else in a manner that could readily be associated with the individual. (f) We have already had numerous contact with individuals requesting testing and expressing questions/concerns relating to PCBs and the Allendale School Community. Our contact information will also be included with any correspondence including notification of test results.

A frequently asked questions document has also been drafted to communicate these comments to the Allendale School Community.

6. Comment:

The participant questionnaire was not included in my packet, and I think it is important for us to review. The Anniston study mentioned that the accuracy of the correlation coefficient between blood PCB concentrations and length of residency will depend on how questions about residency are asked. Many families will move, but their moves are still within a ½ to 1-mile radius of the OPCAs and other PCB contaminated sites.

Response:

It is standard policy to not release a questionnaire before it is administered to the participants. The questions relating to residency on the questionnaire address the possibility of having lived at several previous addresses within a $\frac{1}{2}$ to 1-mile radius of a contaminated site as well as address the possibility of living near other PCB contaminated sites. The questions ask for:

- Current address and length of residency.
- Previous address and length of residency. This question includes space for four previous addresses, when they lived there, and total number of years.
- The questionnaire also includes space for information regarding a child's time spent at additional (current) addresses, a child who may spend a significant amount of time at the residence of a family membér or other parent/guardian. The question asks for:
- Additional current addresses (e.g. split residency) or an address where the child spends a significant amount of time.
- % time at each address.
- Length of time he/she has lived or visited there.

7. Comment:

Will you be doing GIS mapping of the PCB blood results that includes prior residencies within 1 mile of OPCA and other contaminated sites?

Response:

Individual test results will be analyzed on a case by case basis. The purpose of the questionnaire is to supply supplementary (exposure) information for the individual and provide necessary information to interpret any findings. MDPH/BEH in collaboration with CDC will decide whether individual or group findings need further follow-up investigation based on review of all of the information. This includes looking closely at individual addresses if warranted by the findings of the PCB serum testing, GIS mapping is available if needed.

8. **Comment:**

Also, if there are "detects" in the PCB blood testing, you may also want to do GIS mapping of participants' maternal residencies. Maternal proximity to the sites may be linked to participants' potential exposures while in utero or breastfeeding.

Response:

In situations where the mother is also being tested, we are gathering additional information to answer this type of question if detects are found. If warranted further investigation can be conducted to obtain any information that was not provided by the participant on the questionnaire.

- We are asking for residence history on both the parent/staff questionnaire and the student questionnaire.
- There are questions regarding breast feeding on both questionnaires:
 - Parent/Staff- If they have ever breastfed, how many children and how long each child was breastfed.
 - Student- If they were breastfed. If they have older siblings who have also been breast fed, birth order, and how long each sibling was breastfed.
- We have added a question that specifically asks mother's address at time of child's birth to clarify the address. This will provide the exact address if information is left out of either the mother's or child's residence history.
- We are also asking for parent/student information, which will match family members to one another in order adequately interpret answers to these questions that directly affect both participants.

9. Comment:

<u>Background Research</u>: Thank you for sharing these papers with us. Rich Rosenfeld has mentioned some additional papers that may be helpful in the interpretation of PCB blood testing as it relates to children. In addition, I would like to see the *ATSDR's Health Consultation titled Evaluation of soil, blood, and air data from Anniston, Alabama, Calhoun County, Alabama* included because it specifically discusses potential links with airborne PCBs. The executive summary concludes that exposure to PCBs in the air presents an indeterminate public health hazard, and recommends additional investigation to a. identify persons living near air monitors at which elevated air PCB levels have been detected and b. define the limits of the area with elevated air levels for PCBs. The health consultation can be found by going to: <u>www.atsdr.cdc.gov/HAC/PHA/annpc/ann_p1.html</u>. Also, do you know if there have been any follow-up studies in Anniston?

Response:

The literature on PCB's is voluminous; we provided only a sample of some of the recent publications. Thank you for providing these references, we will add them to our background information.

Comments from HMPRT Member 3:

10. Comment:

The protocol has been carefully thought through and addresses the key methodologies and interpretative issues that often arise in studies of this type. The strengths of the protocol include the following.

- The <u>timing of the indoor environmental testing</u> during the warmer months when the site will be more active makes good sense.
- The <u>serum samples</u> will be tested for 38 PCB congeners at the CDC using the latest equipment, methods, and quality control procedures.
- A great deal of thought has been given to <u>selecting comparison survey data</u> that will provide a reasonable set of background levels to which the Allendale School results can be compared.
- Background levels will be ascertained separately for children and adults.
- The <u>epidemiologic questionnaire</u> is very comprehensive and will permit the investigators to assess behaviors that are likely to create opportunities for exposures to environmental PCB contamination.
- The Berkshire Medical Center (BMC), which has participated in previous PCB studies in the area, will continue to provide <u>standardized phlebotomy services</u> for the proposed study.
- The <u>informed consent form</u> has been reviewed and approved by the MSDPH Internal Review Board and the Department's Office of General Council.

In summary, the protocol provides scientifically sound methods for collecting and testing serum samples from children and adults for interpreting study results in relation to normal background levels of PCBs.

Response:

Noted

Comments from HMPRT Member 4:

11. Comment:

Are the General Electric PCB congeners within the mix of those in the CDC testing? I assume they all are, so that a fingerprint of subtyping can be accomplished.

Response:

The congeners that we are testing for and the congeners that the CDC has included in NHANES III and IV were chosen on the basis that proven methods for testing are established, that there is exposure data for the population for comparison, and that they are congeners most commonly found in human serum when testing is done. These congeners tend to be the more environmental persistent congeners; the more volatile congeners would be less likely to be found in serum because of their short half life. Aroclors 1260 and 1254 are the PCB mixtures that were thought to be most readily used at the GE site. The 38 congeners that CDC is testing for are included in the composition of Aroclor mixtures 1260 and 1254 listed by the EPA.

12. Comment:

In the consent for both the adults and the children, there's a relative paucity of language about:

"An acknowledgement of their understanding that the meaning of elevated PCBs in children's blood is not clear and cannot necessarily be interpreted clinically and that they consent to testing knowing that they cannot be effectively counseled about how to interpret the results. Elevated PCBs in a child's blood cannot necessarily be related to future risks of disease development in that individual child."

"An acknowledgement that a parent consents for testing of their children's blood even though they realize that there is no effective treatment available for elevated blood PCB levels in children."

Response:

The Allendale School Community has been notified of the limitations of testing and interpretation through public meetings and letters sent home with all students and staff. These outreach efforts emphasized that testing was being offered as a public service to address concerns of parents, students, and Allendale School staff, not as a result of an MDPH recommendation. The published analyses from the Centers for Disease Control and Prevention (CDC) Third National Report on Human Exposure to Environmental Chemicals (NHANES III, July 2005) will be used to interpret serum testing results for the students being tested. Counseling on recommendations for future behavior to reduce the potential for exposure to PCBs will be provided for those with elevated PCB levels. We have inserted language into paragraph 1 of the consent form to address the fact that there is no medical treatment to reduce current PCB levels and that counseling on behaviors to reduce the risk of future exposure will be provided. Information is contained in the Frequently Asked Questions (FAQs) of the ATSDR toxicology profile for PCBs and we have also drafted an FAQ sheet specific to the Allendale School PCB testing.

13. Comment:

I did not see any agreement in the consent or the material about the testing that contracts for future monitoring of the children should they have elevated blood PCB levels. (or it's certainly possible I overlooked it) And yet a parent (and pediatrician) would reasonably want to know how often the child's blood PCB level should be drawn periodically, for what duration of time, and what other ancillary monitoring by laboratory assessments (thyroid tests? blood counts? hormonal levels?) should be monitored during subsequent well child care. Would the DPH be expecting to serially test the cohort of children who are found to have elevated PCB levels? If so, at what frequency and over what span of time? If not, the parents and physicians need to know in advance the limits of the contracting, I think.

Response:

If we see any unusual findings follow-up will occur as appropriate as well as counseling to avoid any future exposure. We do not expect increased levels in children this young, however if increased levels of PCBs are found they will be examined on a case by case basis. PCBs have an approximate 1-10 year half life depending on the congener so simply offering follow up testing would not be prudent in terms of public health. We are offering serum testing only and do not expect that ancillary testing would be needed for exposure information; clinical effects are not expected at these levels. We will encourage participants to share the results with their physicians if they wish to do so. The limits of testing Allendale School Community has been explained during public meetings and letters distributed to all students and staff.

APPENDIX C: CONSENT FORMS FOR SERUM PCB TESTING

MASSACHUSETTS DEPARTMENT OF PUBLIC HEALTH ALLENDALE SCHOOL PCB SERUM TESTING

PARENT/GUARDIAN CONSENT FORM

I understand that the Massachusetts Department of Public Health (MDPH) is offering PCB serum testing to the administration, faculty, and students of Allendale School as a public service. I understand that if my child is found to have elevated serum PCBs that there is no medical treatment to reduce his/her current PCB levels and I will be counseled on behaviors to reduce his/her risk of future exposure. I have requested to have my child participate in this effort.

A blood sample will be taken from my child to determine the level of PCBs in his/her blood. The blood will be taken from a vein in my child's arm and will require the use of a hypodermic needle and vacutainer. Approximately 20 ml of blood will be drawn. This procedure usually involves little pain or discomfort, but occasionally some discomfort may occur after the blood sample is obtained. Other risks, while unlikely, will be explained by the staff from Berkshire Medical Center who will be taking the blood sample. My child's blood sample will only be tested for PCBs. The blood sample will be destroyed after the analysis and quality control measures are completed.

I agree to participate in a short interview (approximately 15 minutes) that will be conducted by MDPH staff in order to collect important information that may be associated with individual PCB exposure and that may help with the interpretation of results.

I understand that staff from MDPH and Berkshire Medical Center who conduct this effort will use the information that I provide and the results of my child's tests only for the purpose of evaluating my PCB exposure. The information I provide and the blood test results will be treated as confidential information and will not be published or shared with anyone else in a manner that could readily be associated with me and my child.

I understand that I will be notified of the result of my child's PCB blood test after all laboratory testing and quality control measures have been completed. This is to ensure the scientific integrity of the final result of my child's blood test.

I agree to being re-contacted for follow-up questions at a later date. I also understand that I am not under any obligation to have my child participate in this blood testing and that I can end my child's participation at any time. I have read and understand the above statement, and I hereby agree to have my child participate in this blood test and interview.

Name of Child

Name of Parent/Guardian:	Date:

Signature of Parent/Guardian _

MASSACHUSETTS DEPARTMENT OF PUBLIC HEALTH ALLENDALE SCHOOL PCB SERUM TESTING

ADULT CONSENT FORM

I understand that the Massachusetts Department of Public Health (MDPH) is offering PCB serum testing to the administration, faculty, and students of Allendale School as a public service. I understand that if I have elevated serum PCBs that there is no medical treatment to reduce my current PCB levels and that I will be counseled on behaviors to reduce my risk of future exposure. I have requested to participate in this effort.

A blood sample will be taken from me to determine the level of PCBs in the blood. The blood will be taken from a vein in my arm and will require the use of a hypodermic needle and vacutainer. Approximately 20 ml of blood will be drawn. This procedure usually involves little pain or discomfort, but occasionally some discomfort may occur after the blood sample is obtained. Other risks, while unlikely, will be explained by the staff from Berkshire Medical Center who will be taking the blood sample. My blood sample will only be tested for PCBs. The blood sample will be destroyed after the analysis and quality control measures are completed.

I agree to participate in a short interview (approximately 15 minutes) that will be conducted by MDPH staff in order to collect important information that may be associated with individual PCB exposure and that may help with the interpretation of results.

I understand that staff from MDPH and Berkshire Medical Center who conduct this effort will use the information that I provide and the results of my tests only for the purpose of evaluating my PCB exposure. The information I provide and the blood test results will be treated as confidential information and will not be published or shared with anyone else in a manner that could readily be associated with me.

I understand that I will be notified of the result of my PCB blood test after all laboratory testing and quality control measures have been completed. This is to ensure the scientific integrity of the final result of my blood test.

I agree to being re-contacted for follow-up questions at a later date. I also understand that I am not under any obligation to participate in this blood testing and that I can end my participation at any time. I have read and understand the above statement, and I hereby agree to participate in this blood test and interview.

Name:

Date:

Signature:

APPENDIX D: NIST STANDARD AND ANALYSIS RESULTS

APPENDIX D: NIST STANDARD AND ANALYSIS RESULTS

The results included in this report are based on analyses performed by three different laboratories using different analytical techniques to measure PCBs (Aroclor or congenerspecific). As part of the QA/QC protocol developed prior to the start of sampling, SWRI and SUNY agreed to analyze a sample from the National Institute of Standards and Technology (NIST) that contained known quantities of certain congeners. The purpose of this step was to determine the comparability of the SWRI and SUNY analyses and how closely their results matched with the known quantities of congeners in the NIST sample. NIST does not produce samples for Aroclor analysis.

The Table shows the results of the NIST sample analyses. The NIST standard reference material had 30 congeners at certified levels ranging from 0.00414 to 0.0402 mg/kg. SWRI detected 24 of the 30 congeners with concentrations with an average recovery rate of 95%. The recovery rate is the concentration of the congeners detected by the laboratory divided by the known concentration multiplied by 100. SUNY detected all of the 29 congeners that were included in its analysis, with an average recovery rate of 43%. All of the SUNY concentrations for the congeners reported in the NIST standard were less than the certified concentrations.

Congener #	NIST standard (mg/kg)	SWRI (mg/kg)	SUNY (mg/kg)
18	0.0128 ± 0.001	0.018	0.0029
28	0.0134 ± 0.0005	0.011	0.0057
31	0.014 ± 0.0005	0.015	0.0041
44	0.0181 ± 0.0019	ND (0.0094)	0.0066
52	0.0218 ± 0.0019	0.015 [+69]*	0.0091
56	0.00442 ± 0.00028	ND (0.0094)	0.0010
70	0.0131 ± 0.0012	0.018	0.0075
74	0.00522 ± 0.00051	ND (0.0094)	0.0018
87	0.0166 ± 0.0008	ND (0.0094) [+115]*	0.0068
92	0.00548 ± 0.00072	0.0047	0.0013
95	0.0227 ± 0.0026	0.017 [+93]*	0.0099
99	0.0116 ± 0.0004	0.0077	0.0053
101	0.0298 ± 0.0023	0.025	0.0145 [+90]*
105	0.0132 ± 0.0014	0.013	0.0050

Congener #	NIST standard (mg/kg)	SWRI (mg/kg)	SUNY (mg/kg)
107	0.00414 ± 0.00047	ND (0.0094)	Not reported**
110	0.0281 ± 0.0037	0.024	0.0129
118	0.0263 ± 0.0017	0.026	0.0135
138+163	0.0348 ± 0.0033	0.0321 [+164]	0.0163 [+164]
146	0.00489 ± 0.00038	0.0042	0.00268
149	0.0244 ± 0.0019	0.02	0.0102 [+123]
151	0.00692 ± 0.00064	0.0062	0.0031
153+132	0.0402 ± 0.0018	0.023	0.0122
158	0.00450 ± 0.00043	ND (0.0094)	0.0017
170	0.0088 ± 0.0010	0.012	0.003
174	0.00883 ± 0.00047	0.0073	0.0063
180	0.0184 ± 0.0032	0.014	0.0072
183	0.00527 ± 0.00039	0.008	0.0015
187	0.0113 ± 0.0014	0.012	0.0069
206	0.00381 ± 0.00013	0.0031	0.00285

* Combined analytical result with congener number in brackets

** Congener 107 was not reported in results from SUNY lab

*** Congeners 138 and 163 were reported as combined in SUNY lab report. NIST standard values for both congeners were added in line for combined congeners.

APPENDIX E: EXPOSURE CALCULATIONS

APPENDIX E: EXPOSURE CALCULATIONS

In order to further assess contaminants, such as PCBs, and possible related health concerns, calculations are made to estimate the amount of a contaminant people may come into contact with each day (i.e., exposure dose). These calculations account for several factors that are specific to the location and the medium being analyzed. The maximum concentration is the highest amount of the contaminant found during sampling for each medium. This is a conservative assumption since it is unlikely that an individual would be continuously exposed to the highest concentration. Exposure frequency is the rate of exposure within a given time period. For Allendale Elementary School, it is estimated that students and teachers are inside the school for 180 days/year and could be exposed each day. Exposure duration is the length of time of a continuous exposure. For students and teachers, this is estimated to be 6 and 30 years, respectively. The averaging time is the number of days in which an exposure is averaged. For cancer concerns, the default value is the number of days in a 70-year lifespan. Once the exposure dose is calculated, it is multiplied by the cancer slope factor to produce a theoretical cancer risk. The cancer slope factor for PCBs is 2 mg/kg/day⁻¹ (USEPA 1997a). Dermal exposures have several factors that are specific to them alone. The dermal absorption fraction is the percent of the contaminant that is absorbed through the skin. For PCBs, it is 0.14 (USEPA 2004). Event frequency is the estimated number of times that an individual will have contact with the maximum concentration of the contaminant. It is estimated that students and teachers could come into contact with two areas containing PCBs each day. The skin surface area is the amount of skin that is exposed and may come into contact with the contaminant. For students and adults, it is estimated to be 1433 cm² and 2479 cm², respectively, corresponding to the amount of skin on hands, forearms, and face (USEPA 2004). Air exposures include inhalation rates, which are the volume of air that children and adults breathe each day. For children, it is 10 cubic meters per day (m³/day). For adults, it is 15.3 m³/day (USEPA 1997b). Carpet and vacuum dust exposures include soil ingestion rates, which for children is 200 milligrams per day (mg/day) and for adults is 100 mg/day (ATSDR 2005). Soil ingestion rates were used because dust ingestion rates are not available.

<u>1. SURFACE WIPES</u>

<u>Child</u>

Maximum Concentration:	0.00000294 mg/cm ² -event
Dermal Absorption fraction for PCBs:	0.14
Exposure Frequency:	180 days/year
Exposure Duration:	6 years
Event Frequency:	2 events/day (1 inside and outside classroom)
Skin Surface Area:	1433 cm^2
Body Weight:	35 kg
Averaging Time:	25,550 days
Cancer Slope Factor:	2 mg/kg/day^{-1}

 $ExposureDose = \frac{Concentration * DermalAbsorptionFraction * ExposureFrequency * ExposureDuration * EventFrequency * SkinSurfaceArea}{Concentration * DermalAbsorptionFraction * ExposureFrequency * ExposureDuration * EventFrequency * SkinSurfaceArea}{Concentration * DermalAbsorptionFraction * ExposureFrequency * ExposureDuration * EventFrequency * SkinSurfaceArea}{Concentration * DermalAbsorptionFraction * ExposureFrequency * ExposureDuration * EventFrequency * SkinSurfaceArea}{Concentration * DermalAbsorptionFraction * ExposureFrequency * ExposureDuration * EventFrequency * SkinSurfaceArea}{Concentration * DermalAbsorptionFraction * ExposureFrequency * ExposureDuration * EventFrequency * SkinSurfaceArea}{Concentration * DermalAbsorptionFraction * ExposureFrequency * ExposureDuration * EventFrequency * SkinSurfaceArea}{Concentration * DermalAbsorptionFraction * ExposureFrequency * ExposureDuration * EventFrequency * SkinSurfaceArea}{Concentration * DermalAbsorptionFraction * EventFrequency * ExposureDuration * EventFrequency * SkinSurfaceArea}{Concentration * DermalAbsorptionFraction * EventFrequency * ExposureDuration * EventFrequency * SkinSurfaceArea}{Concentration * EventFr$

BodyWeight * AveragingTime

Exposure Dose = $1.4 \times 10^{-6} \text{ mg/kg/day}$

Theoretical Cancer Risk = Exposure Dose * Cancer Slope Factor

Theoretical Cancer Risk = 3×10^{-6}

<u>Adult</u>

Maximum Concentration:
Dermal Absorption fraction for PCBs:
Exposure Frequency:
Exposure Duration:
Event Frequency:
Skin Surface Area:
Body Weight:
Averaging Time:
Cancer Slope Factor:

0.00000294 mg/cm²-event 0.14 180 days/year 30 years 2 events/day (1 inside and outside classroom) 2479 cm² 70 kg 25,550 days 2 mg/kg/day⁻¹ $ExposureDose = \frac{Concentration * DermalAbsorptionFraction * ExposureFrequency * ExposureDuration * EventFrequency * SkinSurfaceArea}{Concentration * DermalAbsorptionFraction * ExposureFrequency * ExposureDuration * EventFrequency * SkinSurfaceArea}{Concentration * DermalAbsorptionFraction * ExposureFrequency * ExposureDuration * EventFrequency * SkinSurfaceArea}{Concentration * DermalAbsorptionFraction * EventFrequency * ExposureDuration * EventFrequency * SkinSurfaceArea}{Concentration * DermalAbsorptionFraction * EventFrequency * ExposureDuration * EventFrequency * SkinSurfaceArea}{Concentration * DermalAbsorptionFraction * EventFrequency * ExposureDuration * EventFrequency * SkinSurfaceArea}{Concentration * DermalAbsorptionFraction * EventFrequency * ExposureDuration * EventFrequency * SkinSurfaceArea}{Concentration * DermalAbsorptionFraction * EventFrequency * ExposureDuration * EventFrequency * SkinSurfaceArea}{Concentration * EventFrequency * EventFrequ$

BodyWeight * AveragingTime

Exposure Dose = $6.2 \times 10^{-6} \text{ mg/kg/day}$

Theoretical Cancer Risk = Exposure Dose x Cancer Slope Factor

Theoretical Cancer Risk = 1×10^{-5}

<u>2. AIR</u>

<u>Child</u>

•
Maximum Concentration of PCBs:
Inhalation Rate:
Exposure Frequency:
Exposure Duration:
Body Weight:
Averaging Time:
Cancer Slope Factor:

0.0000117 mg/m³ 10 m³/day 180 days/year 6 years 35 kg 25,550 days 2 mg/kg/day⁻¹

ExposureDose = $\frac{Concentration * InhalationRate * ExposureFrequency * ExposureDuration}{BodyWeight * AveragingTime}$

Exposure Dose = $1.4 \times 10^{-7} \text{ mg/kg/day}$

Theoretical Cancer Risk = Exposure Dose x Cancer Slope Factor

Theoretical Cancer Risk = 3×10^{-7}

Adult

Maximum Concentration of PCBs: Inhalation Rate: 0.0000117 mg/m³ 15.3 m³/day Exposure Frequency: Exposure Duration: Body Weight: Averaging Time: Cancer Slope Factor: 180 days/year 30 years 70 kg 25,550 days 2 mg/kg/day⁻¹

 $ExposureDose = \frac{Concentration * InhalationRate * ExposureFrequency * ExposureDuration}{BodyWeight * AveragingTime}$

Exposure Dose = $5.4 \times 10^{-7} \text{ mg/kg/day}$

Theoretical Cancer Risk = Exposure Dose x Cancer Slope Factor

Theoretical Cancer Risk = 1×10^{-6}

3. CARPET SURFACE DUST

Child

Maximum Concentration of PCBs:		0.526 mg/kg
Soil Ingestion:		200 mg/day
Conversion Factor:		0.000001 kg/mg
Exposure Frequency:		180 days/year
Exposure Duration:		6 years
Body Weight:	35 kg	
Averaging Time:	· ·	25,550 days
Cancer Slope Factor:		2 mg/kg/day ⁻¹

 $ExposureDose = \frac{Concentration * IngestionRate * ExposureFrequency * ExposureDuration * ConversionFactor}{ConversionFactor}$

BodyWeight * AveragingTime

Exposure Dose = $1.3 \times 10^{-7} \text{ mg/kg/day}$

Theoretical Cancer Risk = Exposure Dose x Cancer Slope Factor

Theoretical Cancer Risk = 2×10^{-7}

<u>Adult</u>

Maximum Concentration of PCBs:0.526 mg/kgSoil Ingestion:100 mg/dayConversion Factor:0.000001 kg/mgExposure Frequency:180 days/yearExposure Duration:30 yearsBody Weight:70 kgAveraging Time:25,550 days

25,550 days 2 mg/kg/day^{-1}

 $ExposureDose = \frac{Concentration * IngestionRate * ExposureFrequency * ExposureDuration * ConversionFactor}{BodyWeight * AveragingTime}$

Exposure Dose = $1.6 \times 10^{-7} \text{ mg/kg/day}$

Theoretical Cancer Risk = Exposure Dose x Cancer Slope Factor

Theoretical Cancer Risk = 3×10^{-7}

4. VACUUM BAGS

Cancer Slope Factor:

<u>Child</u>

Maximum Concentration of PCBs: Soil Ingestion: Conversion Factor: Exposure Frequency: 1.29 mg/kg 200 mg/day 0.000001 kg/mg 180 days/year **Exposure Duration:** 6 years Body Weight: Averaging Time: Cancer Slope Factor:

35 kg 25,550 days 2 mg/kg/day⁻¹

 $ExposureDose = \frac{Concentration * IngestionRate * ExposureFrequency * ExposureDuration * ConversionFactor$ *BodyWeight* * *AveragingTime*

Exposure Dose = $3.1 \times 10^{-7} \text{ mg/kg/day}$

Theoretical Cancer Risk = Exposure Dose x Cancer Slope Factor

Theoretical Cancer Risk = 6×10^{-7}

Adult

Maximum Concentration of PCBs:	1.29 mg/k
Soil Ingestion:	100 mg/da
Conversion Factor:	0.000001
Exposure Frequency:	180 days/y
Exposure Duration:	30 years
Body Weight:	70 kg
Averaging Time:	25,550 day
Cancer Slope Factor:	2 mg/kg/d

g ay kg/mg 'year ays day⁻¹

Concentration * IngestionRate * ExposureFrequency * ExposureDuration * ConversionFactor ExposureDose = BodyWeight * AveragingTime

Exposure Dose = $3.9 \times 10^{-7} \text{ mg/kg/day}$

Theoretical Cancer Risk = Exposure Dose x Cancer Slope Factor

Theoretical Cancer Risk = 8×10^{-7}
APPENDIX F: CDC LETTER FOR SERUM PCB TESTING

Centers for Disease Control and Prevention (CDC) Atlanta GA 30341-3724

October 24, 2006

Suzanne K. Condon, Associate Commissioner Director, Center for Environmental Health Massachusetts Department of Public Health 250 Washington Street, 7th Floor Boston, MA 02108

Dear Ms. Condon:

As you know, the U.S. Centers for Disease Control and Prevention (CDC) has been providing analytical support to the Massachusetts Department of Public Health, Center for Environmental Health (MDPH/CEH), on serum PCB analyses for residents of Pittsfield, Massachusetts. In collaboration with your office and the MDPH State Laboratory Institute, we developed a protocol for the collection of blood samples and analysis for participants who asked to have their blood tested for the presence of PCBs, particularly those children, parents, and staff of the Allendale Elementary School in Pittsfield.

CDC has now completed analyses of the Pittsfield blood samples for PCBs. The analyses were done on a total of 32 samples using congener-specific analytical methods. Results were provided for 35 specific congeners, as well as for the total sum of congeners (lipid adjusted) and the total sum of PCBs on a whole weight basis, the latter of which is most comparable to values previously reported in the scientific literature based on Aroclor analytical techniques. We compared the results for Pittsfield participants with the Third National Report on Human Exposure to Environmental Chemicals, published in 2005. These data represent a random sample of the non-institutionalized U.S. population participating in the 2001-2002 National Health and Nutrition Examination Survey (NHANES). These data are the most recent national data available for comparison and include congener-specific results for U.S. residents aged 12 years and older.

We specifically compared the Pittsfield participants aged 8-19 years of age to the Third National Report data for children aged 12-19 years (the youngest age group collected for NHANES). The Pittsfield participants aged 20 or older were compared to that same age group from the Third National Report.

Results of the analyses on the Pittsfield participants revealed that the Pittsfield participants showed low PCB levels on either a whole weight basis or on a lipidadjusted congener basis as compared with the Third National Report data (available at www.cdc.gov/nchs/about/major/nhanes/guidelines.htm). For example, the seven children who were current Allendale School students (aged 8-10 years) all had serum PCB concentrations below the 50th percentile concentrations for NHANES, based on children aged 12-19 (e.g., median sum PCB concentration of 0.117 ppb [whole weight, 15 congeners] versus 50th percentile concentration in NHANES of 0.345 ppb). Among adult participants, the median serum PCB levels were less than the NHANES 50th percentile values, and all adults were similar to or less than the NHANES 90th percentile values for the U.S. population. In addition, we qualitatively compared congener patterns for PCBs-118, 138/158, 153, 170, 180. and 187 (the most prevalent congeners typically seen in US background samples and in Pittsfield participants) from the Pittsfield participants to see whether any patterns in the participants may have been different from patterns CDC typically sees in the U.S. population. This review did not reveal any unusual patterns among the Pittsfield participants to suggest that exposures that may have led to any evidence of PCBs in blood samples are different than the U.S. population.

We look forward to continued collaboration with the MDPH/CEH on future projects. If you have any questions, please feel free to contact us.

Sincerely,

Yand A. Centern h.

Donald G. Patterson Jr. Supervisory research Chemist Division of Laboratory sciences National center for environmental Health

APPENDIX G: COMMENTS ON DRAFT REPORT - An Evaluation of PCB Testing Conducted at the Allendale Elementary School

COMMENTS ON DRAFT REPORT – An Evaluation of PCB Testing Conducted at the Allendale Elementary School

Introduction

The Allendale School report was released as a public comment draft on October 25, 2006 and a 30-day comment period was established. Upon request this comment period was extended until December 20, 2006. The following are comments received by the Massachusetts Department of Environmental Protection (MDEP), Housatonic River Area Advisory Committee (HRAAC), Allendale School Task Force, City of Pittsfield and responses from MDPH/BEH. Comments related to typographical errors or other minor clarifications are not listed but resulted in corrections or additions to the final report. Additionally, comments were received on appendices attached to the draft report that were final documents and therefore not subject to further comment. These documents (Protocol for Indoor Environmental Testing Protocol and Summary Protocol for Testing Serum PCBs) had already undergone review by members of the Allendale Indoor PCB Environmental Sampling Workgroup and the Health and Medical Peer Review Team.

General Comments

1. Comment:

When reporting total polychlorinated biphenyl (PCB) concentrations (as either total Aroclors or total congeners), all laboratories sum only the detectable concentrations (i.e., non-detects are considered to have concentrations of 0). In discussing the data provided by the various labs, DPH interchanges the terms "detected" and "reported" and uses both terms to represent both data as reported by the labs (as described above) and calculated total Aroclor or congener concentrations that DPH has made using an extremely conservative summing procedure that treats all non-detects as values constituting one-half the reported detection limit. MassDEP has several comments concerning this approach that apply to a number of sections of text in the Report.

- The term "detected" should be used in the Report solely to describe detected concentrations of PCBs.
- The term "reported" should only be used in the Report to describe data as it was reported by the labs and should not be used to describe total PCB concentrations that have been calculated by DPH using a summation

procedure that incorporates one-half the detection limit for all nondetects.

- Unless DPH routinely performs a very conservative calculation for total PCB concentrations that incorporates one-half the detection limits for all non-detected Aroclors or congeners, MassDEP recommends just discussing total PCB concentrations excluding all non-detects from the summation procedure. (Note: MassDEP does add in one-half the detection limit for all non-detects when determining very conservative estimates of exposure point concentrations (i.e., average concentrations) for risk assessment purposes, but not for representing concentrations at individual sampling locations.) MassDEP's proposed approach for depicting total PCBs would allow for more meaningful comparisons with MassDEP's data or with data from EPA or other sources who do not use DPH's convention of adding in non-detects. It will also lead to less confusion in the text of the Report and make the text more focused on the essential aspects and significance of the data (i.e., that the concentrations of PCBs present in environmental media are not at levels that pose health concerns).
- If DPH considers it important to demonstrate that even when using onehalf the detection limit for all non-detects, the maximum total PCB concentrations in all environmental media would still be very low and well below and existing standards or guidelines, then MassDEP recommends emphasizing the significance of this result (as being excessively conservative) in the text of the Report. In addition to including more definitive language to that effect, it would be helpful to list the total PCB concentrations as reported by the labs first and then list the total PCB concentrations as conservatively recalculated by DPH second. In many sections of the text, DPH's recalculated concentrations are listed first and the more conventional PCB totals are listed later in the text.

Response:

As is discussed in the Indoor PCB Environmental Testing Protocol in Appendix A, and in more detail in our eight public health assessments for the General Electric site in Pittsfield, to be conservative, when evaluating environmental data, MDPH treats non-detected analytes as being present at one-half the detection limit, consistent with the protocol. It is important to note, however, that MDPH presented and discussed total PCB concentrations both by assuming NDs were 0 and by assuming NDs were one-half the detection limit, consistent with the protocol. MDPH believes the report was clear and comprehensive in discussing total PCBs using either summary method. The final report has been edited to ensure that "reported" and "detected" are used consistently throughout the document, and that it is clear when PCB concentrations discussed are those reported by the laboratories (ND = 0) or those calculated by MDPH (ND =one-half detection limit).

2. **Comment:** Several different environmental testing guidelines are used throughout the assessment. For clarity and ease of interpretation, we recommend that a chart similar in format to the one below be included in the document.

Summary of PCB Environmental Testing Guidelines used by the Massachusetts Department of Public Health for this Assessment

Agency	Soil	Surface wipe	Air
ATSDR	CREG (lifetime daily exposure for cancer health effects) = 0.4 mg/kg Chronic EMEG (exposures lasting one year or longer for noncancer health effects) =1 mg/kg for children; 10 mg/kg for adults		CREG = 0.01 ug/m3 = 10 nanograms/m3
MDEP	2 mg/kg		
EPA		10 ug/100cm2	
California		0.1 ug/100cm2	

Response: A table has been added to the report similar to the above (see Table 3a).

3. **Comment:** The document contains two very distinct types of testing, the environmental testing and the serum testing. The environmental testing was done to resolve differences posed by conflicting results in earlier school environmental testing, and methodologies were established with that specific purpose in mind. In contrast, serum testing was offered as a service to those parents, students, and staff who voiced concern that their serum levels may be elevated; and results were never intended to be used to scientifically assess whether students, staff, or near-by residents were unduly exposed to PCBs secondary to proximity to the OPCAs. With that in mind, we strongly urge the MDPH to refrain from linking the two types of testing; to keep discussion and conclusions separate; and, to specifically discuss the limitations of drawing any definitive conclusions from the limited number of PCB serum testing results obtained.

Response:

The draft report evaluated and discussed separately the results from indoor environmental testing and PCB serum testing. We have added clarifying language to indicate that the serum PCB testing was a public service offer and hence not designed to collect a statistically representative sample of the community (page 36). However, the environmental data would suggest that elevated serum PCB levels would not be likely as a result of attending or working at the Allendale School.

We also believe that the serum PCB results provide useful information that indicates that for the individuals who were tested, there did not appear to be unusual opportunities for PCB exposures, regardless of potential sources of PCB exposure. This finding was clearly stated in the first sentence of the conclusions: "Results from indoor environmental and serum PCB testing at the Allendale School did not appear to reveal unusual opportunities for PCB exposure to the Allendale School community or to other participants in the serum PCB testing." In addition, CDC's conclusions were similarly stated and were added to the conclusions section of the report (pages 35 and 36). We believe results of indoor environmental and serum PCB testing should provide some level of reassurance to the Allendale School community. Further, MDPH has published a separate report on PCB exposure prevalence in the HRA. That 1997 study concluded that serum PCB levels in Housatonic River area residents have not shown unusual patterns compared to the U.S. population. Serum PCB testing results among HRA residents indicate that increasing age, occupational exposures, and fish consumption are the most important predictors of serum PCB levels, and these predictors are consistent with the scientific literature.

4. Comment:

Discussion and conclusions regarding the environmental testing should include hypotheses as to why some samples, particularly those reflecting areas not cleaned on a regular basis, tested above California safety guidelines.

Response:

As is discussed in the report, one of 98 wipe samples (a sample collected from a windowsill in the gymnasium located 10 feet above the floor) slightly exceeded the California-recommended cleanup standard for PCBs on surface areas. When the data are evaluated assuming all non-detected congeners are present at one-half the detection limit, two wipe samples, one from the gymnasium windowsill and one from a ceiling vent in the health office, had PCB concentration that slightly exceeded the Californiarecommended cleanup standard. As is noted in the report, MDPH hypothesized that since both locations appeared to be visibly dirty it seemed likely that they have not been cleaned in some time. The recommendation that more aggressive cleaning of surfaces not routinely cleaned, such as the gymnasium windowsill, be conducted regularly was based on that hypothesis

5. Comment:

Those who sought out PCB serum testing were a very small, self-selected, non-randomized group of individuals, and wordage throughout the document, especially in its discussion and conclusions, should underscore the fact that results can not be viewed as a representative sample of the Allendale community. Similarly, medians and means calculated from the 32 individual serum tests will also have minimal interpretive value. For example, on p. 29 [page 32 of final report], the document states that "results of the serum testing for Pittsfield children show that participants, especially current Allendale students had low levels when compared with national data." Given the extremely small sample size and the lack of ageappropriate comparisons, we feel that it would be much more accurate to say the following: "Results of serum testing for 7 current Allendale students aged 8-10 showed that they had low PCB levels when compared with NHANES data for children aged 12-17 years." We also feel that it is important to include in your discussion and conclusion that 2 of the 7 former Allendale students aged 12-19 years (28.6%) had PCB serum levels that were higher than the NHANES median.

Response: See response to comment number 3 on clarifying language added regarding the representativeness of the sample. We believe that the report was clear on how the current Allendale student's levels compared to NHANES and other available data in the scientific literature, but we have edited the document, for example, to restate the age categories under discussion (e.g., pages 29,

32). MDPH disagrees with the comment that the serum PCB results have "minimal interpretive value." We believe that the results do not indicate unusual exposure opportunities to PCBs among the participants compared to the general U.S. population. Further, as mentioned previously MDPH's 1997 PCB exposure prevalence study demonstrated that serum PCB levels in Housatonic River area residents have not shown unusual patterns compared to the U.S. population.

Finally, with respect to the comment that two (of five) former Allendale students in the 12-19 age group had serum PCB levels above the NHANES median for this group, MDPH emphasizes that all former Allendale students had serum PCB levels consistent with national data from NHANES. As with any population, there is always variation around a median, with some individuals above and some individuals below the median value. As noted in the report, CDC concluded that there were no unusual patterns among the Pittsfield participants to suggest that exposures that may have led to any evidence of PCBs in blood samples are different than the U.S. population.

- 6. **Comment:** HRI believes it is the right of Allendale school children, the staff, and surrounding neighbors to not be exposed to PCBs and the other chemicals associated with the TEN General Electric Tier one hazardous waste sites. Assurances from the agencies that these families will grow up with no health effects are at best speculation.
 - **Response:** This comment is noted; however, no revisions are warranted. This comment will be shared with agencies charged with environmental remediation.
- 7. **Comment:** There is interest in longitudinal studies of the health of the "children" who attended Allendale School over the years these children are now young adults and older who feel they were at risk when they attended Allendale School before the GE cleanup of the yard and school building.

Response:

MDPH has been conducting serum PCB testing in the Housatonic River Area for more than a decade. Results of serum PCB testing has consistently indicated that serum PCB levels in Housatonic River area residents have not shown unusual patterns compared to the U.S. population. Serum PCB testing results among HRA residents indicate that increasing age, occupational exposures, and fish consumption are the most important predictors of serum PCB levels, and these predictors are consistent with the scientific literature. MDPH continues to offer, as a public service, serum PCB testing to any Housatonic River area resident who wishes to have such testing.

It is important to note that MDPH did seek and receive funding in 2001 from the U.S. Centers for Disease Control and Prevention (CDC) under its national Environmental Public Health Tracking initiative to evaluate possible links between developmental disabilities and PCB exposure in children of Berkshire County. MDPH planned on using readily available Massachusetts Department of Education (MDOE) data on children enrolled in special education programs, as well as data from the MDPH Early Intervention Program, to explore possible unusual patterns of developmental disabilities among Berkshire County children versus the state as a whole in relation to exposure opportunities to PCBs. However, the project was significantly impacted by an interpretation at the federal level of an existing statute (FERPA, or Family Education Rights and Privacy Act) that prohibited MDOE or the MDPH's Early Intervention program from sharing their data with MDPH for public health research. Hence, MDPH was unable to conduct the work as originally planned. Active parental consent was sought to try to overcome this barrier, but low participation limited analyses that could be performed. The results of this effort are available at www.mass.gov/dph/ceh.

8. Comment:

Why was there poor attendance at the DPH meetings at Allendale School? The dates of the various DPH meetings were somehow not well

communicated to the public. There was relatively poor attendance at the meetings at Allendale School. This problem needs to be addressed before future public meetings of this kind. I am not sure if there was apathy or lack of awareness of the meetings.

MDPH cannot answer the question about why attendance at meetings may **Response:** have been lower than expected. However, the Department did coordinate with the Allendale School principal to send two letters out in April 2006 notifying the Allendale community of its offer to conduct serum PCB testing and how to participate. MDPH also coordinated with the principal to send out letters to the Allendale community notifying them of its May 2, 2006 public meeting to address questions and concern about the serum PCB testing effort. There was also considerable press coverage in the Berkshire Eagle throughout this time period, including coverage of MDPH's offer to conduct serum PCB testing for any individual who wanted it. Upon completion of serum PCB testing and environmental testing at the Allendale School, a letter was sent to parents, faculty, and staff to notify them of the October 25, 2006, public meeting to share results of testing. For this latter meeting, a meeting announcement was sent to the Berkshire Eagle.

- 9. **Comment:** After the clean-up of the Allendale schoolyard the EPA told the public the school was cleaned from top to bottom for PCBs. The PCBs have to be coming from somewhere.
 - **Response:** As the report noted, of the 98 environmental samples collected and analyzed for PCB Aroclors and congeners, one sample slightly exceeded any regulatory or screening guideline, in this case a California recommended clean-up guideline of 0.1 ug/wipe. This was a surface wipe sample that was collected from a gymnasium windowsill located 10 feet above the floor. MDPH recommended that cleaning of surfaces not routinely cleaned (e.g. windowsills) be undertaken and regularly conducted as part of routine building maintenance.

10. **Comment:** HRI believes the two OPCAs violate Mass DEP regulations: 310 CMR 16.40(3)(15). Prohibits "any area of waste deposition [that] would be within 1000 feet of an occupied residential dwelling, health care facility, prison, elementary school, middle school or high school or children's pre-school, licensed day care center, senior center or youth center, excluding equipment storage or maintenance structures;....

We believe that any state agency should not be in the business of trying to assure a community that two high level PCB dumps within yards of the school and neighborhood properties has no risks.

Response: This comment is referred to MDEP.

11. Comment:

Many of the Allendale school children live in the same neighborhood which is in proximity to the two OPCAs and several of the Tier 1 GE hazardous waste sites on the GE facility. Of the 300 plus homes tested for contaminated fill in Pittsfield, over 175 had to be cleaned up. The majority of Pittsfield homes have never been tested. Tests of ambient air inside homes around the GE facility have shown low level PCBs in the air. The multiple pathways of exposure would indicate that the children have to be exposed to PCBs in many ways.

Response:

This comment does not refer to the Allendale School indoor environmental testing and hence, no revisions are warranted. Comments related to other environmental data in Pittsfield are beyond the scope of the Allendale School report. However, it is important to note that MDPH has also been conducting public health assessments for the General Electric sites (available at <u>www.mass.gov/dph/ceh</u>), which evaluate available environmental data for these sites and surrounding areas.

Specific Comments

Introduction

12. Comment:

On Page 2, the text in the top paragraph states that MassDEP was a member of the indoor environmental testing workgroup. Although iterated in the Q & A sheet that was handed out at the public meeting on October 25, 2006, this statement is a misrepresentation and MassDEP respectively requests that all such references be removed from this part of the text and from the protocol included in Appendix A of the Report. This comment was previously made to DPH when MassDEP commented on the draft protocol that went out for public comment in May 2006. DPH's response to comments on the draft protocol (included in Appendix A of the Report) stated in response to comment 1 that MassDEP received materials and participated in regular conference calls which gave it the opportunity to provide input into the development of the protocol. This and other statements made in DPH's response to comments are incorrect. The text on Page 2 states that meetings of the workgroup were held between January and May. MassDEP was not requested to become a member of the workgroup until after the first meeting had been held. The second meeting was held at a date and time when MassDEP could not participate. After having missed two of the meetings, MassDEP informed DPH that it would not be participating as a workgroup member. MassDEP did participate in weekly calls between representatives of DPH, EPA and MassDEP beginning in April, but no workgroup members other than DPH staff participated in these calls and, by then, the protocol had already been formulated without MassDEP's input (i.e., the protocol was not provided to MassDEP until it went out for public comment). Furthermore, these calls constituted progress reports on sampling and project status, and occurred after the draft protocol had gone out for public comment and the sampling had been initiated.

Response:

MassDEP will not be mentioned as a workgroup member in the final

Allendale report.

Indoor Environmental Testing: Methods

13. Comment:

In the first paragraph on Page 6, and on Page 7 of Appendix A, the text states that Spectrum Analytical, Inc. (SAI) analyzed vacuum cleaner bag dust samples for Aroclors using the United States Environmental Protection Agency's (EPA's) Method TO-4A and that the Southwest Research Institute (SWRI) used a Method TO-4A to analyze vacuum cleaner bag dust for both Aroclors and congeners. Appendix A also specifies that a modified Method TO-4A was used by SWRI to analyze carpet surface dust, surface wipe and unit ventilator samples for both Aroclors and congeners. Considering that suitable EPA SW-846 methods exist for analyzing Aroclors (Method 8082) in soils and surfaces wipes and congeners (Methods 1668A) in soils and that Method TO-4A is specific to air, it is unclear why Method TO-4A was selected for analysis of these other media. It would be helpful if a rationale and justification were provided in the text for using Method TO-4A for non-air samples and if both the modified Method TO-4A used by SWRI and the lab-specific, notuniversally-accepted method used by the State University of New York at Albany (SUNY) were included as attachments to the Report for public review. The use of different analytical methods by different labs for the same media and constituents (i.e., Aroclors and congeners) is problematic relative to making comparisons between the analytical data produced by

each lab and in making health-based conclusions about PCB levels in the various media that were sampled at the Allendale School. In addition, the text later treats dust as soil for purposes of comparing concentrations of dust found at the school with soil-based standards and guidelines. If dust is to be treated by DPH as a soil for purposes of making these types of comparisons, then it would have been more appropriate to analyze these other media using methods that are specific to soils, rather than air.

Response: SAI analyzed the carpet dust sample using USEPA Method TO-4A and the vacuum bag samples using USEPA Method 8082. This is corrected in the final report.

SWRI used a modified Method TO-4A to analyze samples for congeners and USEPA Method 8082 for Aroclor analysis. This will be noted in the final report. SWRI provided an additional, technical description of the modified Method TO-4A analysis. This additional description was added to the report (page 6).

It is also important to note that the U.S. Environmental Protection Agency Office of Environmental Measurement and Evaluation, Quality Assurance Unit in Chelmsford, MA, provided technical assistance by commenting on the SOPs for all three labs prior to sample collection and analysis.

14. Comment:

There has been considerable public concern expressed at past public meetings at the school concerning the perceived merits of performing congener analysis versus Aroclor analysis, with the misunderstanding being that congener analysis is able to pick up more congeners and yield higher total PCB concentrations that can be obtained by analyzing for a number of Aroclors. In reality, each Aroclor contains mixtures of a number of different congeners and when the samples are analyzed for the normal spectrum of Aroclors typical of those in the SW-846 Methods, these analyses will pick up on most, if not all, of the same congeners as can be detected using the congener-specific analyses. The text on page 6, paragraph 1 downgrades the significance of the Aroclor analyses by stating that these analyses targeted seven (7) Aroclors whereas the congener analysis targets 101 (of the 209) possible congeners. Later in the text, the protocol for indoor environmental sampling (page 8) lists the individual congeners that can be detected by the congener-specific analyses and the text concerning blood serum sampling states that serum samples were analyzed for 36 specific

congeners and these congeners are later listed in Table 5. MassDEP believes that it is important that the report explain to the public the similarity between the numbers and types of congeners that can be detected by the congener-specific and Aroclor-specific methods and emphasize that the analyses performed by all of the labs yielded similar concentrations for PCBs in the various environmental media for both types of analyses. A page on the EPA website (located at the following web address:

http://www.epa.gov/toxteam/pcbid/aroclor_comp.htm) contains useful information concerning the distribution and percentages of the congeners contained in each Aroclor and MassDEP recommends including this reference within the text of the report as well as some text that summarizes the numbers and types of congeners contained in each Aroclor.

Response:

On page 6, MDPH added more explanation on aroclor and congener analyses to address this comment. MDPH also added a sentence referring the reader to the EPA website,

www.epa.gov/toxteam/pcbid/aroclor_comp.htm.

15. **Comment:** In paragraph 1 on Page 6 of the main text, the text briefly states that both SUNY and SWRI agreed to analyze a sample from the National Institute of Standards and Technology (NIST) that contained known quantities of a number of congeners. The text states that the purpose of this procedure "was to determine the comparability of the SWRI and SUNY analyses and how closely their results matched the known quantities of congeners in the NIST sample." MassDEP considers the primary importance of performing this quality assurance/quality control (QA/QC) procedure is to evaluate each lab's ability to perform accurate analyses. Generally, when labs cannot achieve adequate recoveries of contaminants, the operating procedures and the data obtained are considered suspect. The importance of the results of this QA/QC exercise should be emphasized and expanded upon in the text.

Response: MDPH has added text to the Results and Discussion section about the NIST sample results (pg. 14, pg 23).

16. Comment: On page 6 last paragraph [page 7 of final report] – Given that many of the non-cancer effects of PCBs have only recently been appreciated, and to our knowledge, no known safe level of PCBs has been determined for neurodevelopmental and endocrine disrupting health effects, the task force requests that the health effects to which the chronic EMEGs refer be specified. We also request that the peer-reviewed research used to determine the chronic EMEGs for these health effects be made available in your references. A request for this information was also made at the public meeting in October, 2006.

Response:

Environmental media evaluation guides (EMEGs) are values for different environmental media (e.g., air, water) that have been published by the U.S. Agency for Toxic Substances and Disease Registry (ATSDR). EMEGs correspond to concentrations in the specified medium below which adverse health effects are not expected. They are based on ATSDR-derived minimal risk levels (MRLs) and conservative assumptions about exposure and body weight. MRLs are an estimated daily human exposure to a substance that is unlikely to cause non-cancer health effects over a specified period of time. The ATSDR MRL derivations are contained in peer-reviewed toxicological profiles that are comprehensive reviews of the published and unpublished scientific literature for the chemical under review. The specific immunological health effects have been added to the report (page 8) and referenced.

17. Comment:

The last paragraph on Page 7 [page 8 of final report] of the main text discusses comparing carpet surface dust samples and vacuum cleaner bag dust samples with standards for soils. MassDEP does not agree that such comparisons are appropriate, particularly when it comes to comparing these dust samples with standards. MassDEP believes that this comparison is inappropriate because its risk-based Method-1 Standards are based on a series of specific assumptions relative to the properties of soil which affect dermal adhesion, inhalation and ingestion and are expected to be very dissimilar to those of dust, partly since dust is expected to be a more heterogeneous medium.

MDEP provided similar comments on two other sections:

In the top paragraph on Page 19 [page 21 of final report], DPH compares PCB concentrations in dust to health-based standards for soil. For the reasons already discussed in these comments, MassDEP cautions against making such comparisons and drawing any associated conclusions about health risks, unless DPH emphasizes that the concentrations in dust are very low and the comparison to the MCP standard is an overly conservative one.

And:

MassDEP believes that the concluding language in the second paragraph on Page 19 [page 21 of final report] should have greater emphasis than "opportunities for exposures to PCBs would not be expected to result in health concerns." Since Appendix E indicates that the risk calculations that were done for dust included only ingestion and not dermal absorption, there is no concern that soil adherence assumptions were improperly used. However, the fact that dust would most likely not be ingested as readily, as frequently, or in large quantities as soil is still of concern. Because of this, MassDEP believes that any risk estimates made using soil-based assumptions for dust will necessarily be overly conservative, and the nature of the conservativeness of these estimates should be emphasized in the text of the Report.

Response:

The sampling protocol as well as the text of the report clearly notes that there are no available comparison values for dust and hence, to be conservative, MDPH used comparison values for soil as an initial screening to assess dust concentrations in carpet surface and vacuum cleaner bag dust samples. MDPH believes this is an appropriate screening method that the protocol in Appendix A also noted would be used. We have added clarifying language on this point on page 8.

Indoor Environmental Testing: Results

18. Comment:

The third paragraph on page 10 [page 11 of final report] discusses one of two air samples taken from classroom 28 and one outdoor air sample that was collected for comparison purposes. However, Table 3 [Table 3b of final report] indicates that two outdoor samples were collected, so the reference to one outdoor air sample needs to be clarified. In addition, if both samples collected from classroom 28 will not be discussed in the text, then the text should emphasize that only the sample having the highest concentration will be discussed.

Response: The first paragraph of the section discussing air sample results states that two indoor and one outdoor samples were collected on two different days for a total of six samples. Similar to the other sample results sections, only the samples that detected PCBs are discussed; hence, no revisions were warranted based on this comment.

19. Comment: In the fourth paragraph on Page 10 [page 11 of final report], the text describes the range of concentrations in the four (4) SUNY blanks. However, because the blanks were run to perform two different and distinct QA/QC tests, it may be more applicable to separately discuss the range of concentrations in both sets of blanks.

Response: The report text has been edited to present results for the field blanks and the laboratory method blanks separately (page 12).

20. Comment:

In the last paragraph on Page 10 [page 11 of final report], the text states that PCB concentrations reported by SUNY "may be overestimates of what was actually in the indoor or outdoor air," because SUNY also reported detections of PCB congeners for all of the air samples that were part of the OA/OC protocol. MassDEP believes that this sentence should more concisely read, "are overestimates of what was actually in the indoor or outdoor air." In cases where constituents are detected in blanks, analytical methods can be considered questionable and the results should be viewed with certain skepticism. The paragraph goes on to explain that field blanks are used to evaluate "whether improper handling of sampling equipment in the field or during shipment may result in contamination that may not originate from the environmental media being sampled.' In this paragraph, DPH also acknowledges that SUNY's detection of PCB congeners in the two matrix blanks that consisted of new filters sent directly from the ECS office to SWRI, may indicate that these clean and unused sampling cartridges themselves were contaminated. If only one lab had analyzed all of these blanks, these conclusions could be considered to be plausible. However, considering that both SAI and SWRI did not detect any PCBs in any of its four blanks that were obtained from the same source and underwent the same field and shipping procedures, this could suggest an alternative plausible explanation that SUNY employed poor QA/QC procedures, such as improper instrument calibration, using poorly cleaned glassware, etc., that could have resulted in detection of field blanks, matrix blanks, and samples, even if these were properly handled in the field and during shipment. The fact that SUNY also obtained relatively low recoveries of all of the congeners present in the NIST standard samples relative to the high recoveries obtained by SWRI for its analysis of the same standard provides additional reason to question some of SUNY's QA/QC procedures. These QA/QC concerns should be discussed in more detail in the text.

Response: MDPH has added details in the report about the SUNY blank detections (page 11-12). Because it is difficult to know for certain the source of the PCBs in the blanks, MDPH feels the blank results are appropriately described.

21. Comment:

The text in the last paragraph on page 13 [page 15 of final report] states that an additional evaluation of the filter results from "this room" was performed, but it is unclear if "this room" is classroom 21 or classroom 24. This should be clarified. This additional evaluation involved calculating estimates of PCB concentrations in the filter samples per kg filter weight, in order to compare these concentrations to an ATSDR value for soil. MassDEP cautions against attempts to compare concentrations per filter weight to standards that are based on concentrations in soil per kilogram weight of soil, since the soil comparison values are presumably risk-based and incorporate assumptions about soil adherence to the skin, soil inhalation and soil ingestion.

Response:

MDPH emphasized results of other environmental testing in the rooms where filter samples were taken, as we believe (and documented in the protocol for environmental testing contained in Appendix A) that measurements of dust, air, and wipe samples were more important in terms of opportunities to come into contact with PCBs in this indoor environment and hence assessing health concerns. As an added qualitative evaluation that we believe was appropriately discussed in the report, we estimated concentrations of PCBs in the filter samples to compare with soil comparison values, a highly conservative comparison given that individuals would not likely be exposed to PCBs in the filters themselves. This qualitative comparison indicated that even for filter designed to capture particulates, the estimated concentration of PCBs was less than soil comparison values. This helps put into context the low concentration of PCBs found in the filters.

22. Comment: Many of the samples tested positive for low level PCBs. It is logical to assume that if low level PCBs are accumulating on surfaces in the school, then low level exposure has to be taking place. If PCBs are measured in the air, then the children and staff have to be breathing them in. This cannot be a healthy scenario.

Response: In the Discussion section of the report, MDPH evaluates possible health concerns based upon the concentrations of PCBs detected in the building. The report concludes that the results do not appear to reveal any unusual opportunities for exposure to PCBs.

Indoor Environmental Testing: Discussion

23. Comment:

The last sentence in the first paragraph on Page 15 [page 17 of final report] contains some very important health-related, exposure-related information that merits emphasis in more than one sentence. The risk levels shown in Appendix E are below the levels of concern for either MassDEP or EPA and this fact should be emphasized.

Response: This sentence in the Discussion section on surface wipe sample states: "Under the most conservative exposure scenario available, i.e., if the maximum concentration detected in a wipe sample was readily accessible on surfaces throughout the school on a daily basis, for six years for children or 30 years for adults, opportunities for exposure to PCBs would not be expected to result in health concerns (see Appendix E for calculation)." A similar statement is in the Discussion section on carpet and vacuum bag dust, and MDPH believes that additional emphasis is not warranted.

24. Comment:

In the second paragraph on Page 15 [page 17 of final report], MassDEP believes that there should be emphasis on the fact that the surfaces having detectable, yet very low levels of PCBs in dust are inaccessible. Presumably, the EPA comparison value is based on dermal contact and incidental ingestion, as is the California comparison value (as stated on page 7 [page 8 of final report] of the main text of the Report). As such, it appears inappropriate to compare these guidelines to the values for dust on these inaccessible surfaces or to recommend more aggressive cleaning of the ceiling vent or gymnasium windowsill, since there is no opportunity for contact and, hence, no opportunity for either dermal contact or incidental ingestion. If DPH chooses to keep text concerning these comparisons in the text on Page 15 [page 17 of final report], and elsewhere, the text should emphasize the ultra-conservative nature of making such comparisons. If there is no risk to students or adults in the school (as is clearly indicated by the calculations in Appendix E), then it may prove confusing to the public to read this recommendation that seems to suggest just the opposite. This became very apparent from the comments made at the October 2006 public meeting. Furthermore, if DPH decides to include this text in the final Report, then more justification should be provided for why more aggressive cleaning is necessary if there are no public health risks present.

Response:

MDPH believes that the report appropriately describes the noted locations. The Department also believes that to minimize any potential future exposure opportunities, it is appropriate and makes public health sense to

recommend more aggressive cleaning. No revisions in the report were made based on these comments.

25. **Comment:** Two samples tested above clean-up thresholds established by the California Department of Toxic Substance Control. Even though these samples were in hard to reach places, the school should be cleaned again.

Response: See response to previous comment.

26. Comment: On p. 16 [page 18 of final report]– second paragraph – Berkshire Community College is located at or very close to potential site of transformer fluid contamination and the site should not be used for determination of baseline/background concentrations of outdoor air PCBs in Pittsfield.

Response: MDPH investigated the origin of the Berkshire Community College air monitoring station that was established for background comparison for ambient air monitoring related to the General Electric sites. Based on MDPH review of information contained in numerous General Electric site assessment reports as well as historical information provided by U.S. EPA staff who worked on the project at the time, it appears that the Berkshire Community College air monitoring location was established during a public meeting in Pittsfield in the mid-1990s. According to U.S. EPA, this public meeting was attended by MDEP, the Housatonic River Initiative and others; consideration was given to wind patterns and that there was no known PCB contamination on the west side of Pittsfield. This monitoring station is referenced in the September 1997 document of Ambient Air Monitoring for PCBs (at the GE site) prepared by Berkshire Environmental Consultants, Inc summarizing air monitoring data collection from 1991-1996.

> MDPH is not aware of PCB contamination near the Berkshire Community College air monitoring location. Questions regarding locations of transformer fluid contamination should be directed to the Massachusetts

Department of Environmental Protection (MDEP) Bureau of Waste Site Clean-up.

MDPH has added details on the origin of the ambient air background locations in the report (see page 18).

27. Comment: On Page 16 [page 18 of final report], the third sentence in paragraph 3 states that EPA had set up an ambient air monitoring station at Berkshire Community College. Please note, this statement is incorrect, since the General Electric Company established that air monitoring station as a background monitoring location and performed all air sampling there.

Response: See response to previous comment.

28. **Comment:** Since the health office ceiling vent dust contains PCBs, are those in the health office at risk?

Response:The ceiling vent samples are described in the Discussion section. Under
the most conservative exposure scenario, opportunities for exposure to
PCBs in dust at this level would not be expected to result in health
concerns. To minimize any potential future exposure opportunities,
MDPH recommended more aggressive cleaning of surfaces not routinely
cleaned, such as vents and windowsills.

29. Comment: Since the Allendale School air monitoring data that is listed in Tables 3 [Table 3b of final report] and 4 of the Report seems to indicate that PCB concentrations in air at the school are significantly lower than concentrations in air samples collected in the New Bedford homes, the North Carolina day care centers, the United Kingdom buildings and the Cape Cod homes, MassDEP recommends emphasizing this issue in the text by using stronger language than the existing text which states: "...samples at the Allendale School were within or less than the concentrations..."

Response: MDPH believes the current language is appropriate and hence no changes were made.

30. Comment: On page 19 [page 21 of final report] first paragraph – We take issue with the assumption that maximum daily exposure to children attending the Allendale School would be for only 6 years. Many of the children attending the Allendale School live in residences that are adjacent to the

OPCAs. For this subset of children, cumulative PCB exposure would be potentially higher than those only attending Allendale School for 6 years but living elsewhere.

Response:

The samples collected at the school provide data on exposure opportunities inside the school, and hence, the report evaluated exposure opportunities to PCBs in school given the concentrations reported by the analytical labs. Since the school services children from kindergarten through grade five, it was appropriate to assume a six year exposure period in order to assess health concerns that may be associated with the concentrations found in the indoor environmental testing at the school.

However, MDPH did conduct an additional analysis assuming daily exposure for 30 years to the concentrations found in the school. Results of this evaluation did not change the conclusions.

The paragraph that begins at the bottom of Page 19 [page 22 of final report] 31. Comment: states that carpet dust from a series of North Carolina day care centers was analyzed for 20 PCB congeners, whereas dust from the Allendale School was analyzed for 101 congeners. The data listed on Pages 19 and 20 [page 22 of final report] for North Carolina and the Allendale School show that concentrations in Allendale School dust was at least an order of magnitude lower than that found the North Carolina dust. MassDEP believes it is significant that the Allendale School results are that much lower, considering that the Allendale study evaluated five times as many congeners as did the North Carolina study. It is very possible that the North Carolina results would have been even higher if analyses had evaluated as many congeners as were evaluated at the Allendale School. To properly put the Allendale results in context, it is recommended that the text emphasize this large difference in PCB concentrations more so than state that these "concentrations are within the range found within the North Carolina study."

Response: MDPH believes that the report clearly states that the sampling results for Allendale and North Carolina are summarized for different numbers of congeners.

32. Comment: In the last paragraph on Page 20 [page 23 of final report], DPH iterates its discussion about comparing PCB levels in air filters (on a mg/kg basis) to soil standards. MassDEP has already commented on this above, but iterates

its comment here. It would be preferable if the discussion emphasized that since the air filters are collecting minor amounts of PCBs and there are very low levels of PCBs in indoor air, the air filters are functioning successfully, as designed, to remove all particulate matter from the air. MassDEP also believes that it is important to emphasize here that there are not existing health-based standards for dust in air filters and that indoor air concentrations (for which some standards exist) are a better indicator of potential inhalation exposures in indoor air than are the inaccessible filters.

Response:

MDPH discusses in the Results section (page 14), Discussion section (page 23), and in the Indoor Environmental Sampling Protocol (Appendix A) that there are no regulatory standards or guidance values for air filters and that sampling results from other media in those rooms (e.g., air, dust) are better indicators of exposure opportunities. We believe the report (including Appendix A) adequately emphasizes the other testing results in the rooms where filter samples were taken and the role of filters to capture particulate matter, and hence, no changes in the text are warranted. We also believe the qualitative analysis of PCBs in filters indicates that the filters did not appear to contain unusual levels of PCBs.

33. Comment:

For years HRI has been asking for a peer reviewed, published number based on scientific studies for long term, low level exposure to PCBs for both adults and children. The only number ever presented to the Pittsfield public is an OSHA number based on adult exposure for an 8 hour work day. No one knows the implications of this type of long term, low level exposure.

Response:

MDPH is confused about this comment, as we have conducted numerous public health assessment activities in the Housatonic River area over the past decade, including eight separate public health assessments specifically focused on the General Electric sites, that use ATSDR comparison values. ATSDR has derived comparison values, which are peer reviewed, that are applicable to long term exposure to children or adult, assuming exposure opportunities in residential settings. ATSDR has published a Toxicological Profile on PCBs (2000) that includes a comprehensive assessment of the scientific literature on health effects from PCBs and details the toxicity information used to derive their comparison values. These comparison values were also used for the appropriate media in the Allendale School report. The PHAs can be viewed at <u>www.mass.gov/dph/ceh</u> or individuals can receive a copy of them by calling (617) 624-5757 or (800) 240-4266.

34. **Comment:** Fingerprinting of the Aroclor types might indicate whether the PCBs were the types GE used at the facility.....1254,1260.

Response: The indoor environmental testing conducted by MDPH was not designed to determine the source of any detectable PCBs, but rather to quantify the presence of PCBs and assess potential health concerns.

PCB Serum Testing: Methods

35. Comment:

In paragraph 4 on Page 27 [page 30 of final report], the first sentence states that some of the 18 adults whose blood serum were tested in the Allendale study were not affiliated with the school and live either elsewhere in Pittsfield or in neighboring communities. It is unclear if any of these individuals had a past association with the school or if they were selected as controls (although the latter is unlikely considering the small total sample size). If neither is true, then it is unclear why these individuals were sampled. The rationale for including these individuals in the sampling effort should be explained.

Response:

The MDPH has had an open offer to the Housatonic River Area residents for approximately 10 years to assess individual resident's opportunities for exposure to PCBs and analyze their blood if they so requested. This public service activity was offered following MDPH 1997 exposure prevalence study which concluded that assessment of exposure to PCBs has shown that serum PCB levels among participants with the highest risk of exposure were generally within the background range reported for the nonoccupationally exposed population in the U.S. The Allendale report summarizes results for all individuals who responded most recently to this ongoing offer. The report specified the breakdown of participants who responded to our latest public service offer in terms of whether the individuals were current or former students or staff, parents of students, or nearby residents.

36. Comment: Younger children (age 8-10) were compared to NHANES data for older children.

Response:

se: As described in the Methods section (Sample Analysis) of the report, children under 12 were not sampled for PCB serum analysis in the CDC's 2005 Third National Report on Human Exposure to Environmental Chemicals and therefore no direct comparison data were available. Thus results of the Allendale children less than 12 years old were compared to children ages 12-19 years old from NHANES. We also explained that because of the well-known fact that serum PCB levels rise with increasing age, we would expect the Allendale children younger than 12 years of age would have similar or lower serum PCB levels.

PCB Serum Testing: Results

37. Comment:

Why weren't there more volunteers for blood testing? A few days after Elaine Krueger sent out fliers for volunteers to call and sign up for PCB blood testing - I found out from some Allendale staff that this telephone number did not connect. I emailed Elaine, told her the problem, she followed up with the telephone company who resolved the problem in a few days. When I informed staff that the telephone problem was fixed and they could then call to sign up, many were discouraged, apathetic and/or suspicious of the DPH 's intentions. There are some people who just have a fixed negative opinion about GE and any government agency. Others feared finding out their blood level of PCB's because "I can't do anything about it anyway" since it binds to lipids and there is no treatment for high blood levels of PCB's. I heard comments from some community residents around the school that they would have volunteered for blood testing but this study was only for the Allendale School community.

Response:

As noted, MDPH quickly responded to the alert that the 800 number was not working for those calling from Berkshire County, and resolved the problem the day that it was identified. In addition, MDPH coordinated with Allendale School officials to distribute a letter to all students, parents, faculty, and staff to notify Allendale School community members of the offer to test serum PCB levels. In addition the MDPH/BEH in partnership with the Pittsfield Board of Health, held an informational meeting on May 2, 2006 at the Allendale School to discuss this effort and answer questions.

There was extensive media coverage, e.g., by the Berkshire Eagle, throughout this period that also reported on the serum PCB testing offer. It is also important to re-emphasize here MDPH's continuing offer to administer an exposure assessment questionnaire and blood testing for HRA residents concerned about their individual exposure opportunities to PCBs.

38. **Comment:** Some of the Allendale school adults had higher median PCB blood levels than other city adults.

Response: To address this comment, MDPH further evaluated the serum PCB data for the current Allendale staff. The median serum PCB level for the four Allendale staff (1.618 ppb) was above the NHANES 50th percentile (1.062 ppb) but below the 75th percentile (1.883 ppb). MDPH evaluated the length of employment for each of the four participants by serum PCB level. While there were too few individuals to evaluate any meaningful trends, there are some observations that may be useful in considering any possible association between the school and individual serum PCB levels. That is, the individual with the lowest level worked there for more than a decade, while the individual with the highest level worked there for the least amount of time. If the school was the major source of PCBs, we would expect higher levels in individuals working longest at the school. These additional details were added to the report (page 31).

It is also worth noting here that the CDC concluded that "Results of the analyses in the Pittsfield participants revealed that the Pittsfield participants showed low PCB levels on either a whole weight basis or on a lipid-adjusted congener basis as compared with the third National Report data."

PCB Serum Testing: Discussion

39. Comment:

The data summarized in the second-to-the-last paragraph on Page 30 [page 33 of final report] indicates that the PCB blood serum levels in the Allendale School children was an order of magnitude lower than the PCB levels

detected in the children from Anniston, Alabama. Although this difference in blood serum levels seems to be significant, the text does not elaborate or distinguish that the Allendale levels will necessarily be lower if non-detects are added into the summation using one-half the detection limit divided by the square root of two, whereas total PCB calculations for the Anniston study used one-half the detection limit for all non-detects. MassDEP recommends that DPH use one-half the detection limit for all non-detects for the Allendale data for purposes of making comparison to the Anniston data. In addition, it should be noted that Allendale blood serum concentrations could be slightly higher if the study had analyzed for 37 congeners.

Response:

The CDC method of assuming non-detected congeners were present at the method detection limit divided by the square root of 2 actually results in HIGHER, not lower, total serum PCB concentrations than if one assumed the non-detected congeners were present at one-half the method detection limit. While it is correct that the Anniston serum PCB levels were reported under the assumption that non-detected congeners were present at one-half the detection limit, it is not necessary to re-calculate the Allendale results under the same assumptions, as the Allendale results for children are already lower than the Anniston results. The Allendale serum PCB results were calculated to be consistent with CDC's treatment of non-detected congeners so that comparisons to the national NHANES data could be made. Additional discussion about the comparison between Anniston and Allendale results was added to the report on page 33.

Additionally for clarification, the Pittsfield samples were analyzed for a total of 38 PCB congeners (clarification to the Methods section of the report). As described in the Results section of the report, results for PCB congener 18 were not reported because one or more of the quality assurance/quality control parameters did not meet the specified criteria, which CDC reports is common for congener 18 for all labs. CDC also reported congener 18 is a minor contributor to total serum PCBs. Thus, 37 congeners were reported in the results, including congeners 138,158 and congeners 196,203, which cannot be separated by this methodology and are reported together. Therefore when the sum of total PCBs for 35 congeners are

described in the report, the sum includes 33 individual congeners (excluding congener 18) and 2 pairs of co-congeners (i.e. co-eluting congeners) totaling 37 congeners. See Table 5 of the report for a complete list of congeners. The Anniston Alabama study (Orloff et al. 2003) also analyzed for the same 37 congeners and reported serum levels for congeners 138-158 and 196-203 as co-congener pairs as well. Therefore the assessment of total serum levels is accurately comparing the same congeners.

40. Comment:

On page 31 first paragraph [page 34 of final report] – Similarly, did the questionnaires reflect possible reasons for why one adult had a PCB concentration of 3.595 ppb? Did this person share potential PCB exposure pathways related to living or working in proximity to the OPCAs?

Response:

The individual with a serum PCB level of 3.595 ppb had reported occupational exposures to PCBs, was in the oldest age group of all participants (>50 years old), reported living in a residence near GE, and reported being a fish eater. Thus, the individual did have a number of other factors that would suggest higher serum PCB levels, particularly age and occupational exposures, than individuals without these factors. Again, it is important to note that CDC concluded that the serum PCB results for Pittsfield participants were low and consistent with national data (e.g., all participants had serum PCB levels less than the NHANES 90th percentile). Additional language about this individual's serum PCB result was added to the text on page 34.

Conclusions

41. **Comment:** On page 31 [page 35 of final report] second paragraph – We recommend that the wording of the first sentence be changed to the following: "Although the Allendale School has an unusual opportunity for PCB exposure, results from indoor environmental testing and serum PCB testing from a very limited number of students and adults did not appear to reveal significant PCB exposure".

Response: This comment refers to the sentence in the original report: "Results from indoor environmental and serum PCB testing at the Allendale School did

not appear to reveal unusual opportunities for PCB exposures to the Allendale School community or to other participants in serum PCB testing." The sentence was not intended to imply that the close proximity of Hill 78 to the Allendale School was not atypical, but rather that environmental and blood tests did not result in unusual levels of PCBs in the indoor environment nor unusual body burdens of PCBs. MDPH did add language in the conclusions section (page 35) that we believe addresses this comment.

42. Comment:

The last sentence in the first paragraph on Page 32 [page 35 of final report] compares indoor air levels at the school with historical outdoor air samples collected in past years and arrives at conclusions about anticipated indoor and outdoor air concentrations in the colder and warmer months. Whereas, MassDEP does not disagree with DPH's predictions about slight seasonal variations in outdoor air concentrations, MassDEP does not believe that enough seasonal indoor air data has been collected at the Allendale School to make comparisons with seasonal variations in outdoor air concentrations in outdoor air concentrations and recommends either removing this sentence from the text or writing it in a different manner.

Response: MDPH believes the sentence is clear as written with appropriate qualifications. No change.

43. Comment: The sample size of both environmental and blood testing is extremely limited to draw any conclusions yet the public is led to believe there are no health implications working or living near the two OPCAs.

Response:We believe that the available data, including two rounds of indoor
environmental testing in 2005 and 2006 (a total of 186 samples) and serum
PCB testing do not indicate unusual opportunities for exposure to PCBs at
the Allendale School. MDPH continues to offer serum PCB testing to any
Pittsfield resident who may have concerns about their individual exposure
opportunities to PCBs.

44. **Comment:** The Allendale School Taskforce is relieved that an extremely small sample of current Allendale children aged 8-10 years had serum PCB levels below the CDC 50th percentile for children aged 12-19 years.

However, other findings in this evaluation give us pause and include the following:

1. That two of the seven former Allendale School students (28.6%) had PCB serum levels above the NHANES mean.

2. That current Allendale staff had higher median PCB serum levels compared with other Pittsfield adults tested.

3. That indoor environmental testing of areas not subject to cleaning on a regular basis (but thankfully out of reach from most children) tested above clean-up thresholds established by the California Department of Toxic Substance Control.

We feel that the Allendale Community deserves a discussion about the potential reasons for these observations, along with specific mention that the limited number of serum PCB tests obtained prohibits the drawing of any conclusions regarding potential PCB exposure from the OPCA sites.

We also feel, at an absolute minimum, that this information should be viewed as a lesson learned regarding the wisdom of sitting hazardous waste sites adjacent to schools. Our knowledge of the safety of exposure to low dose PCBs over time is extremely limited, and the finding of even small amounts of PCBs in the Allendale School should prompt both General Electric and public agencies to thoroughly investigate alternative particularly relevant now given the fact that Hill 78, the OPCA closest to Allendale School, is unlined and will require continued monitoring in perpetuity and that future clean-ups of the Housatonic River will pose similar concerns about the safety of PCB consolidation.

We also appreciate the sincere effort put forth by the Massachusetts Department of Public Health, specifically Elaine Krueger and her staff, in trying to resolve some of these complicated and often politically contentious issues; and would like to thank them for listening and responding to our concerns about the proximity of the GE/EPA PCB consolidation sites to the Allendale Elementary School.

Response:

We agree that these findings should be reassuring to the Allendale School community. Much of the language within the above comment has been addressed elsewhere in this document. For example, with respect to the comment about the two former Allendale students, see response to comment number 5; about the current Allendale staff, see response to comment number 38; about the representativeness of the sample, see response to comment number 3; and about the surface wipe samples and cleaning, see response to comments 24, 25, and 28. The statements

relative to the proximity of Hill 78 to the school will be shared with state and federal regulatory agencies.

Finally, the MDPH appreciates the kind words offered in support of our fallen colleague and her staff. Elaine's contributions continue to be missed on a daily basis but her commitment to public service continues to inspire all of us at the Bureau of Environmental Health.

Recommendations

45. **Comment:** Does DPH recommend that the ceiling vents/air ducts be vacuumed out? If so, who would pay for this?

Response: MDPH recommends that areas not routinely cleaned, which may include ceiling vents/air ducts, be routinely cleaned. We believe school maintenance staff can best assess most appropriate techniques for cleaning.